Guide to Laboratory Services
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Purpose

The purpose of this manual is to provide our clients with information about laboratory testing availability and to provide a guide for collecting and submitting specimens.

This edition can also be accessed on the SC DHEC website at:
http://www.scdhec.gov/Health/FHPF/LabCertificationServices/LabServicesGuide/

Mission Statement

The mission of the Public Health Laboratory (PHL) is to provide specialized laboratory testing for accurate screening, diagnosis, prevention and surveillance of disease, foodborne illness, and congenital disorders to improve public health and the quality of life for the South Carolina community.

General Information

Laboratory Certification for Clinical Testing- CLIA ID# 42D0658606

The Public Health Laboratory of the South Carolina Department of Health and Environmental Control (SC DHEC), formerly named the Bureau of Laboratories, is a multi-disciplinary, integrated source of diagnostic services including analytical support and consultation for physicians, private laboratories, hospitals, and county health departments. The PHL is prepared to assist in any national public health emergency.

Physical Address

The Public Health Laboratory is located in the James A. Hayne Building at 8231 Parklane Road, Columbia, SC 29223, on the campus of the State Park Health Center. State Park is located between Highway 555 (Farrow Road), Parklane Road and the I-77 connector (or SC I-277) two miles north of I-20; 2 miles west of Columbia Mall. Using the Parklane Road Entrance, the Hayne Building is at the end of the first left turn.

Hours of Business

The official working hours are from 8:00 A.M. to 4:00 P.M. Monday through Friday.

After Hours, Weekends and Holidays

The laboratory maintains an On-Call Roster for all weekends and holidays. Individuals requesting information or services of an emergency nature can call the main number, 803-896-0800. This number transfers to an answering service who will contact the Director on call.

A list of state holidays for the current year can be found on the SCDHEC website at:

Emergency Response / Disaster Preparedness

As part of DHEC’s Emergency Preparedness Plan of Action, the Public Health Laboratory is equipped and staff are trained to respond rapidly and effectively to a medical emergency, natural disaster, and / or act of bioterrorism or chemical terrorism. If the emergency occurs outside of regular working hours,
personnel will be called back or work overtime as needed to provide laboratory support.

**Specimen Receiving**

With the exception of Newborn Screening, specimens transported by DHEC’s courier service are placed in specially marked boxes and are picked up by lab staff from the Sims-Aycock building between 5:00 AM and 6:00 AM Tuesday through Saturday. Specimens sent by first class mail are picked up by lab staff from the U.S. Post Office at 8:30 AM Monday through Saturday. The U.S. Post Office delivers any overflow packages at approximately 12:30 PM, Monday through Friday.

On DHEC-observed non-federal holidays, specimens are picked up by laboratory staff between 7:00 AM and 8:00 AM from the Sims-Aycock building and the U.S. Post Office. These are sorted and stored according to established protocols to be accessioned on the next working day.

Specimens are accepted at the Hayne Building during the business hours of 8:00 AM to 4:00 PM Monday through Friday, except for state holidays. Private couriers delivering specimens at the back entrance of the Hayne Building should call Specimen Management Section at 803-896-0898 for pick up at the loading dock. Private couriers and/or individuals delivering specimens through the front entrance are assisted by the Security Officer at the front desk.

**After-Hours Delivery of Specimens**

Specimens other than Newborn Screening specimens will not be accepted after hours unless special arrangements have been made with the laboratory section conducting the test. This person will notify the Security Officer on duty that a delivery is expected.

The after-hours depository located in the rear of the Hayne Building is for animal heads being delivered for rabies testing only. Please do not put specimens and cultures in the depository.

Newborn screening specimens can be accepted at the Security Desk of the Hayne Building after business hours. Couriers delivering from hospitals will sign the specimens in on a log kept at the Security Desk. Holiday and Saturday delivery of Newborn Screening specimens shipped using FedEx/UPS are also accepted by the Security Desk.

**Contact Persons and Phone Numbers**

- Laboratory Test Results: (803) 896-0800
- Laboratory Request Forms/Mailing Containers: (803) 896-0913
- Facilities Maintenance (Laboratory Instrument Services): (803) 896-0919
- Laboratory Director: (803) 896-0965
- Assistant Laboratory Director: (803) 896-9725
- Director, Chemistry Division: (803) 896-0991
- Director, Microbiology Division: (803) 896-0870
- Support Division Manager: (803) 896-2331
- Director, Logistic Division: (803) 896-0923
- Office of Quality Assurance: (803) 896-3897
- Office of Laboratory Safety: (803) 896-0956
- Laboratory Information Management Systems (LIMS) Administrator: (803) 896-4777
- Complaints: (803) 896-3897 or (803) 896-0899
Testing Policies

Persons Authorized to Order Tests
The Laboratory will accept clinical laboratory specimens for testing from physicians, health departments, and hospital laboratories, or as provided by South Carolina statues. These senders will be responsible for receiving, relating, interpreting, and/or distributing the data. A clinical laboratory specimen is described as any material derived from the human body for the purpose of diagnosis, prevention, treatment or assessment for medical or legal purposes. Inanimate substances and other samples submitted for examination (e.g., environmental lead samples, etc.) may be accepted from private citizens at the discretion of the Division Director, Laboratory Supervisor, Assistant Laboratory Director, or Laboratory Director.

Verification of Orally Ordered Tests
When additional tests are requested by telephone, the caller is asked to follow up with a written request on letterhead or to send an additional laboratory requisition form for that test to the Public Health Laboratory. Please send written requests to the attention of the Section Supervisor or to the Specimen Management Section. The additional test(s) will not be performed until the written request is received. With time sensitive tests, the specimen(s) may be tested immediately, and the results held until the written request is received. Exception: No HIV tests will be performed without written request at the time of testing. All blood specimens will be discarded if a written request is not received within seven working days.

Requesting Additional Testing on a Serology Specimen
To request an additional serology test, call the Specimen Management Section at (803) 896-0898. Specimens are discarded after seven working days. A request must be made within that time period. Additional testing on the same specimen may not always be feasible. The testing laboratory may request additional information to determine specimen acceptability. In some cases, a second (new) specimen for testing may be recommended. In other cases, the patient’s clinical history may provide an explanation for the initial result, and additional testing or retesting may not be necessary.

Specimens Referred for Testing to the CDC
Laboratories wishing to send specimens directly to the CDC should contact the Microbiology Division at (803) 896-0870. The sender will be assigned a State Health Department Number and will be asked to forward the Public Health Laboratory (PHL) a copy of the information being sent. CDC forms are also available from the PHL.

Other Reference Laboratories
If a specimen is sent to a reference laboratory for initial, follow-up, or verification testing by the Public Health Laboratory, the sender will be notified that the specimen has been referred. Either the original result report from the reference laboratory is forwarded to the sender, or the results will be reported using the PHL’s laboratory information system, noting where the test was performed. A copy of the report is maintained by the laboratory.

STAT Testing
Requests received in the morning will be put in the day’s run. The results will be telephoned to the requestor, followed by a hard copy report or electronic accessible report. If the request is for a test that
will not be performed immediately, the requestor will be informed by telephone when the test will be performed and the result available.

**Confirmatory Testing**
When confirmatory tests are necessary, preliminary results are reported until all testing is completed. Once all testing is complete, a final report will follow.

**Laboratory Specimens Sent to the Public Health Laboratory in Error**
Specimens sent to the laboratory in error will be returned to the sender as soon as possible.

**Correction of Patient Information**
All requested changes to the request form by the sender must be documented on letterhead, dated, and signed by the requestor. A returned copy of the original laboratory report requesting the missing information is also acceptable to communicate changes needed as long as the sender states clearly what is needed, dates, and signs the report. The patient’s record will be updated to reflect the change.
Specimen Rejection & Disclaimer Criteria

The following rejection criteria and disclaimers are considered universal, as they apply to all specimens submitted for testing. Specific test related rejections are listed in the Alphabetical Listing of Test (Section II) and the Collection Procedures (Section III).

No Specimen Received
When a request form is received without a specimen, notification to the client should be made about the specimen and the laboratory’s inability to perform testing. In addition, the laboratory test report will state that no specimen was received and that testing was not performed.

No Requisition Received
If a specimen is received without a request form and the sender cannot be identified from the specimen label, the specimen will be held awaiting telephone inquiry or delayed receipt of requisition form. After seven days, blood specimens are discarded. Gen-Probe Aptima swab specimens are discarded after 60 days and the Gen-Probe Aptima urine specimens are discarded after 30 days.

No Name on Specimen
When a specimen is received without an identifying number or patient name, it WILL NOT be tested. An exception may be made at the discretion of the Supervisor, Division Director, or Laboratory Director for a specimen that cannot be recollected because of its unique anatomic source, collection method, or time of collection. Examples include: CSF, peritoneal pleural and synovial fluids, autopsy, biopsy, or organ specimens, and specimens collected prior to the initiation of antimicrobial therapy.

No Name on Requisition
When a requisition form is received without a name, and there is no other identification on the form that matches the information on the specimen, a call will be placed to the submitter requesting a corrected requisition. The specimen will be tested ONLY after the corrected lab requisition form is received.

No Test Requested
When a specimen is received and there is no test marked on the lab requisition, the sender should be notified by telephone, and the following documentation will be entered into the computer system if a corrected requisition is not provided, “No test requested. For this specimen to be tested, write the test name on this form and return it to the lab. This specimen will be discarded 7 days after the received date shown.” When a corrected requisition form is received, only then will the specimen be tested.

Note: Blood specimens are discarded after 7 days.

Other Missing Information
If other necessary information is missing, the specimen will be tested, and the missing information will be requested by phone, fax, mail, or email. The result will be held until the missing information is received.

Mismatched Information
When the name on the request form and the specimen do not match, the specimen will NOT be tested. It will be reported as, “Name on specimen differs from name on request form”.

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Partial Information Matches
When there is a partial name match and other data on the request form matches, it is most probably the same patient. The name on the tube is written on the request form, and the test is run, and a disclaimer added to the report.

Verification should be made by contacting the client about the specimen, and the data/time and name of the person spoken with should be documented.

Specimen Broken or Leaked in Transit
When a broken or leaking specimen is received, every attempt will be made to salvage it without compromising the integrity of the specimen.

If the sample CANNOT be salvaged, a notation that the sample broke/leaked in transit should be made in the lab information system (LIS) to be included on the report. The laboratory report should also inform the client that testing was not performed.

Notification should be made to the client about the specimen, and the date/time and name of the person notified should be documented in the computer system.

Incorrect Specimen Received
If the specimen received is incorrect for the test requested, notification should be made to the client about the specimen and laboratory’s inability to perform testing. In addition, the laboratory test report will state that the incorrect specimen type was received and that testing was not performed.

Unsatisfactory Specimens
The Public Health Laboratory will discard specimens which are received in unsatisfactory condition. The reasons for the rejection will be reported to the sender on the standard laboratory report form. Unsatisfactory conditions include, but are not limited to:

- Hemolyzed, chylous, or contaminated specimen,
- Specimen received beyond the acceptable time for testing,
- Specimen collected too soon or too late during the disease-state for the test requested,
- Specimen was stored and shipped at improper temperature,
- Specimen is nonviable, or decomposed,
- Specimen quantity insufficient

Specimens that have some degree of hemolysis, icteric, or chylous, will be tested if the degree of hemolysis or lipemia does not interfere with the analysis. The unsatisfactory condition will be indicated on the report form.
Results Reporting Policies
All laboratory reports generated are considered confidential information. The reports will be released only to authorized persons. Reports can be accessed via the internet through a laboratory web portal, allowing immediate access to results. Reports are mailed daily to clients without access to the laboratory web portal, as requested. Clients can only view information on orders that have been logged in with their customer ID. Contact the laboratory at 803-896-4777 for any issues regarding web portal access.

Telephone Results
Panic/critical values and/or public health emergencies are telephoned to the appropriate person(s). A result will NOT be left on voice mail or an answering machine. A message to call the Public Health Laboratory for results will be left.

Copies of Test Reports
Newborn Screening: Laboratory reports are available via the internet through eReports, a laboratory web portal for the hospital submitting the specimen and for the physician whose name has been entered on the request form as the healthcare provider. An account must be set up by the LIMS office to access reports on eReports. All other tests: Reports can be accessed electronically through the OpenELIS web portal or Citrix. Copies of test reports will be provided to the name entered in the sender section of the request form or to the provider, upon request.

Remailing of Test Reports
If a physician or clinic to which the patient has been referred requests a copy of a test results, the report will be reprinted with the original sender number and sent as requested. If the report is not received, please call 803-896-0800 or 803-896-4777.

Correcting Reporting Errors
If an error or the possibility of an error is discovered by the laboratory after results have been reported, the sender should be notified immediately by telephone. The error should be explained, and the correct result given. A corrected report will be issued with the comment, “Corrected Report”.

If an error in reporting is discovered by the sender, the laboratory should be notified immediately. The error will be corrected, and an updated report will be generated. The corrected report will include the comment, “Corrected Report” if a result has been changed, or “Amended Report” for other error types (e.g., patient demographics).
**Disease Reporting**
The Code of Laws of South Carolina (1976) Section 44-29-10: Regulation 61-20 mandates that the Commissioner of DHEC is to publish annually a list of diseases to be reported by physicians and laboratories. This list can be found on the Internet at [https://scdhec.gov/sites/default/files/Library/CR-009025.pdf](https://scdhec.gov/sites/default/files/Library/CR-009025.pdf).

All communicable disease outbreaks and unusual disease occurrences should be reported, so that appropriate control measures can be implemented.
## SECTION II
### ALPHABETICAL LISTING OF TEST INFORMATION

<table>
<thead>
<tr>
<th>Test</th>
<th>BACILLUS ANTHRACIS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Anthrax</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>72 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical specimens / isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with the patient's first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Clinical specimen / Pure isolate on slant</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 520; “Suspect Agent” = Bacillus anthracis</td>
</tr>
<tr>
<td></td>
<td>DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be pre-approved by Special Pathogens Supervisor prior to submission.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at (803) 896-0777 / (803) 767-8118</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Availability:</td>
<td>24 hours / 7 days a week</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Preliminary results (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Bacillus anthracis is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of Bacillus anthracis detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect B. anthracis in clinical specimens or referred isolates.</td>
</tr>
<tr>
<td>Method:</td>
<td>A variety of sentinel and Laboratory Response Network (LRN) Methods are used to detect Bacillus anthracis.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>BRUCELLA</td>
</tr>
<tr>
<td>-------------------------------------------</td>
<td>--------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Brucellosis</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As Needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>7- 21 days from time of specimen receipt in the laboratory</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical Specimens / isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Call the Special Pathogens Laboratory.</td>
</tr>
<tr>
<td>Form:</td>
<td>Form 1335; Test Number 520; “Suspect Agent” = <em>Brucella</em> sp. DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be pre-approved by Special Pathogens Supervisor prior to testing.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at (803) 896-0777 / (803) 767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Availability:</td>
<td>24 hours / 7 days a week</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Preliminary (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td><em>Brucella</em> abortus, <em>melitensis</em>, and <em>suis</em> are designated as Select Agents (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of <em>Brucella</em> detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect <em>Brucella</em> organisms in clinical specimens / To confirm suspect isolates</td>
</tr>
<tr>
<td>Method:</td>
<td>A variety of sentinel and Laboratory Response Network (LRN) Methods are used to detect and speciate <em>Brucella</em> organisms.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>BRUCELLA ANTIBODY (TOTAL) by AGGLUTINATION</td>
</tr>
<tr>
<td>------</td>
<td>------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Brucella Microagglutination Test (BMAT)</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / (803) 896-0777 or (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday-Thursday</td>
</tr>
<tr>
<td>Turnaround Time</td>
<td>5 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>500 µL</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator Tube (SST).</td>
</tr>
<tr>
<td>Form:</td>
<td>Form 1335; Test Number 522; check “BMAT”. DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be pre-approved by Special Pathogens Supervisor prior to testing.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Serum specimens should be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until receipt at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Hemolysis; lipemia; gross bacterial contamination; improper temperature; For universal rejections, See Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
</tbody>
</table>
| Results and Interpretations: | - A single serum titer of 1:160 or higher is suggestive of exposure to Brucella at some time. Titer results ≥ 1:160 will automatically reflex to a repeat test with the “reduced” serum for acute/convalescence determination. 
- Cross-reactions may occur between Brucella and F. tularensis antigens and antisera 
- Preliminary (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. |
<p>| Additional Information: | N/A |
| Purpose of Test: | To presumptively detect smooth strain brucella antibodies in human sera. This test will not detect exposure to Brucella canis or Brucella abortus RB51 rough strains. |
| Method: | Semi-Quantitative Agglutination |
| Interfering Substances: | Hemolysis; lipemia; gross bacterial contamination |
| Comment: | Please call the Special Pathogens Laboratory with any questions or concerns. |</p>
<table>
<thead>
<tr>
<th>Test</th>
<th>BURKHOLDERIA MALLEI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Glanders</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time</td>
<td>7 to 10 days from the time of specimen receipt in the laboratory.</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical Specimens / isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 520; “Suspect Agent” = B. mallei DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be pre-approved by Special Pathogens Supervisor prior to testing.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at (803) 896-0777 / (803) 767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Preliminary results (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Burkholderia mallei is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of Burkholderia mallei detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect B. mallei in clinical specimens / To confirm referred isolates.</td>
</tr>
<tr>
<td>Method:</td>
<td>A variety of sentinel and LRN methods are used to grow, confirm, or rule-out bacterial isolates.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogen Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td><strong>Test</strong></td>
<td><strong>BURKHOLDERIA PSEUDOMALLEI</strong></td>
</tr>
<tr>
<td>----------</td>
<td>--------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Melioidosis</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time</td>
<td>7 to 10 days from the time of specimen receipt in the laboratory.</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical Specimens and clinical isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
</tbody>
</table>
| Form: | DHEC 1335; Test Number 520; “Suspect Agent” = *Burkholderia pseudomallei*  
DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen. |
| Special Instructions: | Specimen must be pre-approved by Special Pathogens Supervisor prior to testing. |
| Packing and Shipping*: | Special handling criteria apply. Please contact the laboratory for special instructions at (803) 896-0777 / (803) 767-8118. |
| Transport Conditions: | Determined during pre-approval consultation. |
| Specimen Rejection Criteria: | Determined during pre-approval consultation. |
| Availability: | As needed |
| Results and Interpretations: | Preliminary (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. |
| Additional Information: | *Burkholderia pseudomallei* is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of *Burkholderia pseudomallei* detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation. |
| Purpose of Test: | To detect *Burkholderia pseudomallei* in clinical specimens / To confirm referred isolates |
| Method: | A variety of sentinel and LRN methods are used to grow, confirm or rule-out bacterial isolates. |
| Interfering Substances: | N/A |
| Comment: | Please call the Special Pathogen Laboratory with any questions or concerns. |
### Test

**Synonym:** Blood Metals (Cadmium (Cd), Lead (Pb), and Mercury (Hg) in Whole Blood)

**Lab Section/Phone:** Analytical Chemistry, 803-896-0886 or 803-896-0991

**Days Performed:** As requested

**Turnaround Time:** 5 Days

**Specimen Required:** 1 mL of whole blood from venipuncture

**Specimen Identification:** Specimen container must be labelled with patient’s full name, and a second patient identifier such as DOB, Specimen #, etc. DHEC requisition must be completed in full.

**Specimen Volume (optimum):** > 1 mL

**Specimen Volume (minimum):** 500 µL

**Collect:** In general, if more than one evacuated tube of blood is to be drawn from an individual, the blood metals tube should be drawn second or later. Draw the blood through a stainless-steel needle into a Vacutainer™.

**Form:** DHEC 1332, Test #882

**Special Instructions:** N/A

**Packing and Shipping**: See Transporting and Shipping Infectious Substances in Section IV.

**Transport Conditions:** Specimens can be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until receipt at the PHL. Specimens must be received for testing within 10 days of collection.

**Specimen Rejection Criteria:** Clotted blood, insufficient quantity (QNS), improper temperature. For universal rejections, See Section I.

**Availability:** Monday – Friday

**Results and Interpretations:** Blood lead levels in children under the age of 16 are considered elevated at or above 3.5 µg/dL and chelation treatment should be considered at confirmed blood lead levels of 45 µg/dL. The Occupational Safety and Health Administration regulations use a blood lead level of 40 µg/dL as cause for written notification and a medical exam, and a blood lead level of 60 µg/dL as cause for medical removal from exposure. Levels of concern for cadmium in blood is > 5 µg/L. The American Conference of Governmental Industrial Hygienists has a biological exposure index (BEI) of 15 µg/L for inorganic mercury in blood.

**Additional Information:** N/A

**Purpose of Test:** Identify exposure to Cadmium, Lead, and Mercury.

**Method:** Inductively Coupled Plasma Mass Spectrometry (ICP-MS)

**Interfering Substances:** N/A

**Comment:** N/A
<table>
<thead>
<tr>
<th>Test</th>
<th>CAMPYLOBACTER</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Organism for ID, Enteric Culture</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology – 803 896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate or PCR+ stool transport if unable to isolate Campylobacter isolate</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolates and Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Organism or Specimen should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Isolate – send culturette or slant.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Specimen – send a walnut sized portion of feces or 5-10ml of liquid stool in stool transport. Infant specimens may be collected in a disposable diaper with outside facing in.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Isolate ship on slant or culturette. Stool must be in transport medium.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, isolate Organism for ID, Stool Transport Enteric Cx.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store stool preserved in transport media in refrigerator at 2-8°C. Ship ALL specimens and isolates on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient, specimen too old, improper transport media or conditions, improper temperature. For universal rejections, See <a href="#">Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Campylobacter genus and species</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. If unable to isolate, ship stool in transport media, such as Cary Blair and Para-Pak ASAP, as the recovery of Campylobacter goes drastically down after 3 days from collection. Isolates once incubated overnight in microaerophilic conditions can be shipped within 3 business days on frozen cold packs in approved and specialized insulated shippers to maintain a temperature range of 2-8°C until received at the PHL.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>SC Disease Reportable Conditions required submission, Confirm or identify Campylobacter.</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>None</td>
</tr>
<tr>
<td>Comment:</td>
<td>Important – Maintain specimen at 2-8°C and ship to be received within 3 days of collection at the PHL in the temperature range of 2-8°C.</td>
</tr>
<tr>
<td>Test</td>
<td>CAMPYLOBACTER STOOL CULTURE</td>
</tr>
<tr>
<td>------</td>
<td>-----------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Enteric Culture</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology – 803 896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Stool in transport media.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Organism or Specimen should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Specimen – send a walnut sized portion of feces or 5-10 mL of liquid stool in stool transport. Infant specimens may be collected in a disposable diaper with outside facing in.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 – Enteric Culture</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store the stool preserved in transport media in refrigerator at 2-8°C. Ship ALL specimens and isolates in approved and specialized shippers on frozen cold packs to maintain a temperature range of 2-8°C for receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient, specimen too old, improper transport media or conditions, improper temperature. For universal rejections, See Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Campylobacter genus and species</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. If unable to isolate, ship stool in transport media, such as Cary Blair and Para-Pak ASAP, as the recovery of Campylobacter goes drastically down after 3 days from collection. Isolates once incubated overnight in microaerophilic conditions can be shipped within 3 business days on frozen cold packs in approved and specialized insulated shippers to maintain a temperature range of 2-8°C.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>SC Disease Reportable Conditions required submission, isolate Campylobacter from culture</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>None</td>
</tr>
<tr>
<td>Comment:</td>
<td>Important – Maintain specimen at 2-8°C and ship to be received within 3 days of collection at the PHL in the temperature range of 2-8°C.</td>
</tr>
<tr>
<td>Test</td>
<td>CANDIDA AURIS</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Candida not Candida albicans, Candida unable to speciate</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate submission on slant</td>
</tr>
<tr>
<td><strong>Submitter Identification Method</strong></td>
<td><strong>Isolates to Submit</strong></td>
</tr>
<tr>
<td>No identification</td>
<td>A random subset of isolates</td>
</tr>
<tr>
<td>Germ tube only</td>
<td>Germ tube-negative isolates</td>
</tr>
<tr>
<td>Chromagar only</td>
<td>Isolates that are <strong>NOT</strong> green or blue (so no C. albicans or C. tropicalis or C. dubliensies)</td>
</tr>
<tr>
<td>API 20C or API 32C</td>
<td>Isolates that are <strong>NOT</strong> C. albicans or C. tropicalis or C. dubliensies</td>
</tr>
<tr>
<td>VITEK 2, MicroScan, Phoenix</td>
<td>Isolates that are <strong>NOT</strong> C. albicans or C. tropicalis or C. dubliensies</td>
</tr>
<tr>
<td>MALDI-TOF or molecular identification</td>
<td>Isolates that are <strong>NOT</strong> C. albicans, C. tropicalis, C. parapsilosis, C. lusitaniae, C. dubliensies or C. krusei</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolates must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Organism isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Isolate of Candida possible auris on slant. See chart below.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 - mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Write on form any testing performed</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <strong>Transporting and Shipping Infectious Substances in Section IV.</strong></td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Isolate mixed, isolate not a Candida species, improper temperature; For universal rejections, See <strong>Section I</strong>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Candida species identified</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To identify possible Candida auris, or other rare newly emerging Candida species.</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>CHIKUNGUNYA IgM Capture ELISA</td>
</tr>
<tr>
<td>------</td>
<td>--------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Chik IgM Serology</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>15 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.5 mL</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator vacuum tube (SST), centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged, and serum is removed from the clot and put into a labeled secondary container/tube. Please follow manufacturer’s guidelines. See Specimen Collection: Venipuncture Procedure in Section III if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Serum specimens can be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until received at the PHL or can be stored frozen at -20°C or lower and shipped on dry ice to maintain the specimen at -20°C or lower until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>N/A</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Positive specimens will be referred to CDC for additional testing.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect IgM antibodies for the Chikungunya virus to determine a current infection.</td>
</tr>
<tr>
<td>Method:</td>
<td>IgM Capture ELISA (Enzyme-Linked Immunosorbent Assay)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>CHLAMYDIA (CT) DETECTION BY NUCLEIC ACID AMPLIFICATION</td>
</tr>
<tr>
<td>------</td>
<td>------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Gen-Probe, C. trachomatis Amplified Nucleic Acid Probe, Chlamydia rRNA, CT Aptima</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday-Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td><strong>Swab specimen</strong>: Endocervical, rectal and pharyngeal swab, and/or male urethral specimens in Gen-Probe Aptima Unisex Swab Specimen Collection Kit for Endocervical and Male Urethral Swab Specimens (Blue label/Blue collection swab). <strong>Vaginal specimens</strong>: Vaginal specimens are collected using the Gen-Probe Aptima Multitest Swab Specimen Collection kit. (Orange label/ Pink collection swab). <strong>Urine specimen</strong>: Patient should not have voided within one hour of collection. Collect first 20 - 30 mL of the first-catch urine stream into collection cup. Transfer 2 mL of urine into Aptima Urine Specimen Transport tube so that the urine level falls within the two lines on the transport tube labeled: “fill area”. (Yellow Label). Urine must be transferred to the Urine Collection Tubes within 24 hours. See GC/Chlamydia Gen-Probe Collection Procedure, Section III.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, or Specimen #. DHEC requisition must be completed.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Urine should be collected up to fall within the “fill area” lines. Swab collection kits should contain an adequate amount of transport media for testing.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Urine should be collected up to the “fill area” lines. Swab collection kits should contain the adequate amount of transport media for testing.</td>
</tr>
<tr>
<td>Collect:</td>
<td>See specimen requirements and GC/Chlamydia Gen-Probe Collection Procedure, Section III.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332, Test – CT only, Test – GC only and CT/GC, GC/Chlamydia/Trich. vaginalis</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Only use Gen-Probe Aptima specimen collection kit (unisex swab, multitest swab, or urine). <strong>Patients under the age of twelve should be tested by culture</strong>.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship urogenital swabs at 2-30°C, rectal and pharyngeal swabs at 4-30°C, and urine at 2-30°C. Swab specimens must be tested within 60 days of collection. Urine specimens should be tested within 30 days of collection. For longer storage, freeze transport tube within 7 days of collections at ≤ -20 °C.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen with no swab or incorrect swab in transport media; white swab in transport media; two swabs in transport media; urine above or below designated black lines on transport tube labeled fill area; swab specimen more than 60 days old, or urine specimen more than 30 days old. Specimens received at improper temperature. For universal rejections, See Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Positive, Negative, or Indeterminate</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>This test is not appropriate in cases of sexual assault or abuse; <strong>patients under the age of twelve should be tested by culture</strong>. A negative result does not preclude the presence of a CT or GC infection because results are dependent on adequate specimen collection, absence of inhibitors, and sufficient rRNA to be detected. Test results may be affected by improper specimen collection, improper specimen storage, technical error, or specimen mix-up. Therapeutic failure or success cannot be determined with the Aptima Combo 2 Assay since nucleic acid may persist following appropriate antimicrobial therapy.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>For the detection of Chlamydia in pharyngeal, rectal, vaginal, cervical, urethral and urine specimens.</td>
</tr>
<tr>
<td>Method:</td>
<td>Nucleic Acid Amplification Test</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>CORYNEBACTERIUM DIPHTHERIAE, CULTURE &amp; ID</td>
</tr>
<tr>
<td>----------------------------------------------------------------------</td>
<td>-----------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>C. diphtheriae</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate on slant; culture upon approval by CDC (throat swab, NP swab, skin, clinical material on Loeffler’s slant)</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolates and specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Organism or specimen should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, Organism for ID (referred isolate), Non-Enteric culture (CDC approval)</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Notify Clinical Microbiology Laboratory Section prior to submission; Specimens must be received within 24 hours of collection.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship specimens in approved shippers to maintain temperature of specimen within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Transport swab not used or ampule in transport swab not crushed. Culture: must be received within 24 hours of collection unless submitted on Loeffler’s medium. Specimens received at improper temperature. For universal rejections, see <a href="#">Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Per SC List of Reportable Conditions, specimen submission to the Public Health Laboratory (PHL) is required within 1 business day of reporting.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Conventional culture methods, Traditional Biochemicals</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>COVID-19</td>
</tr>
<tr>
<td>----------------------------------</td>
<td>--------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>SARS-CoV-2</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology/Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>6 days/week</td>
</tr>
<tr>
<td>Turn a Round time:</td>
<td>24 - 48 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Upper respiratory specimens (nasopharyngeal (NP), oropharyngeal, anterior nasal, and mid-turbinate nasal swabs, nasopharyngeal aspirate) and bronchoalveolar lavage (BALS) specimens</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Swab in 2 - 3 mL of viral transport media.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Swab in 1 mL of viral transport media.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Use only synthetic fiber swabs with plastic shafts. Do not use calcium alginate swabs or swabs with wooden shafts, as they may contain substances that inactivate some viruses and inhibit PCR testing. Place swabs immediately into sterile tubes containing 2 - 3 mL of viral transport media.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335OE</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV if needed.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store specimens at 2 - 8°C for up to 72 hours after collection. If specimens will ship without delay, ship overnight on frozen cold packs to maintain specimens at 2-8°C until received at the PHL. If a delay in shipping will result in receipt of the specimen at the PHL more than 72 hours after collection, store at ≤-20°C and ship specimens on dry ice to maintain temperature until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>6 days/week</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td></td>
</tr>
<tr>
<td></td>
<td><strong>Hologic Panther Fusion SARS-CoV-2 Assay</strong></td>
</tr>
<tr>
<td>Detected</td>
<td>SARS-CoV-2 detected</td>
</tr>
<tr>
<td>Not Detected</td>
<td>SARS-CoV-2 not detected</td>
</tr>
<tr>
<td>Invalid</td>
<td>Recollect specimen</td>
</tr>
<tr>
<td><strong>ThermoFisher COVID-19 Real Time PCR Assay Multiplex, ThermoFisher TaqPath</strong></td>
<td></td>
</tr>
<tr>
<td>Detected</td>
<td>Positive SARS-CoV-2</td>
</tr>
<tr>
<td>Not Detected</td>
<td>SARS-CoV-2 not detected</td>
</tr>
<tr>
<td>Inconclusive</td>
<td>Recollect specimen</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Qualitative detection of nucleic acid from the 2019-nCoV in upper respiratory specimens (such as nasopharyngeal swabs) collected from individuals who meet 2019-nCoV clinical and/or epidemiological criteria (e.g., clinical signs and symptoms associated with 2019-nCoV infection, contact with a probable or confirmed 2019-nCoV case, history of travel to geographic locations where 2019-nCoV cases were detected, or other epidemiologic links for which 2019-nCoV testing may be indicated as part of a public health investigation).</td>
</tr>
<tr>
<td>Method:</td>
<td>• ThermoFisher COVID-19 Real Time PCR Assay Multiplex, ThermoFisher TaqPath</td>
</tr>
<tr>
<td></td>
<td>• Hologic Panther Fusion SARS-CoV-2 Assay</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>CRE, CRPA, CRAB</td>
</tr>
<tr>
<td>------</td>
<td>-----------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>CRE = <strong>Carbapenem-resistant Enterobacterial</strong> (former Enterobacteriacea), Ship All, Do Not send duplicates. Only one isolate per patient regardless of source. Includes the following: <em>E.coli, Enterobacter, Klebsiella, Proteus, Providencia, Serratia, and Morganella.</em> (With the exceptions of <em>Serratia</em> which are both resistant to carbapenems and sensitive to a 3rd generation cephalosporin and <em>Enterobacter</em> spp. which are sensitive to Cefepime. These both represent a different mechanism of resistance than a carbapenemase.) Ertapenem non-susceptibility is the most sensitive indicator of carbapenemase production.&lt;br&gt;CRPA = Carbapenem resistant <em>Pseudomonas aeruginosa</em> <strong>Send all non-mucoid P. aeruginosa resistant to imipenem, meropenem, or doripenem AND Not Susceptible to cefepime or ceftazidime.</strong> Do not send duplicates.&lt;br&gt;CRAB = <strong>Carbapenem resistant Acinetobacter baumannii complex</strong> Send in all pan-resistant <em>Acinetobacter</em> spp.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate submitted on slant.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolates must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Organism Isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Carbapenem-resistant Enterobacteriaceae and Acinetobacter baumannii from all specimen types are required to be submitted.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition 1335, Mark CRE/CRPA/CRAB line</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a> if needed.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature of specimen within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient, specimen too old, improper transport media or conditions, improper temperature; For universal rejections, See <strong>Section I</strong>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Organism identification will be confirmed and reported, mCIM test will be set up and reported, all Positive and indeterminant mCIM isolates will have a PCR test performed to identify carbapenemase enzyme, and an AST (antimicrobial sensitivity test) will be performed.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>INCLUDE DRUG SUSCEPTIBILITY PROFILE, Specimen submission to the Public Health Laboratory (PHL) is required. Ship within 3 business days.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS, mCIM, Cepheid, STRECK kit, KBS, Sensititre</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>CRYPTOSPORIDIUM ANTIGEN</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Cryptosporidium antigen testing is available for outbreaks as determined by the SC DHEC Division of Acute Disease Epidemiology.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Walnut sized portion fresh stool or 3 mL of liquid stool, 10% formalin preserved stool, Cary-Blair, C &amp; S, or concentrated stool sediment. Specimen must be placed in a leakproof container. Do Not use PVA.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition form 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>, if needed.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store stool preserved in transport media in refrigerator at 2-8°C. Ship ALL specimens and isolates on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td><strong>Specimen preserved in PVA</strong>, improper labeling, improper temperature; For universal rejections, see <a href="#">Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Negative = Cryptosporidium antigen is absent or below detectable levels. Positive = Cryptosporidium antigen detected.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the presence of <em>Cryptosporidium</em> oocysts.</td>
</tr>
<tr>
<td>Method:</td>
<td>Rapid immunoassay for the qualitative detection of <em>Cryptosporidium parvum</em> antigen.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>The test is designed for use with stool specimens collected in an acceptable transport media. The use of colonic washes, aspirates or other diluted specimen types has not been established and could affect the performance of the assay. Stool specimens contaminated by products with an oily or particulate base (e.g., Barium, mineral oil, etc.) could interfere with the test and are not recommended.</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>DENGUE IgM</td>
</tr>
<tr>
<td>------------------------------</td>
<td>---------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Dengue IgM Serology</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>CDC Submission</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient's full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.5 mL serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator vacuum tube (SST) centrifuged appropriately according to manufacturer's guidelines. (Red top vacuum tubes may be used if the specimen is centrifuged, and serum is removed from the clot and put into a labeled secondary container/tube.) Please follow manufacturer’s guidelines. See <a href="#">Specimen Collection: Venipuncture Procedure in Section III</a>, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Paired specimens are NOT required.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Serum specimens may be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until received at the PHL or may be stored frozen at -20°C or lower and shipped on dry ice to maintain the specimen at -20°C or lower until received by the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Weekly</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td></td>
</tr>
<tr>
<td></td>
<td><img src="https://example.com/table.png" alt="Table" /></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Result</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative</td>
<td>No detectable IgM antibody, individual does not appear to be infected with Dengue virus. The result does not rule out Dengue virus infection.</td>
</tr>
<tr>
<td>Equivocal</td>
<td>Dengue virus IgM antibody cannot be determined. Submit another specimen for testing.</td>
</tr>
<tr>
<td>Positive</td>
<td>Presence of detectable IgM antibody, presumptive infection with Dengue virus. Confirmatory testing to follow. A positive IgM result may not indicate a recent infection because IgM may persist for several months after infection.</td>
</tr>
</tbody>
</table>

Additional Information: Positive results will be referred to CDC for additional testing.

Purpose of Test: To detect IgM antibodies for the Dengue virus to determine a current infection.

Method: IgM Capture ELISA (Enzyme-Linked Immunosorbent Assay)

Interfering Substances: N/A

Comment: N/A
<table>
<thead>
<tr>
<th>Test</th>
<th>EBOLA VIRUS REAL-TIME RT-PCR ASSAY (EBOLA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Zaire ebolavirus</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>24 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Whole blood, serum, and plasma; urine (cannot be the sole specimen)</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient's first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 521; check “Ebola”</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Pre-approval Needed - Hospitals must obtain approval from SC DHEC DADE (Division of Acute Disease Epidemiology) and the Special Pathogens Laboratory prior to submitting specimens. Contact information can be located on the back of the List of Reportable Conditions. Contact the Special Pathogens Laboratory (803-896-0777 / 803-767-8118) for test notification, specimen collection, storage conditions, and shipping conditions/methods.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>- Final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail.</td>
</tr>
<tr>
<td></td>
<td>- The definitive identification of Ebola virus requires additional testing to be performed by CDC.</td>
</tr>
<tr>
<td></td>
<td>- Negative EBOV NP rRT-PCR results do not preclude Ebola virus infection and should not be used as the sole basis for patient management decisions.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Ebola Virus is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of Ebola detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To presumptively identify Ebola RNA in clinical specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>CDC/LRN Real-Time RT-PCR Assay, EUA</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>ENTERIC GI PANEL by FilmArray (PCR)</td>
</tr>
<tr>
<td>------------------------------</td>
<td>------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>GI Panel, GI Outbreak</td>
</tr>
<tr>
<td>Bacteria: Campylobacter, Clostridium difficile toxin A/B, Plesiomonas shigelloides, Salmonella, Vibrio species, Vibrio cholerae, Yersinia enterocolitica;</td>
<td></td>
</tr>
<tr>
<td>Diarrheagenic E. coli/Shigella: Enteroaggregative E. coli (EAEC), Enteropathogenic E. coli (EPEC), Enterotoxigenic E. coli (ETEC) lt/st. Shiga-like producing E. coli (STEC) stx1/stx2, E. coli O157, Shigella/Enteroinvasive E. coli (EIEC);</td>
<td></td>
</tr>
<tr>
<td>Parasites: Cyclospora cayetanensis, Cryptosporidium, Entamoeba histolytica, Giardia lamblia;</td>
<td></td>
</tr>
<tr>
<td>Viruses: Adenovirus F 40/41, Astrovirus, Norovirus GI/GII, Rotavirus A, Sapovirus</td>
<td></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>Note: For same day test results, specimen must be received by noon.</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Stool (walnut sized portion of formed or 5 - 10 mL of liquid) preserved in Cary Blair media in transport tube.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Walnut sized portion of formed stool or 5 - 10 mL of liquid stool</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Stool preserved in Cary-Blair media transport tube</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Call Virology Laboratory</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store the stool preserved in transport media at 2-8°C. Ship on frozen cold packs to maintain a temperature range of 2-8°C for receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Unpreserved stool and specimen preserved in PVA, improper temperature; For universal rejections, see Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>N/A</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the presence of enteric pathogens in a GI outbreak situation.</td>
</tr>
<tr>
<td>Method:</td>
<td>Film Array GI panel (PCR)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
</tbody>
</table>
**Test** | **ENTERIC PATHOGENS CULTURE**
--- | ---
**Synonym:** | Fecal culture, stool culture, enteric culture, *Salmonella* culture, *Shigella* culture, *E coli* 0157 culture, *Campylobacter* culture, *Vibrio* culture, toxin culture for *Staphylococcus aureus*, *Bacillus cereus*, and *Clostridium perfringens*.
**Lab Section/Phone:** | Clinical Microbiology 803-896-3360
**Days Performed:** | Monday - Friday
**Turnaround Time:** | 10 Business days
**Specimen Required:** | Walnut sized portion of feces or 5-10 mL of liquid stool in stool transport. Infant specimens may be collected in a disposable diaper with plastic side facing inside.
**Specimen Identification:** | Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.
**Specimen Volume (optimum):** | N/A
**Specimen Volume (minimum):** | N/A
**Collect:** | Use stool transport such as Cary Blair or ParaPak
**Form:** | DHEC requisition 1335, mark Enteric Culture
**Special Instructions:** | Specimen must be maintained at 2-8°C.
**Packing and Shipping:** | See [Transporting and Shipping Infectious Substances in Section IV](#).
**Transport Conditions:** | Store the stool preserved in transport media at 2-8°C. Ship ALL specimens and isolates in approved and specialized shippers on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.
**Specimen Rejection Criteria:** | Quantity insufficient; specimen too old; improper transport media or conditions, improper temperature; For universal rejections, refer to [Section I](#).
**Availability:** | Monday - Friday
**Results and Interpretations:** | N/A
**Additional Information:** | N/A
**Purpose of Test:** | Culture and identification of the following pathogens: *Salmonella*, *Shigella*, *Campylobacter*, *Vibrio*, Shiga-toxin producing *Escherichia coli*, *Aeromonas*, *Yersinia enterocolitica*, *Plesiomonas shigelloides*, *Staphylococcus aureus*, *Bacillus cereus*, *Clostridium perfringens*.
**Method:** | Traditional culture, conventional biochemicals, serotyping, bioMerieux VITEK MS
**Interfering Substances:** | Do not use PVA
**Comment:** | Enteric Pathogen culture testing is available for outbreaks as determined by the SC DHEC DADE (Division of Acute Disease Epidemiology). Epidemiology to note on requisition slip which pathogens are suspected.
<table>
<thead>
<tr>
<th>Test</th>
<th>ENTERIC PATHOGENS submitted by NON-CULTURE INDEPENDENT METHODS (PCR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Walnut sized portion of feces or 5 - 10 mL of liquid stool in stool transport. Infant specimens may be collected in a disposable diaper with plastic side facing inside.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Note: For same day test results, specimens must be received by noon.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Use stool transport such as Cary Blair or ParaPak.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition 1335, mark Enteric Culture</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be maintained at 2-8°C.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store the stool preserved in transport media at 2-8°C. Ship ALL specimens and isolates in approved and specialized shippers on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient; specimen too old; improper transport media or conditions, improper temperature; For universal rejections. See <a href="#">Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. Ship ASAP on cold packs in insulated shippers to improve recovery of PCR+ organism.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Culture and identification of the following pathogens: <em>Salmonella</em>, <em>Shigella</em>, <em>Campylobacter</em>, <em>Vibrio</em>, Shiga-toxin producing <em>Escherichia coli</em>, <em>Aeromonas</em>, <em>Yersinia enterocolitica</em>, <em>Plesiomonas shigelloides</em>.</td>
</tr>
<tr>
<td>Method:</td>
<td>Traditional culture, conventional biochemicals, serotyping, bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>Do not use PVA</td>
</tr>
</tbody>
</table>
| Comment: | Important – Maintain specimen at 2-8°C and ship to be received within 3 days of collection at the PHL in the temperature range of 2-8°C.
<table>
<thead>
<tr>
<th>Test</th>
<th>ESCHERICIA COLI – SHIGA-TOXIN PRODUCING</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td><em>E. coli</em> O157:H7, <em>E. coli</em> non-O157, STEC, EHEC, Shiga toxin positive</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate, or PCR+ stool transport/ broth if unable to isolate.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate, Broth, or Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, mark Culture/Isolate for Shiga toxin producing <em>E. coli</em> or Broth/specimen for Shiga toxin producing <em>E. coli</em>, as appropriate.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <strong>Transporting and Shipping Infectious Substances in Section IV</strong>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature of specimen within the range of 15-25°C until received at the PHL. Store the stool preserved in stool transport and broth at 2-8°C and ship ALL specimens and broths in approved and specialized shippers on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient; specimen too old; improper transport media or conditions, improper temperature; For universal rejections, refer to <strong>Section I</strong>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. Ship PCR + stool transport specimens and broths ASAP to increase ability to recover isolate. Ship Shiga toxin positive isolates within 1 business day.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Culture as needed and identification of Shiga-toxin producing <em>E. coli</em></td>
</tr>
<tr>
<td>Method:</td>
<td>Immunochromatographic rapid test for Shiga-toxin</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Test</th>
<th>FOODBORNE ILLNESSES (FOOD POISONING)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Days Performed:</td>
<td>Monday- Friday</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>The Food Microbiology section assists in the epidemiological investigation of suspected foodborne illness. A physician with a patient suspected of having a foodborne illness should contact <strong>Food Protection in the county health department</strong>. The laboratory does not accept specimens from patients.</td>
</tr>
<tr>
<td>Test</td>
<td>FRANCISELLA TULARENSIS</td>
</tr>
<tr>
<td>------</td>
<td>------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Tularemia, rabbit fever, deerfly fever</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803)767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>7 to 10 days from the time of specimen receipt in the laboratory</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical Specimens / isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during pre-approval consultation</td>
</tr>
</tbody>
</table>
| Form: | DHEC 1335; Test Number 520; “Suspect Agent” = *F. tularenia*  
DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen. |
<p>| Special Instructions: | Specimen must be pre-approved by Special Pathogens Supervisor prior to testing. |
| Packing and Shipping*: | Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118. |
| Transport Conditions: | Determined during pre-approval consultation |
| Specimen Rejection Criteria: | Determined during pre-approval consultation |
| Availability: | As needed |
| Results and Interpretations: | Preliminary (when applicable) and final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. |
| Additional Information: | <em>Francisella tularensis</em> is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of <em>Francisella tularensis</em> detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation |
| Purpose of Test: | To detect <em>F. tularensis</em> in clinical specimens / To confirm referred isolates |
| Method: | A variety of sentinel and LRN methods are used to grow, confirm, or rule-out bacterial isolates. |
| Interfering Substances: | N/A |
| Comment: | Please call the Special Pathogen Laboratory with any questions or concerns |</p>
<table>
<thead>
<tr>
<th>Test</th>
<th>GIARDIA ANTIGEN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>N/A</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Walnut sized portion fresh stool or 3 mL of liquid stool, 10% formalin preserved stool, Cary-Blair, C &amp; S, or concentrated stool sediment. Specimen must be placed in a leakproof container. Do Not use PVA.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition form 1335, Mark Cryptosporidium Antigen line.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transferring and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store the stool preserved in stool transport at 2-8°C. Ship ALL specimens and isolates in approved and specialized shippers on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL within 3 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td><strong>Specimen preserved in PVA</strong>, improper labeling, improper temperature; For universal rejections, refer to <a href="#">Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Negative = Giardia antigen is absent or below detectable levels. Positive = Giardia antigen detected.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the presence of Giardia antigen.</td>
</tr>
<tr>
<td>Method:</td>
<td>Rapid immunoassay for the qualitative detection of <em>Cryptosporidium parvum</em> and <em>Giardia lamblia</em>.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>The test is designed for use with stool specimens collected in an acceptable transport media. The use of colonic washes, aspirates or other diluted specimen types have not been established and may affect the performance of the assay. Stool specimens contaminated by products with an oily or particulate base (e.g., Barium, mineral oil, etc.) may interfere with the test and are not recommended.</td>
</tr>
<tr>
<td>Comment:</td>
<td><strong>Giardia antigen testing is available for outbreaks as determined by the SC DHEC Division of Acute Disease Epidemiology (DADE).</strong></td>
</tr>
</tbody>
</table>
**Test** | **GI OUTBREAK**
--- | ---
Synonym: | Norwalk Virus, Norovirus PCR, Enteric Outbreak
Lab Section/Phone: | Virology & Rabies, 803-896-0819
Days Performed: | Monday - Friday
Turnaround Time: | N/A
Specimen Required: | Two separate collections are required:
1. For Norovirus Detection by Real-Time PCR, a peanut-sized or tablespoon volume of fresh diarrheal stool. Specimens collected within 48 - 72 hours of symptom onset are preferred. Specimens collected within 10 days of symptom onset will be accepted. Rectal swabs are not acceptable.
2. For Enteric Pathogens Culture, a walnut sized portion of feces or 5 - 10 mL of liquid stool in stool transport. Infant specimens may be collected in a disposable diaper with plastic side facing inside.
Specimen Identification: | Patient’s full name and patient ID # (or other unique identifier) is required on both specimens and requisition.
Specimen Volume (optimum): | N/A
Specimen Volume (minimum): | 1. For Norovirus Detection by Real-Time PCR, a peanut-sized or tablespoon volume of fresh diarrheal stool.
2. For Enteric Pathogens Culture, a walnut sized portion of feces or 5 - 10 mL of liquid stool in stool transport.
Collect: | 1. For stool for Norovirus Detection by Real-Time PCR, use a sterile, screw capped, leak-proof, 50 mL conical tube or urine container.
2. For Enteric Pathogens Culture, use transport tube with Cary-Blair medium included in Enteric Kit provided by the Public Health Laboratory
Form: | DHEC 1335; When ordering this test panel, please write GI Outbreak on the submission form.
Special Instructions: | Use of this test is restricted to Epidemiological investigations. This test should be used when a GI outbreak is suspected, and multiple etiologies are suspected. Please consult your Regional Epidemiological contact.
Packing and Shipping*: | See Transporting and Shipping Infectious Substances in Section IV.
Transport Conditions: | Store and ship to be maintained at 2-8 °C until received at the PHL.
Specimen Rejection Criteria: | See Specimen Rejection & Disclaimer Criteria in Section I.
Availability: | N/A
Results and Interpretations: | | Result | Interpretation |
| | Detected | Organism detected |
| | Not detected | No organism detected |
Additional Information: | Please write GI Outbreak on the submission form. This panel designates a testing algorithm for GI outbreak of unknown etiology. This panel includes tests for Norovirus rRT-PCR, BioFire FilmArray GI Panel, and enteric culture (in this order). Testing will cease when a positive identification is made. If enteric pathogens other than Salmonella, E. coli O157:H7 or Shigella are suspected, please specify.
Purpose of Test: | GI Outbreak testing is available for outbreaks as determined by the SC DHEC Division of Acute Disease Epidemiology.
Method: | 1. Norovirus: Real-time reverse transcriptase polymerase chain reaction (real-time RT-PCR)
2. BioFire GI Panel: Multiplex PCR Panel
3. Enteric Pathogens Culture: Traditional culture, conventional biochemicals, serotyping, bioMerieux VITEK MS
Interfering Substances: | N/A
Comment: | N/A
<table>
<thead>
<tr>
<th><strong>Test</strong></th>
<th><strong>GONOCOCCAL (GONORRHEA) CULTURE</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>GC culture, <em>Neisseria gonorrhoeae</em> culture</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Transgrow bottle</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Transgrow bottle must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full. <strong>Do Not place label over clear glass viewing area, layer patient label over existing label.</strong></td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>See <em>N. gonorrhoeae Collection Procedure, Section III.</em></td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Bring transgrow bottle to room temperature before inoculating. <strong>Hold bottle upright</strong> and roll swab over entire surface of medium; discard swab. <strong>NOTE:</strong> Ship for overnight delivery. Do not ship specimens for arrival over a weekend.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, Mark GC Culture &amp; ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td><strong>Collect specimens Monday thru Wednesday ONLY.</strong></td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <em>Transnporting and Shipping Infectious Substances in Section IV.</em> If an incubator is available, incubate inoculated transgrow bottle upright at 35°C until shipped, and indicate incubation time on request form. If an incubator is not available, make sure culture is shipped on the same day as collected. <strong>DO NOT REFRIGERATE AFTER INOCULATION.</strong></td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Transgrow media not used; specimen in transit more than 5 days, improper temperature; For universal rejections, See <em>Section I.</em></td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Wednesday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td><em>Neisseria gonorrhoeae</em> isolated or not isolated.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td><strong>If Drug Treatment failure is expected, notate this on DHEC requisition. If <em>Neisseria gonorrhoeae</em> is isolated, isolate will be sent out for Antimicrobial Susceptibility Testing (AST).</strong></td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Culture for growth of <em>Neisseria gonorrhoeae</em>, this is needed if drug treatment failure is expected.</td>
</tr>
<tr>
<td>Method:</td>
<td>BioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>GONOCOCCAL (GC) DETECTION by NUCLEIC ACID AMPLIFICATION</td>
</tr>
<tr>
<td>----------------------------------</td>
<td>--------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Gen-Probe N. gonorrhoeae Amplified Nucleic Acid Probe, Gonorrhea rRNA, GC Aptima</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday-Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1-5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td><strong>Swab specimen</strong>: Endocervical, rectal and pharyngeal swab, and/or male urethral specimens in Gen-Probe Aptima Unisex Swab Specimen Collection Kit for Endocervical and Male Urethral Swab Specimens (Blue Label/blue collection swab). <strong>Vaginal specimens</strong>: Vaginal specimens are collected using the Gen-Probe Aptima Multitest Swab Specimen Collection kit. (Orange label/ Pink collection swab). <strong>Urine specimens</strong>: Patient should not have voided within one hour of collection. Collect 20 - 30 mL of the first-catch urine stream into collection cup. Transfer 2 mL of urine into Aptima Urine Specimen Transport tube so that the urine level falls within the two lines on the transport tube labeled: “fill area”. (Yellow Label). Urine must be transferred to the Urine Collection Tubes within 24 hours. See GC/Chlamydia Gen-Probe Collection Procedure, Section III.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, or Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Urine should be collected up to fall within the “fill area” lines. Swab collection kits should contain the adequate amount of transport media for testing.</td>
</tr>
<tr>
<td>Collect:</td>
<td>See specimen requirements and GC/Chlamydia Gen-Probe Collection Procedure, Section III.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Only use Gen-Probe Aptima specimen collection kit (unisex swab, vaginal swab, or urine). Patients under the age of twelve should be tested by culture.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship urogenital swabs at 2-30°C, rectal and pharyngeal swabs at 4-30°C, and urine at 2-30°C. Swab specimens must be tested within 60 days of collection. Urine specimens must be tested within 30 days of collection. For longer storage, freeze transport tube within 7 days of collection at ≤ -20 °C and ship on dry ice to maintain at temperature of ≤ -20°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen with no swab or incorrect swab in transport media; white swab in transport media; two swabs in transport media; urine above or below designated black lines on transport tube labeled fill area; swab specimen more than 60 days old, or urine specimen more than 30 days old; specimens received at the improper temperature; For universal rejections, See Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Positive, Negative, or Indeterminate</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>This test is not appropriate in cases of sexual assault or abuse; patients under the age of twelve should be tested by culture. A negative result does not preclude the presence of a CT or GC infection because results are dependent on adequate specimen collection, absence of inhibitors, and sufficient rRNA to be detected. Test results may be affected by improper specimen collection, improper specimen storage, technical error, or specimen mix-up. Therapeutic failure or success cannot be determined with the Aptima Combo 2 Assay since nucleic acid may persist following appropriate antimicrobial therapy.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>For the detection of Neisseria gonorrhoeae in pharyngeal, rectal, vaginal, cervical, urethral and urine specimens.</td>
</tr>
<tr>
<td>Method:</td>
<td>Nucleic Acid Amplification Test</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HAEMOPHILUS INFLUENZAE</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td><em>H. influenza</em></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Agar slant that will support growth of isolate</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Pure bacterial isolate on agar slant (chocolate agar is preferred)</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition form 1335. Mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Inoculate chocolate agar slant with isolated organism, incubate overnight in 35°C CO2 incubator, observe for growth, and ship isolate to maintain temperature of specimen within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <em>Transporting and Shipping Infectious Substances in Section IV</em>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates to maintain temperature of specimen within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Culture non-viable; culture mixed, improper temperature; For universal rejections, See <em>Section I</em>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Only <em>H. influenzae</em> isolates from normally sterile sites should be tested. Always specify site of isolate. Urgently reportable; ship within 1 business day.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Confirm identification of <em>Haemophilus influenzae</em> and serotype.</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS, serotyping</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEMOGLOBIN (Hgb) ELECTROPHORESIS</td>
</tr>
<tr>
<td>------</td>
<td>----------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Adult Sickle Cell Screen</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Newborn Screening, 803-896-0874 or 803-896-0891</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Upon Request</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Dried blood spot; collected on DHEC 1339 collection form</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and date of birth written on DHEC 1339 collection form</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 filled circles on DHEC 1339 collection form</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 filled circle on DHEC 1339 collection form</td>
</tr>
<tr>
<td>Collect:</td>
<td>Fingerstick</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1339</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Fill each circle with one large blood drop that soaks through to the other side. Do not layer blood drops. Allow the specimen(s) to dry horizontally for 3 - 4 hours before packing for shipment. To protect the specimen, fold over the biohazard labeled flap once the specimen is dry.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Place dried and covered specimen in paper/cardboard mailer. Do not ship in plastic.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Do NOT use plastic bags or any other airtight, leakproof, or sealed containers.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen(s) received in plastic bags; specimens collected on expired collection forms; specimens older than 14 days; specimen quality and/or quantity inadequate.</td>
</tr>
<tr>
<td>Availability:</td>
<td>N/A</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Screen for abnormal hemoglobin’s</td>
</tr>
<tr>
<td>Method:</td>
<td>Iso-electric focusing and/or High-Performance Liquid Chromatography</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS A SEROLOGY</td>
</tr>
<tr>
<td>----------------------</td>
<td>--------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>HAVAB-G; Anti-HAV; HAVAB-IgG; Antibody to Hepatitis HAV-IgG; Anti-HAV, IgG; Antibody to Hepatitis A Virus, IgG; HAVAB-M; HAVAB-IgM; Antibody to HAV-IgM; Anti-HAV, IgM; Antibody to Hepatitis A Virus, IgM</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Upon request</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1.0 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.5 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332, Test - Hepatitis A, IgG; Test - Hepatitis A, IgM</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>All Hepatitis A outbreak investigations should be reported to the laboratory supervisor (803-896-0811) or Division Director (803-896-0870) prior to shipment of specimens. After collecting the specimen, invert the serum-separator tube gently 5 times, allow to clot for at least 30 minutes and centrifuge within 2 hours from the time of collection. Specimens must be centrifuged within 2 hours of collection to separate the serum from the clot.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 7 days of collection at the PHL; for storage longer than 7 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 7 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Testing performed as needed.</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reactive or Nonreactive</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>For the detection of Hepatitis A in serological specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence (CMIA)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B CORE IgM ANTIBODY</td>
</tr>
<tr>
<td>------</td>
<td>-----------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Anti-HBc, IgM; HBcAb, IgM; Antibody to Hepatitis B Core Antigen, IgM</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Available upon request</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>0.5 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.25 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 7 days of collection at the PHL; for storage longer than 7 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 7 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reactive or Nonreactive</td>
</tr>
<tr>
<td>Additional Information:</td>
<td><em>Test automatically performed on patients with reactive anti-HBc total antibody in absence of reactive HBsAg or anti-HBs on Diagnostic Profile (test #223), and test automatically performed on patients with reactive Hepatitis B surface antigen on Diagnostic Test Panel #223</em>.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B CORE TOTAL ANTIBODY SCREEN</td>
</tr>
<tr>
<td>------</td>
<td>--------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Anti-HBc; Core Antibody; HBcAb, Total; Antibody to Hepatitis B Core Antigen</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient's first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1.0 mL of Serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.5 mL of Serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332, Test #226</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>After collecting the specimen, invert the serum-separator tube gently 5 times, allow to clot for at least 30 minutes and centrifuge within 2 hours from the time of collection. Specimens must be centrifuged within 2 hours of collection to separate the serum from the clot. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 7 days of collection at the PHL; for storage longer than 7 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 7 days old when received, not shipped on dry ice and received at -20°C or colder. A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reactive or Nonreactive</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B DIAGNOSTIC PROFILE</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Includes tests for HBsAg, anti-HBs, and anti-HBc, and anti-core IgM are performed if indicated.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>2 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>After collecting the specimen, invert the serum-separator tube gently 5 times, allow to clot for at least 30 minutes and centrifuge within 2 hours of collection. Specimens must be centrifuged within 2 hours of collection to separate the serum from the clot. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 6 days of collection at the PHL; for storage longer than 6 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 6 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Includes tests for HBsAg, anti-HBs and anti-HBc, and anti-core IgM, if indicated.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>N/A</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Specimen requirements allow for HBsAg, anti-HBs and anti-HBc, and anti-core IgM, to be performed, if indicated.</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B IMMUNE STATUS/POST-IMMUNIZATION</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>---------------------------------------------</td>
</tr>
<tr>
<td>Synonym</td>
<td>Anti-HBs and Anti-HBc</td>
</tr>
<tr>
<td>Lab Section/Phone</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum)</td>
<td>1.5 mL of Serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum)</td>
<td>1.0 mL of Serum</td>
</tr>
<tr>
<td>Collect</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions</td>
<td>Tests include Anti-HBs and Anti-HBc</td>
</tr>
<tr>
<td>Packing and Shipping*</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions</td>
<td>Store and ship to be maintained at 2-8°C and received within 7 days of collection at the PHL; for storage longer than 7 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 7 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test</td>
<td>N/A</td>
</tr>
<tr>
<td>Method</td>
<td>Chemiluminescence</td>
</tr>
<tr>
<td>Interfering Substances</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B SURFACE ANTIBODY</td>
</tr>
<tr>
<td>------</td>
<td>-----------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>HBsAb; Anti-HBs; Antibody to Hepatitis B Surface Antigen</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1.5 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1.0 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See <strong>Specimen Collection: Venipuncture Procedure in Section III</strong>, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <strong>Transporting and Shipping Infectious Substances in Section IV</strong>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 7 days of collection at the PHL; for storage longer than 7 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 7 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see <strong>Specimen Rejection &amp; Disclaimer Policies in Section I</strong>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday-Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence (CMIA)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS B SURFACE ANTIGEN</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-----------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>HBsAg; Hepatitis-Associated Antigen</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1.5 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1.0 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 6 days of collection at the PHL; for storage longer than 6 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens; specimens greater than 6 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS C TOTAL ANTIBODY</td>
</tr>
<tr>
<td>------</td>
<td>--------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Antibody to Hepatitis C Virus; Anti-HCV</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday-Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with the patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>3 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.25 mL of serum (if reactive, a total of 2.25 mL serum needs to be collected and sent for confirmatory testing).</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>For sites requesting HCV RNA if total antibody reactive by EIA, collect blood in a serum separator tube, spin down within 6 hours of collection. Label outside of box “HCV Viral Load” with indelible marker or sticker that cannot easily be removed. Viral loads can be shipped with any STD specimen but MUST be packed on frozen cold packs to maintain specimen at a temperature of 2-8 °C until received at the PHL.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 5 days of collection at the PHL; for storage longer than 5 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improperly stored/shipped, heat-inactivated, pooled, grossly hemolyzed or microbial contaminated specimens (Test #224 only); specimens greater than 5 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reactive Confirmed, Reactive Not Confirmed, Nonreactive, Equivocal Confirmed, or Equivocal Not confirmed</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Reactive specimens that were shipped/collected appropriately (in an SST, centrifuged, and shipped on cold packs) will be reflexed to viral load testing automatically.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Chemiluminescence (CMIA)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Positive HCV Total Antibody results will be confirmed using the HCV Viral Load test, provided the Special Instructions listed above are followed. Specimen requirements allow for reflexed testing to be performed, if needed.</td>
</tr>
<tr>
<td>Test</td>
<td>HEPATITIS C QUANTITATION BY PCR (RNA)</td>
</tr>
<tr>
<td>------</td>
<td>------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>HCV Viral Load test</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>3 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube (SST) or serum. Tubes must be adequately centrifuged, and serum from red top tubes must be removed from the clot and put in a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Collect blood in a serum separator tube (SST) or red top tube, allow to clot for at least 30 minutes, spin down within 6 hours of collection, and ship SST or serum at 2-8°C on frozen cold packs to arrive within 5 days of collection (please send in as soon as possible). Label outside of box “HCV Viral Load” with indelible marker or sticker that cannot easily be removed. Viral loads can be shipped with any STD specimen, but MUST be packed on frozen cold packs to maintain specimen at a temperature of 2-8 °C until received at the PHL.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 5 days of collection at the PHL; for storage longer than 5 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Whole clotted blood not centrifuged and separated within 6 hours of collection; specimens greater than 5 days old when received, not shipped on dry ice and received at -20°C or colder; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see Specimen Rejection &amp; Disclaimer Policies in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>The measurable reportable range for this procedure is 10-10,000,000 IU/mL and 1.00-7.0 log 10; Specimen testing within this range will be reported as the measured IU/mL value and the log 10 value of the measured IU/mL value e.g., 30,000 IU/mL and 4.48 log 10. Specimens testing above 10,000,000 will be reported as &gt; 10,000,000 IU/ mL and &gt; 7.0 log 10. Specimens testing less than 10 IU/mL and less than 1.00 log 10 will be reported as less than &lt; 10 IU/mL as and less than &lt; 1.00 log 10. Specimens with nothing detected will be reported as Not Detected.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Used to aid in the detection and quantitation of HCV infections</td>
</tr>
<tr>
<td>Method:</td>
<td>Nucleic acid amplification test (RT-TMA)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
</tbody>
</table>
**Test:** HERPES SIMPLEX 1 & 2 Assay

| **Synonym:** | N/A |
| **Lab Section/Phone:** | Virology & Rabies, 803-896-0819 |
| **Days Performed:** | Monday - Friday |
| **Turnaround Time:** | 5 Days |
| **Specimen Required:** | Swab specimens from anogenital lesions ONLY, placed in the Aptima Multitest Swab Specimen Collection Kit (Orange Tube) |
| **Specimen Identification:** | Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition. |
| **Specimen Volume (optimum):** | N/A |
| **Specimen Volume (minimum):** | N/A |
| **Collect:** | Polyester-tipped swab specimens from anogenital lesions ONLY placed in Aptima Multitest Swab Specimen Collection Kit (Orange Tube) (available upon request; Ordering Supplies in Section III, p.1) |
| **Form:** | DHEC 1335 |
| **Special Instructions:** | N/A |
| **Packing and Shipping*:** | See Transporting and Shipping Infectious Substances in Section IV. |
| **Transport Conditions:** | Store and ship within one week of collection; maintain specimen at 2-30°C until received at the PHL. |
| **Specimen Rejection Criteria:** | See Specimen Rejection & Disclaimer Criteria in Section I. |
| **Availability:** | Monday - Friday |

**Results and Interpretations:**

<table>
<thead>
<tr>
<th>HSV-1 Result</th>
<th>HSV-2 Result</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>HSV-1 neg</td>
<td>HSV-2 neg</td>
<td>Negative: No HSV-1 or HSV-2 mRNA detected</td>
</tr>
<tr>
<td>HSV-1 neg</td>
<td>HSV-2 pos</td>
<td>HSV-2 positive: HSV-2 mRNA detected</td>
</tr>
<tr>
<td>HSV-1 pos</td>
<td>HSV-2 neg</td>
<td>HSV-1 positive: HSV-1 mRNA detected</td>
</tr>
<tr>
<td>HSV-1 pos</td>
<td>HSV-2 pos</td>
<td>HSV-1 and HSV-2 positive: HSV-1 and HSV-2 mRNA detected</td>
</tr>
</tbody>
</table>

**Additional Information:** Please do NOT mark “wound, pus, drainage” or write “lesion”, but rather specify “Genital” as the specimen or include where the lesion is located in the anogenital region.

**Purpose of Test:** Qualitative detection and differentiation of messenger RNA (mRNA) from Herpes simplex virus type1 (HSV-1) and type 2 (HSV-2) DNA.

**Method:** Nucleic Acid Amplification Test (NAAT) using real-time transcription-mediated amplification (TMA)

**Interfering Substances:** N/A

**Comment:** N/A
<table>
<thead>
<tr>
<th>Test</th>
<th>HIV-1 PCR QUANTITATIVE (RNA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>HIV-1 Viral Load Test</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Minimum 2.0 mL EDTA anticoagulated plasma, See Specimen Collection: Venipuncture Procedure in Section III, if needed. If using EDTA vacutainer, separate the plasma from the packed cells within 24 hours of collection by centrifugation for 20 minutes at room temperature (18-30°C). Remove the plasma from the cells using a sterile transfer pipette to a sterile polypropylene transport tube. Note: Remove as much of the plasma from the cells as possible without aspirating the cells. The assay requires 1.0 mL of plasma. The PPT separator tube can be shipped after centrifugation without transferring plasma to another tube. Invert tube after centrifugation to ensure complete separation of cells from plasma. If cells are present in plasma, re-centrifuge before shipping.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2.0 mL of plasma</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1.0 mL of plasma</td>
</tr>
<tr>
<td>Collect:</td>
<td>PPT vacutainer (supplied by the Public Health Laboratory call 803-896-0913 to order) or polypropylene tube to which plasma cells have been transferred from the Lavender top (EDTA) vacuum tube or K2 EDTA with gel separator.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>The specimen MUST BE kept at 2-8 °C. Label outside of shipping container as “HIV-1”. Make sure label will not come off. Please check with the laboratory during a holiday to ensure that it will arrive within 3 days of collection or freeze the specimen at ≤ -20°C and ship on dry ice to maintain specimen at ≤ -20°C until received at the PHL.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship at 2° - 8 °C. Transport on frozen cold packs in a shipping container labeled on the outside of the container as “HIV-1”. Specimens must arrive at the PHL within 3 days of collection. Viral loads can be shipped with any STD specimen but MUST be packed on frozen cold packs to maintain specimen at a temperature of 2° - 8 °C until received at the PHL. If specimen will not be received at the PHL within 3 days of collection, transfer plasma into a secondary container and freeze the plasma at ≤ -20°C and then ship on dry ice to maintain specimen at ≤ -20°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Clotted whole blood specimens and specimens &gt;3 days old not maintained at ≤ -20°C or colder; improper temperature.; For universal rejections, See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>The measurable reportable range for this procedure is 30 - 10,000,000 copies/mL and 1.47 - 7.0 log 10. Specimens testing within this range will be reported as the measured copy value and the log 10 value of the measured copy value e.g., 30,000 copies/mL and 4.48 log 10. Specimens testing above 10,000,000 will be reported as &gt; 10,000,000 copies/mL and &gt; 7.0 log 10. Specimens testing less than &lt; 30 copies/mL and less than &lt; 1.47 log 10 will be reported as less than &lt; 30 copies/mL and less than &lt; 1.47 log 10. Specimens with nothing detected will be reported as Not Detected.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Therapeutic monitoring of HIV infection</td>
</tr>
<tr>
<td>Method:</td>
<td>Nucleic acid amplification test</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
</tbody>
</table>

II-39
**Test**  |  **HIV-1/HIV-2 SEROLOGY**
--- | ---
**Synonym:**  | HIV-1/HIV-2 antibody, HIV-1, HIV-2 antibodies, HIV-1 antigen
**Lab Section/Phone:**  | Diagnostic Serology, 803-896-0811
**Days Performed:**  | Monday – Friday
**Turnaround Time:**  | 1 - 5 Business Days
**Specimen Required:**  | Serum
**Specimen Identification:**  | Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.
**Specimen Volume (optimum):**  | 1.5 mL of serum
**Specimen Volume (minimum):**  | 1 mL of serum
**Collect:**  | Serum-separator tube (SST) or serum. Tubes must be properly centrifuged, and serum from red top tubes must be removed from the clot and put into a different labeled container/tube. See [Specimen Collection: Venipuncture Procedure in Section III](#), if needed.
**Form:**  | DHEC 1332
**Special Instructions:**  | N/A
**Packing and Shipping:**  | See [Transporting and Shipping Infectious Substances in Section IV](#).
**Transport Conditions:**  | Specimens that will be received at the PHL within 2 days of collection can be stored and shipped at 2-30°C. Store specimens at 2 - 8°C that will not be received at the PHL within 2 days of collection, but will be received within 7 days of collection, and transport on frozen ice packs to maintain specimens at 2 - 8°C until receipt at the PHL. For specimens that will not be received at the PHL within 7 days of collection, remove the serum from the clot or gel, place in secondary container, and freeze the serum at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.
**Specimen Rejection Criteria:**  | Specimens greater than 7 days old not maintained at -20 °C or colder when received; specimens received at the improper temperature; A second specimen will need to be collected if any specimens are rejected. For universal rejections, see [Specimen Rejection & Disclaimer Criteria in Section I](#).
**Availability:**  | Monday - Friday
**Results and Interpretations:**  | Reactive, Nonreactive
**Additional Information:**  | Repeatedly reactive specimens are confirmed by Geenius HIV 1/2. Repeat reactive specimens not confirmed by Geenius HIV1/2 will be submitted for HIV-1 NAT. Recommend repeat testing on all first-time positive patient results including CD4 and Viral load (HIV-1 RNA).
**Purpose of Test:**  | To aid in the detection and diagnosis of HIV-1/HIV-2
**Method:**  | Multiplex flow immunoassay
**Interfering Substances:**  | N/A
**Comment:**  | Specimen requirements allow for reflexed testing to be performed, if needed.
<table>
<thead>
<tr>
<th><strong>Test</strong></th>
<th><strong>INFLUENZA A: H5N1 (ASIAN CLADE)</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Avian Flu/ Bird Flu</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evening - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>48 Hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Throat swabs, Nasal washings/aspirates, nasopharyngeal swabs, sputum, bronchoalveolar lavage, tracheal aspirates, and bronchial washings.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335-OE; in the Special Pathogens test section, check “Avian Influenza box” and indicate “H5” as Agents/Organism/or Virus Suspected.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Pre-approval Needed - Hospitals must obtain approval from the DHEC health department and Public Health Laboratory prior to submitting specimens to the Special Pathogens Laboratory. Contact information can be located on the back of the List of Reportable Conditions. Contact the Special Pathogens Laboratory (803)896-0777 / (803)767-8118 for test notification, specimen collection, storage conditions, and shipping conditions/methods.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at (803)896-0777 / (803)767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. The definitive identification of Influenza A:H5N1 virus requires additional testing to be performed by CDC.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Testing for Influenza A: H5N1 will be concurrent with Influenza A:H7N9 testing</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To presumptively detect Influenza A:H5N1 RNA in clinical specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>CDC Real Time RT-PCR Assay, EUA</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td><strong>INFLUENZA A: H7N9 (EURASIAN LINEAGE)</strong></td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Avian Flu / Bird Flu</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>48 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Throat swabs, Nasal washings / aspirates, nasopharyngeal swabs, sputum, bronchoalveolar lavage, tracheal aspirates, and bronchial washings.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 -OE; in the Special Pathogens test section, check “Other” box and indicate “H7” as Agents/Organism/or Virus Suspected.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Pre-approval Needed - Hospitals must obtain approval from the DHEC health department and Public Health Laboratory prior to submitting specimens to the Special Pathogens Laboratory. Contact information can be located on the back of the List of Reportable Conditions. Contact the Special Pathogens Laboratory (803)896-0777 / (803)767-8118 for test notification, specimen collection, storage conditions, and shipping conditions / methods.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during Special Pathogen notification.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. The definitive identification of Influenza A:H7N9 virus requires additional testing to be performed by CDC.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Testing for Influenza A: H5N1 will be concurrent with Influenza A:H7N9 testing</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To presumptively detect Influenza A:H7 RNA in clinical specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>CDC Real Time RT-PCR Assay, EUA</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns</td>
</tr>
</tbody>
</table>
Test | INFLUENZA DETECTION BY REAL-TIME (RT) PCR
--- | ---
Synonym: | Influenza Surveillance, Influenza Detection
Lab Section/Phone: | Virology & Rabies, 803-896-0819
Days Performed: | Monday - Friday
Turnaround Time: | 15 days
Specimen Required: | Nasopharyngeal swab (NP), nasal aspirate (NA), nasal wash (NW), dual nasopharyngeal/throat swab (NP/TS), bronchoalveolar lavage (BAL), bronchial wash (BW), tracheal aspirate (TA), and sputum (SPT) placed in 2 - 3 mL viral transport media.
Specimen Identification: | Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.
Specimen Volume (optimum): | Swab specimen (see above) placed in 2 - 3 mL viral transport media.
Specimen Volume (minimum): | N/A
Collect: | Screw-capped tube of viral transport media.
Form: | DHEC 1335
Special Instructions: | Year round, the Public Health Laboratory (PHL) participates in the World Health Organization’s (WHO) Influenza Surveillance Program. Collection kits are provided. Please contact the Virology laboratory for more information at (803)896-0819.
| ****If Influenza A/H5N1, A H7, or a newly emerging, highly pathogenic human Influenza strain is suspected, please contact your regional public health office for consultation. Contact information for the regional public health offices is located on the back of the South Carolina List of Reportable Diseases. Upon testing approval, please contact the DHEC PHL at 803-896-0777 or 803-767-8118 for specimen collection, storage and transportation. Testing for A/H5N1, A/H7, and for newly emerging highly pathogenic influenza strains is provided in the Special Pathogens Laboratory.
| Packing and Shipping*: | Send to the attention of Virology & Rabies Laboratory. See [Transporting and Shipping Infectious Substances in Section IV](#).
Transport Conditions: | Store specimens at 2-8°C and ship to maintain temperature at 2-8°C for receipt at the PHL within 72 hours of collection. If specimen transport is delayed and will not be received at the PHL within 72 hours, freeze specimens at ≤ -20°C and ship on dry ice to maintain the temperature of ≤ -20°C until received by the PHL.
Specimen Rejection Criteria: | Specimens received on calcium alginate swabs, cotton swabs, or swabs with wooden shafts, improper temperature; See [Specimen Rejection & Disclaimer Criteria in Section I](#).
Availability: | N/A
Results and Interpretations: | N/A
Additional Information: | Influenza testing also includes a full respiratory viral panel to identify other respiratory viral pathogens.
Purpose of Test: | N/A
Method: | Real-time reverse transcription polymerase chain reaction (real-time RT-PCR)
Interfering Substances: | N/A
Comment: | N/A
<table>
<thead>
<tr>
<th>Test</th>
<th>LEAD ANALYSIS, BLOOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Blood Lead (Blood Pb)</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Analytical Chemistry, 803-896-0886</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As requested</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>1 mL whole blood from venipuncture; 500 µL whole blood from finger stick or heel stick for infant screening. Venipuncture preferred for confirmation of an elevated level.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s full name, and a second patient identifier such as DOB, Specimen #, etc. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>&gt;1 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>500 µL</td>
</tr>
<tr>
<td>Collect:</td>
<td>In general, if more than one evacuated tube of blood is to be drawn from an individual, the blood lead tube should be drawn second or later to avoid cross contamination. Draw the blood through a stainless-steel needle into a Vacutainer™.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332, Test #882</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Specimens should be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until receipt at the PHL. Specimens must be received for testing within 10 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Clotted blood, insufficient quantity (QNS), improper temperature; For universal rejections, See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Blood lead levels in children under the age of 16 are considered elevated at or above 3.5 mg/dL and chelation treatment should be considered at confirmed blood lead levels of 45 mg/dL. The Occupational Safety and Health Administration regulations use a blood lead level of 40 mg/dL as cause for written notification and a medical exam, and a blood lead level of 60 mg/dL as cause for medical removal from exposure. Action levels for blood Pb in children and adults print on result reports. Screening (finger stick/heel stick) levels ≥ 3.5 µg/dL requires venipuncture confirmation.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Identify exposure to Lead.</td>
</tr>
<tr>
<td>Method:</td>
<td>Inductively Coupled Plasma Mass Spectrometry (ICP-MS)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>LEGIONELLA URINARY ANTIGEN TEST</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>---------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Lateral-flow immunoassay for Legionella pneumophila serogroup 1 antigen in human urine specimens.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>3 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>1 mL or &gt; of Urine collected in either airtight transport container or airtight Boric Acid Urine Tube.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 mL</td>
</tr>
<tr>
<td>Collect:</td>
<td>Human Urine specimens, Unpreserved: Specimens should be stored at 2 - 8°C in an airtight transport container to prevent leaking. Specimens must be received within 7 days of collection. Test is available only for outbreaks of Public Health importance as determined by a DHEC Epidemiologist.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition 1335 form, mark Legionella Urine Antigen Test</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Urine is considered an Infectious substance. See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store in refrigerator at 2-8°C and ship with frozen cold packs to maintain temperature at 2-8°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Improper transport media or conditions; improper temperature; For universal rejections, See <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td><strong>Negative Test</strong>: Report test results as “No Legionella pneumophila serogroup 1 antigens detected”. A negative result does not exclude infection with Legionella pneumophila serogroup 1, nor does it rule out other microbial-caused respiratory infections or diseases caused by other serogroups of <em>Legionella pneumophila</em>. <strong>Positive Test</strong>: Report test result as Legionella pneumophila serogroup 1 antigens detected. This result does not rule out co-infection with other pathogens.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Rapid, lateral-flow immunoassay for the qualitative detection of <em>Legionella pneumophila</em> serogroup 1 antigen in human urine specimens. It is designed to test specimens from patients with symptoms of pneumonia. Test results are to be used as an aid in diagnosis of <em>Legionella pneumophila</em> serogroup 1 infection. A negative result does not preclude infection with <em>Legionella pneumophila</em> serogroup 1. Test results are to be used in conjunction with information obtained from the patient’s clinical evaluation and other diagnostic procedures.</td>
</tr>
<tr>
<td>Comment:</td>
<td>Test available only for Division of Acute Disease Epidemiology (DADE).</td>
</tr>
<tr>
<td><strong>Test</strong></td>
<td><strong>LISTERIA SPECIES</strong></td>
</tr>
<tr>
<td>----------</td>
<td>---------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td><em>Listeria monocytogenes</em></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Pure bacterial isolate on an agar slant that will support the growth of the isolate.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Pure isolate subcultured from isolated colonies to a slant that is able to support growth.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition, mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature of specimen within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Culture non-viable; culture mixed; improper temperature; For universal rejections, See <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td><em>Listeria monocytogenes</em> isolated or not isolated.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Submission to PHL is required. Ship within 3 business days.</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>MALARIA ANTIGEN TEST (BINAXNOW)</td>
</tr>
<tr>
<td>-------------------------------------------</td>
<td>--------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td><em>Plasmodium falciparum, Plasmodium vivax, Plasmodium ovale, Plasmodium malariae</em></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>24 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>3 - 5 mL EDTA and thin and thick pre-stained slides - See “Malaria Smear” (below).</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>3 - 5 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>3 mL</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>Form 1335 #522; Malaria; DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (ex. Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Notification of the test request must be made to the Special Pathogens Laboratory prior to testing.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the Special Pathogens Laboratory instructions at (803)896-0777 / (803)767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during consultation.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during consultation.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Test results will be verbally communicated, and a hard copy report will be e-mailed or faxed.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Negative results must be confirmed by thin / thick smear microscopy. Microscopy review is required to identify non-falciparum species and to detect potential mixed infections.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To aid in the rapid diagnosis of human malaria infections and in the differential diagnosis of <em>Plasmodium falciparum</em> (P.f.) infections from other less virulent malarial infections.</td>
</tr>
<tr>
<td>Method:</td>
<td>Immunochromatographic assay</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>MALARIA SMEAR</td>
</tr>
<tr>
<td>----------------------</td>
<td>-------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Giemsa stain, Plasmodium</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td></td>
<td><em>THIS TEST IS REFERRED TO AND PERFORMED BY the CDC.</em></td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>24 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Digital images of stained thick and thin blood smears</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Blood smears: Digital images of 2 sets of smears</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Thick and thin stained blood smears</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Write in “Malarial Smear” at the bottom of the Special Pathogen’s test section. DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Notification of the test request must be made to the Special Pathogens Laboratory, (803)896-0777 / (803)767-8118, prior to shipment.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Please contact the Special Pathogens Laboratory for instructions at (803)896-0777 / (803)767-8118.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Smears made from EDTA blood &gt; 1 hour old; blood smears &gt; 3 days old. For universal rejections, see <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>This test is performed by the CDC through the Special Pathogens Laboratory. The Division of Parasitic Disease (DPDx) at the CDC performs microscopic malarial species confirmation and malaria drug resistance surveillance.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Images are submitted to the CDC for rapid identification.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect and speciate plasmodium species in blood smears</td>
</tr>
<tr>
<td>Method:</td>
<td>Microscopic examination of Giemsa-stained smear</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>MEASLES (RUBEOLA) RNA DETECTION BY REAL-TIME (RT) PCR</td>
</tr>
<tr>
<td>------</td>
<td>-----------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Measles (Rubeola) PCR, RT-PCR, or rRT-PCR</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday, weekend and holiday testing approved on a case-by-case basis</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>3 Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>ONLY throat swabs or nasopharyngeal (NP) swabs will be accepted. Ideally, specimens should be collected within 3 days of symptom onset; however, specimens collected up to 14 days from symptom onset will be accepted. Use swabs with synthetic (polyester, nylon, etc.) tips and aluminum or plastic shafts. DO NOT USE swabs with cotton or calcium alginate tips or wooden shafts. Place the swab in viral transport media for storage and shipment.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Sterile, leak-proof, screw-cap tube containing viral transport media.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>All submissions require prior approval from the Virology section supervisor (803)896-0819, the Microbiology Division Director (803-896-0870), or designee.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2-8°C and ship to be maintained at 2-8°C and received within 72 hours of collection at the PHL. If shipment is delayed and specimen will not be received within 72 hours of collection, store specimen at ≤ -20°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen type other than throat or nasopharyngeal (NP) swabs; Swabs with cotton or calcium alginate tips or wooden shafts; Specimens collected more than 14 days after symptom onset; Specimens shipped without transport media; non-frozen specimens received more than 72 hours after collection. Specimens received at improper temperature.; See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday, weekend and holiday testing approved on a case-by-case basis</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td></td>
</tr>
<tr>
<td>Result</td>
<td>Interpretation</td>
</tr>
<tr>
<td>Detected</td>
<td>Measles RNA detected by RT-PCR</td>
</tr>
<tr>
<td>Not Detected</td>
<td>Unable to detect Measles RNA by RT-PCR</td>
</tr>
<tr>
<td>Inconclusive</td>
<td>Indeterminant: Unable to rule out the presence of Measles RNA</td>
</tr>
<tr>
<td>Unable to detect Human DNA. Results suggest sub-optimal specimen collection, transport, and/or storage conditions.</td>
<td>Recollect specimen</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the presence of Measles (Rubeola) virus nucleic acid (RNA). This test will NOT detect the German Measles (Rubella).</td>
</tr>
<tr>
<td>Method:</td>
<td>Real-time RT-PCR</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>MERCURY IN URINE</td>
</tr>
<tr>
<td>------------------------------</td>
<td>--------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Hg in Urine</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Analytical Chemistry, 803-896-0886</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As requested</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Urine</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s full name, and a second patient identifier such as DOB, Specimen #, etc. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 - 5 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>500 µL</td>
</tr>
<tr>
<td>Collect:</td>
<td>Sterile urine cups</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Urine specimens stored at ≤ -20°C and transported frozen by packing on dry ice to maintain ≤ -20°C temperature until received at the PHL is preferred, when possible. Urine may also be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until receipt at the PHL. Urines stored and shipped at 2-8°C must be received at the PHL within 10 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Insufficient quantity (QNS); improper collection container; improper temperature; For universal rejections, See <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Identify exposure to inorganic (metallic) mercury</td>
</tr>
<tr>
<td>Method:</td>
<td>Inductively Coupled Plasma Mass Spectrometry (ICP-MS)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>MERS (MIDDLE EASTERN RESPIRATORY SYNDROME) NOVEL CORONAVIRUS</td>
</tr>
<tr>
<td>------</td>
<td>------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>MERS</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>24 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Nasopharyngeal and/or oropharyngeal swabs, sputum, lower respiratory aspirate/washes, serum; volume depends on specimen type. Call the Special Pathogens Laboratory for more information.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with the patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 521; check “MERS”</td>
</tr>
<tr>
<td>DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
<td></td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Pre-approval needed - hospitals must obtain approval from the DHEC health department prior to submitting specimens to the Special pathogens Laboratory. Contact information can be located on the back of the List of Reportable Conditions. Contact the Special Pathogens Laboratory (803)896-0777 / (803)767-8118) for test notification, specimen collection, storage conditions, and shipping conditions/methods.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. The definitive identification of MERS virus requires additional testing to be performed by the CDC.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To presumptively detect MERS RNA in clinical specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>CDC/LRN Real Time RT-PCR Assay, EUA</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
</tbody>
</table>
### Test

**MUMPS RNA DETECTION BY REAL-TIME RT PCR**

**Synonym:**  
Mumps PCR, Mumps RT-PCR

**Lab Section/Phone:**  
Virology & Rabies, 803-896-0819

**Days Performed:**  
Monday - Friday, weekend and holiday testing approved on a case-by-case basis.

**Turnaround Time:**  
3 days

**Specimen Required:**  
One buccal swab collected within 14 days of symptom onset. Ideal collections occur within 3 days of symptom onset. Use swabs with polyester or nylon tips and aluminum or plastic shafts. DO NOT USE cotton, wood, or calcium alginate swabs. Place swab in viral transport media for storage and shipment. See [Viral Media Collection for Virology Specimens in Section III, p. III-42](#).

**Specimen Identification:**  
Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.

**Specimen Volume (optimum):**  
N/A

**Specimen Volume (minimum):**  
N/A

**Collect:**  
Buccal swab placed in a sterile, leak-proof, screw-cap tube containing viral transport media.

**Form:**  
DHEC 1335

**Special Instructions:**  
All submissions require prior approval from the Virology section supervisor (803-896-0819), the Microbiology Division Director (803-896-0870), or designee. Only specimens submitted as part of an epidemiological investigation will be accepted.

**Packing and Shipping**:  
See [Transporting and Shipping Infectious Substances in Section IV](#).

**Transport Conditions:**  
Store at 2-8°C and ship to be maintained at 2-8°C and received within 72 hours of collection at the PHL. If shipment is delayed and specimen will not be received within 72 hours of collection, store specimen at ≤-20°C and ship on dry ice to maintain specimen at the temperature of ≤-20°C until received at the PHL.

**Specimen Rejection Criteria:**  
Swabs with cotton or calcium alginate tips or wooden shafts; Specimens collected more than 14 days after symptom onset; Specimens shipped without transport media; non-frozen specimens received more than 72 hours after collection. Specimens received at the improper temperature.; See [Specimen Rejection & Disclaimer Criteria in Section I](#).

**Availability:**  
Monday - Friday, weekend and holiday testing approved on a case-by-case basis

**Results and Interpretations:**

<table>
<thead>
<tr>
<th>Result</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Detected</td>
<td>Mumps RNA detected by RT-PCR</td>
</tr>
<tr>
<td>Not Detected</td>
<td>Unable to detect Mumps RNA by RT-PCR</td>
</tr>
<tr>
<td>Inconclusive</td>
<td>Indeterminant: Unable to rule out the presence of Mumps RNA</td>
</tr>
<tr>
<td></td>
<td>Unable to detect Human DNA. Results suggest sub-optimal specimen collection, transport, or storage conditions.</td>
</tr>
<tr>
<td></td>
<td>Recollect specimen</td>
</tr>
</tbody>
</table>

**Additional Information:**  
Only specimens submitted as part of an epidemiological investigation will be accepted.

**Purpose of Test:**  
To detect the presence of Mumps virus nucleic acid (RNA).

**Method:**  
Real-time reverse transcriptase polymerase chain reaction.

**Interfering Substances:**  
N/A

**Comment:**  
N/A
<table>
<thead>
<tr>
<th>Test</th>
<th>MUMPS VIRUS SEROLOGY IgG and IgM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Parotitis Epidemica antibodies</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Per request</td>
</tr>
</tbody>
</table>
| Turnaround Time: | IgG: 10 days  
IgM: 5 days |
| Specimen Required: | Serum |
| Specimen Identification: | Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition. |
| Specimen Volume (optimum): | 2 mL serum |
| Specimen Volume (minimum): | 1 mL serum |
| Collect: | Serum Separator vacuum tube (SST) centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged and serum is removed from the clot and put into a labeled secondary container/tube.) Please follow manufacturer’s guidelines. See [*Specimen Collection: Venipuncture Procedure in Section III*](#), if needed. |
| Form: | DHEC 1332 |
| Special Instructions: | None |
| Packing and Shipping*: | See [*Transporting and Shipping Infectious Substances in Section IV*](#). |
| Transport Conditions: | Store at 2-8°C and ship within 36 hours of collection to maintain specimen at 2-8°C until received by the PHL. If shipment is delayed longer than 36 hours, store specimen at ≤ -20°C and ship on dry ice to maintain the temperature of ≤ -20°C until received by the PHL. |
| Specimen Rejection Criteria: | See [*Specimen Rejection & Disclaimer Criteria in Section I*](#). |
| Availability: | Mumps IgG once/week; Mumps IgM as needed. |
| Results and Interpretations: | Mumps IgG immune status reported as positive, negative, or equivocal.  
Mumps IgM reported as positive or negative. |
| Additional Information: | Results | Interpretations |
| Mumps IgG | Positive | IgG antibodies to the Mumps virus were detected. A positive test indicates a current or past infection, or prior vaccination against the Mumps virus. |
| | Negative | Indicates no detectable IgG antibodies to the Mumps virus. A non-reactive result indicates no current or previous infection with the Mumps virus. Such patients are presumed to be non-immune and are therefore susceptible to a primary infection. A non-reactive result may be obtained early in seroconversion of infected individuals. If this is suspected, obtain an additional specimen in 3 - 5 weeks for re-testing. |
| | Equivocal | Re-evaluate by collecting and testing another specimen after 14 days. |
| Mumps IgM | Positive | Indicates an acute infection. |
| | Negative | Indicates no detectable IgM antibody to the Mumps virus. |
| Purpose of Test: | Mumps IgG: To detect Mumps IgG antibodies for determining immune status.  
Mumps IgM: To detect Mumps IgM antibodies for diagnosing a current infection. |
<p>| Method: | EIA for Mumps IgG; IFA for Mumps IgM. |
| Interfering Substances: | N/A |
| Comment: | N/A |</p>
<table>
<thead>
<tr>
<th>Test</th>
<th>MYCOBACTERIAL CULTURE, BLOOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>TB, AFB</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Mycobacteriology (TB), 803-896-0828</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday-Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>56 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>1 - 5 mL whole blood; with optimum recovery obtained at 3 mL to 5 mL</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen must be labeled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>The range of blood volume which can be cultured is 1 mL to 5 mL, with optimum recovery obtained at 3 mL to 5 mL.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>The range of blood volume which can be cultured is 1 mL to 5 mL, with optimum recovery obtained at 3 mL to 5 mL.</td>
</tr>
<tr>
<td>Collect:</td>
<td>1 - 5 mL whole blood in BD BACTEC Myco/F Lytic Culture Vials</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>The specimen must be collected using sterile techniques to reduce the chance of contamination.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Section IV, Transporting and Shipping Infectious Substances.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Incubate at 37°C if shipping is delayed over 24 hours. Ship in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Detection of mycobacteria in blood.</td>
</tr>
<tr>
<td>Method:</td>
<td>BACTEC FX40 system, bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>Other aerobic organisms including bacteria may, if present, interfere with the recovery of slower growing mycobacteria.</td>
</tr>
<tr>
<td>Comment:</td>
<td>Organisms identified as M. tuberculosis complex referred by PHL for drug susceptibility testing, as indicated.</td>
</tr>
<tr>
<td>Test</td>
<td>MYCOBACTERIAL CULTURE, Other than Blood</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-----------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>AFB, TB</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Mycobacteriology (TB), 803-896-0828</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>56 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Sputum, body fluids, tissue</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen must be labeled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>5 - 10 mL sputum, and other body fluids; 10 mL urine or gastric washings, walnut sized portion of feces or 10 mL liquid stool. See Mycobacterium Culture Collection Procedure.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Screw cap 50 mL polypropylene conical tube</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, Test # 601</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store specimens at 2-30°C and ship specimens to be maintained at 2-30°C until received at the PHL within 3 days of collection. If there is a delay of more than 3 days between collection and shipping, store specimens refrigerated at 2-8°C and ship on frozen cold packs to maintain at 2-8°C until received by the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen &gt; 5 days old when received (Sputum and Urine). Specimens received at the improper temperature.; For universal rejections, see Specimen Rejection and Disclaimer Criteria in Section I, p. 1 - 5.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Detection of Mycobacteria in clinical specimens.</td>
</tr>
<tr>
<td>Method:</td>
<td>Conventional culture methods, GeneXpert MTB/RIF for rapid identification of Mycobacterium tuberculosis DNA and resistance to rifampicin (sputum specimens only), bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Organisms identified as <em>M. tuberculosis complex</em> referred by PHL for drug susceptibility testing, as indicated.</td>
</tr>
<tr>
<td>Test</td>
<td>MYCOBACTERIAL CULTURE, REFERRED FOR IDENTIFICATION</td>
</tr>
<tr>
<td>------</td>
<td>---------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>AFB, TB</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Mycobacteriology (TB), 803-896-0828</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 week</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Send only pure culture with sufficient growth to perform test</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labeled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Sufficient growth to perform test</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Sufficient growth to perform test</td>
</tr>
<tr>
<td>Collect:</td>
<td>Pure culture; LJ slant preferred</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335, Test #602</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Send only pure culture with sufficient growth to perform test</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Section IV, Transporting and Shipping Infectious Substances.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>For universal rejections, see Section I Specimen Rejection and Disclaimer Criteria, p. I - 5</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Identification of Mycobacterium from culture.</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Organisms identified as <em>M. tuberculosis complex</em> referred by PHL for drug susceptibility testing, as indicated.</td>
</tr>
<tr>
<td>Test</td>
<td>NEISSERIA MENINGITIDIS</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>----------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Bacterial meningitis</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Pure bacterial isolate on an agar slant that will support the growth of the isolate (Chocolate agar slant is preferred).</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate container must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Submit well isolated colonies subbed to a slant that will support the growth, incubate overnight in CO₂.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition, mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a></td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store in a 35°C CO₂ incubator and ship in an approved shipper to maintain specimen at 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Culture non-viable; culture mixed; improper temperature; For universal rejections, see <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Isolate will be confirmed and serogrouped.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Submit all <em>N. meningitidis</em> isolated from normally sterile sites within 1 business day.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Confirmation of identification and serogroup</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS, Serogroup</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>NEWBORN SCREENING PANEL</td>
</tr>
<tr>
<td>----------------------</td>
<td>----------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>N/A; Panel includes screening for:</td>
</tr>
<tr>
<td></td>
<td>• Amino Acid Disorders</td>
</tr>
<tr>
<td></td>
<td>• Organic Acid Conditions</td>
</tr>
<tr>
<td></td>
<td>• Fatty Acid Disorders</td>
</tr>
<tr>
<td></td>
<td>• Biotinidase Deficiency</td>
</tr>
<tr>
<td></td>
<td>• Classic Galactosemia</td>
</tr>
<tr>
<td></td>
<td>• Cystic Fibrosis</td>
</tr>
<tr>
<td></td>
<td>• Certain Hemoglobinopathies</td>
</tr>
<tr>
<td></td>
<td>• Primary Congenital Hypothyroidism</td>
</tr>
<tr>
<td></td>
<td>• Congenital Adrenal Hyperplasia</td>
</tr>
<tr>
<td></td>
<td>• Severe Combined Immunodeficiencies</td>
</tr>
<tr>
<td></td>
<td>• Pompe Disease</td>
</tr>
<tr>
<td></td>
<td>• Mucopolysaccharidosis Type I (MPS-I)</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Newborn Screening/ 803-896-0874 or 803-896-0891</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Saturday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>4 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Dried blood spot collected on DHEC 1327 collection form</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient's full name and date of birth written on DHEC 1327 collection form.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>All 5 circles filled</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Varies depending on how full each circle is, how well the blood saturates the paper, and if any repeat testing is needed</td>
</tr>
<tr>
<td>Collect:</td>
<td>Heelstick; See Specimen Collection: Heel-Stick Procedure for Patients Less than 1 Year Old, Section III, p. III-29.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1327</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Allow the specimen to dry horizontally for at least 3 to 4 hours prior to packing; fold over Biohazard labeled flap once specimen is dry; don’t use capillary tubes for collection; don’t layer blood spots.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Place dried specimens in paper envelope/cardboard mailer (no plastic).</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Do NOT use plastic bags or any other airtight, leakproof, or sealed containers.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimens received in plastic bags; specimens collected on expired collection forms; specimens older than 14 days; patient older than 1 year; specimen quality or quantity inadequate/insufficient</td>
</tr>
<tr>
<td>Availability:</td>
<td>N/A</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Identifies newborns that may be at an increased risk of having a certain serious condition</td>
</tr>
<tr>
<td>Method:</td>
<td>• Tandem Mass Spectrometry: Amino Acid Disorders, Organic Acid Conditions, Fatty Acid Disorders</td>
</tr>
<tr>
<td></td>
<td>• Enzymatic &amp; Fluorescence: Biotinidase Deficiency, Classic Galactosemia</td>
</tr>
<tr>
<td></td>
<td>• Fluorimmuno assay and/or PCR: Cystic Fibrosis</td>
</tr>
<tr>
<td></td>
<td>• High Performance Liquid Chromatography and/or Iso-electric focusing: Certain Hemoglobinopathies</td>
</tr>
<tr>
<td></td>
<td>• Fluorimmuno assay: Primary Congenital Hypothyroidism, Congenital Adrenal Hyperplasia</td>
</tr>
<tr>
<td></td>
<td>• PCR: Severe Combined Immunodeficiencies</td>
</tr>
<tr>
<td></td>
<td>• Flow Injection Analysis Tandem Mass Spectrometry: Pompe and MPS-I</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>NOROVIRUS DETECTION BY REAL TIME RT PCR</td>
</tr>
<tr>
<td>------</td>
<td>---------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Norwalk Virus, Norovirus PCR, GI Outbreak</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>A peanut-sized or tablespoon volume of fresh diarrheal stool. Specimens collected within 48 - 72 hours of symptom onset are preferred. Specimens collected within 10 days of symptom onset will be accepted. Rectal swabs are not acceptable.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>A peanut-sized or tablespoon volume of fresh diarrheal stool.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Stool in a sterile, screw capped, leak-proof, 50 mL conical tube or urine container.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2 - 8°C and ship to maintain specimen at 2 - 8°C until received by the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimens placed in any type of media; Specimen more than 10 days old when received. Specimens received at the improper temperature.; See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday; Availability of this test is restricted to epidemiological investigations.</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Results are reported as negative or positive for the presence of genogroup I or genogroup II Norovirus.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the presence of Norovirus nucleic acid (RNA).</td>
</tr>
<tr>
<td>Method:</td>
<td>Real-time reverse transcriptase polymerase chain reaction (real-time RT-PCR)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>ORGANISM for IDENTIFICATION</td>
</tr>
<tr>
<td>---------------------------</td>
<td>----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Any bacterial isolates required to be submitted per the South Carolina List of Reportable Conditions.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Pure bacterial isolates subbed from an isolated colony to an agar slant that will permit growth of the organism.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Pure culture of isolate, subbed from an isolated colony to an agar slant that will permit growth of the organism.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition, mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship according to directions listed under specific organism.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Mixed isolate. Specimens received at the improper temperature.; For universal rejections, See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday unless otherwise noted for specific organism.</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Organism identification confirmed or not. Serotyping and serogrouping as needed.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMeriuxex VITEK MS, Conventional methods, biochemicals, serotyping/grouping</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
</tbody>
</table>
## Test

<table>
<thead>
<tr>
<th><strong>Test</strong></th>
<th><strong>QuantiFERON-TB Gold Plus (QFT Plus)</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Synonym:</strong></td>
<td>QFT, Interferon-Gamma Release Assay (IGRA)</td>
</tr>
<tr>
<td><strong>Lab Section/Phone:</strong></td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td><strong>Days Performed:</strong></td>
<td>Monday-Friday; weekend testing available with prior approval by Supervisor or Division Director.</td>
</tr>
<tr>
<td><strong>Turnaround Time:</strong></td>
<td>7 days</td>
</tr>
<tr>
<td><strong>Specimen Required:</strong></td>
<td>Whole blood in 4 QFT-Plus blood collection tubes</td>
</tr>
<tr>
<td><strong>Specimen Identification:</strong></td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td><strong>Specimen Volume (optimum):</strong></td>
<td>1 mL whole blood</td>
</tr>
<tr>
<td><strong>Specimen Volume (minimum):</strong></td>
<td>0.8 mL – 1.2 mL, as indicated on tube labels with 2 black fill lines</td>
</tr>
<tr>
<td><strong>Collect:</strong></td>
<td>4 QuantiFERON-TB Gold Plus tubes:</td>
</tr>
<tr>
<td></td>
<td>• Nil antigen (Grey cap)</td>
</tr>
<tr>
<td></td>
<td>• TB 1 antigen (Green cap)</td>
</tr>
<tr>
<td></td>
<td>• TB 2 antigen (Yellow cap)</td>
</tr>
<tr>
<td></td>
<td>• Mitogen (Purple cap)</td>
</tr>
<tr>
<td></td>
<td>Specific collection requirements are needed. For detailed collection procedure, see QuantiFERON-TB Gold Plus (QFT-Plus) Collection Procedure in Section III p. III-33.</td>
</tr>
<tr>
<td><strong>Form:</strong></td>
<td>DHEC 1335; be sure to write the incubation start and end times on this form.</td>
</tr>
<tr>
<td><strong>Special Instructions:</strong></td>
<td>QFT-Plus Blood Collection Tubes should be at 17-25°C at time of blood collection.</td>
</tr>
<tr>
<td><strong>Packing and Shipping</strong>:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td><strong>Transport Conditions:</strong></td>
<td>Store tubes at 17-27°C prior to and after incubation. Specimens should be shipped and received within 3 days post-incubation, or within 16 hours of collection if NOT incubated in the regions. Place the specimen inside designated QFT-Plus shipper (large white shipper with pink label) labeled to the attention of Virology and ship to maintain tubes in the temperature range of 17-27°C until receipt at the PHL.</td>
</tr>
<tr>
<td><strong>Specimen Rejection Criteria:</strong></td>
<td>Specimens with volumes below 0.8 mL or above 1.2 mL, as indicated by the black fill lines on tube labels; Specimens not incubated within the proper incubation period; Specimens requiring incubation at 37°C that are not received by the PHL within 16 hours of collection. Specimens received at the improper temperature.; See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td><strong>Availability:</strong></td>
<td>Monday - Friday</td>
</tr>
<tr>
<td><strong>Result</strong></td>
<td><strong>Interpretation</strong></td>
</tr>
<tr>
<td>Positive</td>
<td><em>M. tuberculosis</em> infection likely</td>
</tr>
<tr>
<td>Negative</td>
<td><em>M. tuberculosis</em> infection not likely</td>
</tr>
<tr>
<td>Indeterminate</td>
<td>Likelihood of <em>M. tuberculosis</em> infection cannot be determined</td>
</tr>
<tr>
<td><strong>Additional Information:</strong></td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Purpose of Test:</strong></td>
<td>Aids in the diagnosis of <em>Mycobacterium tuberculosis</em> (TB) infection</td>
</tr>
<tr>
<td><strong>Method:</strong></td>
<td>Detection of interferon-γ by ELISA</td>
</tr>
<tr>
<td><strong>Interfering Substances:</strong></td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Comment:</strong></td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>RABIES EXAMINATION</td>
</tr>
<tr>
<td>------</td>
<td>--------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>N/A</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday only. Weekends and holidays only with notification and emergency testing criteria being met, specifically: (a) An unprovoked wild animal bite to a human, such as bites from a raccoon, fox, skunk, bobcat, coyote, etc.; or (b) A bat when there is an obvious bat bite, or if individuals awaken and find a bat in their room, or if there is a bat in a room with an unattended child or near a mentally impaired or intoxicated person.</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>24 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Brain tissue</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Whole animal head</td>
</tr>
<tr>
<td>Collect:</td>
<td>Ship whole animal head. Heads are only submitted by DHEC Rabies Control Staff.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1308</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Contact the local county health department for the information on specimen collection and shipping instructions. Confirmation is a postmortem procedure; because the standard procedure currently requires the examination of brain tissue, the suspect animal must either be sacrificed or have died before the examination can be performed. All county health departments maintain containers appropriate for shipping specimens for examination, information on the management of animals suspected of being rabid, and information to obtain vaccine for persons exposed to a rabid animal after consultation with the state epidemiologist.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2-8°C and ship on frozen cold packs to maintain the temperature at 2-8°C until receipt at the PHL. See Special Instructions above.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>No brain tissue or tissue decomposed or grossly contaminated. See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>See Days Performed above.</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reported as Positive or Negative. All Positive reports are called directly to the county health department, or after regular working hours, to the county environmentalist who submitted the specimen.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect the rabies viral antigen in brain tissue of suspected animals, for the protection of persons exposed.</td>
</tr>
<tr>
<td>Method:</td>
<td>Fluorescent Antibody (FA)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>The PHL is the only laboratory in SC that performs testing for rabies in animals. Human testing is only performed at CDC with prior approval. Call Virology &amp; Rabies before sending to obtain proper documentation, 803-896-0819.</td>
</tr>
<tr>
<td>Test</td>
<td>RESPIRATORY PANEL 2 by FilmArray (PCR)</td>
</tr>
<tr>
<td>------</td>
<td>--------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Adenovirus (AdV); Coronavirus (CoV) 229E, HKU1, NL63, OC43; Enterovirus (EV); c Human Rhinovirus (RHV); Human Metapneumovirus (hMPV); Influenza A (Flu A) (subtypes H1, H1-2009, and H3); Influenza B (Flu B); Parainfluenza Virus 1 (PIV1); Parainfluenza Virus 2 (PIV2); Parainfluenza Virus 3 (PIV3); Parainfluenza Virus 4 (PIV4); Respiratory Syncytial Virus (RSV); Bordetella pertussis; Bordetella parapertussis; Chlamydophila pneumoniae; and Mycoplasma pneumoniae</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday, only with prior approval as part of a respiratory outbreak investigation</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>5 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Nasopharyngeal (NP) swab placed in viral transport media</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1 - 3 mL of viral transport media containing a nasopharyngeal (NP) swab</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Nasopharyngeal (NP) swab placed immediately into sterile tubes containing 2 - 3 mL of viral transport media. Use only synthetic fiber swabs with plastic shafts. Do not use calcium alginate swabs or swabs with wooden shafts, as they may contain substances that inactivate some viruses and inhibit PCR testing. See Viral Media Collection for Virology Specimens in Section III, p. III-42.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Request BioFire FilmArray RP2 Panel</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Call Virology at 803-896-0819</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2-8°C and ship on frozen cold packs to maintain the temperature at 2-8°C until receipt at the PHL. If shipping is delayed more than 48 hours, freeze at ≤-15°C and ship on dry ice to maintain specimen at temperature of ≤-15°C until received at the PHL. Specimen frozen at ≤-15°C must be received at the PHL within 30 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Specimen type other than nasopharyngeal (NP) swab; Use of calcium alginate swabs or swabs with wooden shafts; Specimens received at the improper temperature; See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>For outbreaks as determined by the SC DHEC Division of Acute Disease Epidemiology</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To identify Adenovirus (AdV); Coronavirus (CoV) 229E, HKU1, NL63, OC43; Enterovirus (EV); Human Rhinovirus (RHV); Human Metapneumovirus (hMPV); Influenza A (Flu A) (subtypes H1, H1-2009, and H3); Influenza B (Flu B); Parainfluenza Virus 1 (PIV1); Parainfluenza Virus 2 (PIV2); Parainfluenza Virus 3 (PIV3); Parainfluenza Virus 4 (PIV4); Respiratory Syncytial Virus (RSV); Bordetella pertussis; Bordetella parapertussis; Chlamydophila pneumoniae; and Mycoplasma pneumoniae.</td>
</tr>
<tr>
<td>Method:</td>
<td>Multiplex Real-time PCR</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
</tbody>
</table>
**Test** | **RPR (RAPID PLASMA REAGIN)**  
--- | ---  
Synonym: | RPR, Non-Treponemal Antibody  
Lab Section/Phone: | Diagnostic Serology, 803-896-0811  
Days Performed: | Monday - Friday  
Turnaround Time: | 1 - 5 Business Days  
Specimen Required: | Serum  
Specimen Identification: | Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.  
Specimen Volume (optimum): | 1.5 mL of serum  
Specimen Volume (minimum): | 1.0 mL of serum  
Collect: | Serum-separator tube or serum. Tubes must be properly centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See **Specimen Collection: Venipuncture Procedure in Section III**, if needed.  
Form: | DHEC 1332 Test #001, 002, 004 or Test #235 (All specimens submitted to the PHL will undergo the reverse-algorithm unless otherwise indicated). Special requests should be in writing on the form under special instructions.  
Special Instructions: | N/A  
Packing and Shipping*: | See **Transporting and Shipping Infectious Substances in Section IV**.  
Transport Conditions: | Store at 2-8°C and ship to be maintained at 2-8°C and received within 5 days of collection at the PHL; for storage longer than 5 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.  
Specimen Rejection Criteria: | Plasma specimen; received after 5 days not maintained at -20°C or colder; received at the improper temperature; For universal rejections, see **Specimen Rejection & Disclaimer Criteria in Section I**.  
Availability: | Monday - Friday  
Results and Interpretations: | N/A  
Additional Information: | Reflex test for reactive and equivocal Syphilis TP’s; Quantitation performed on RPR reactive specimens.  
Purpose of Test: | To aid in the detection, diagnosis, and staging of syphilis  
Method: | Charcoal flocculation  
Interfering Substances: | N/A  
Comment: | N/A
<table>
<thead>
<tr>
<th><strong>Test</strong></th>
<th><strong>RUBELLA SEROLOGY - IgG and IgM</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>German Measles antibody, Rubella immune screen, Rubella IgG and IgM</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>IgG: 10 days  IgM: N/A</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 mL serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator vacuum tube (SST), centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged and serum is removed from the clot and put into a labeled secondary container/tube.) Please follow manufacturer’s guidelines. See <strong>Specimen Collection: Venipuncture Procedure in Section III</strong>, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td><strong>Call Virology, 803-896-0819, prior to sending specimens for IgM.</strong> Rubella IgG does not require notification.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <strong>Transporting and Shipping Infectious Substances in Section IV.</strong></td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2 - 8°C and ship within 36 hours of collection to maintain specimen at 2 - 8°C until received by the PHL. If shipment is delayed longer than 36 hours, specimen should be stored at ≤-20°C and shipped on dry ice to maintain the temperature of ≤-20°C until received by the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See <strong>Specimen Rejection &amp; Disclaimer Criteria in Section I.</strong></td>
</tr>
<tr>
<td>Availability:</td>
<td>IgG: As requested  IgM: Referred to CDC</td>
</tr>
</tbody>
</table>
| Results and Interpretations: | **Rubella IgG**  
<p>| <strong>Result</strong> | <strong>Interpretation</strong> |
| Positive | Indicates a current or previous infection with Rubella virus, or prior vaccination against Rubella virus. |
| Equivocal | Collect and test another specimen. |
| Negative | No detectable IgG antibodies to the Rubella virus. A non-reactive result indicates no current or previous infection with Rubella virus. Such patients are presumed to be susceptible to a primary infection. However, specimens taken too early during a primary infection may not have detectable levels of IgG antibody. If this is suspected, collect and test another specimen in 8 - 14 days. |
| Additional Information: | N/A |
| Purpose of Test: | IgM: Used in diagnosis of German Measles and during possible outbreaks. IgM antibodies usually appear 3 - 5 days after onset of rash. IgG: Used to determine immune status of patient. |
| Method: | EIA (Enzyme Immunoassay) |
| Interfering Substances: | N/A |
| Comment: | N/A |</p>
<table>
<thead>
<tr>
<th>Test</th>
<th>RUBEOLA (Measles) VIRUS SEROLOGY- IMMUNE STATUS/DIAGNOSTIC</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Measles Serology IgM and IgG</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>IgG: 10 days IgM: Referred to CDC</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 mL serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator vacuum tube (SST), centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged and serum is removed from the clot and put into a labeled secondary container/tube. different container/tube). Please follow manufacturer’s guidelines. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Call Virology, 803-896-0819, prior to sending specimen for IgM. Rubeola IgG does not require notification.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2 - 8°C and ship within 36 hours of collection to maintain specimen at 2 - 8°C until received by the PHL. If shipment is delayed longer than 36 hours, store specimen at ≤ -20°C and ship on dry ice to maintain at temperature of ≤ -20°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>IgG: As requested IgM: Referred to CDC</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Measles IgG</td>
</tr>
<tr>
<td></td>
<td>Result</td>
</tr>
<tr>
<td></td>
<td>Positive</td>
</tr>
<tr>
<td></td>
<td>Equivocal</td>
</tr>
<tr>
<td></td>
<td>Negative</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>IgG: Used to determine immune status of the patient. IgM: Used in diagnosis of measles and during possible outbreaks. IgM antibodies usually appear 3 - 5 days after onset of rash.</td>
</tr>
<tr>
<td>Method:</td>
<td>EIA (Enzyme Immunoassay)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>STAPHYLOCOCCUS</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Enteric Pathogen Culture, <em>Staphylococcus aureus</em>, for VISA/VRSA confirmation, see <em>Staphylococcus</em> (VISA/VRSA) isolates.</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical specimens and isolates – Clinical Microbiology, 803-896-0805 Food Samples – Food Microbiology, 803-896-0872 MRSA/VRSA isolates from suspected outbreaks – Molecular Microbiology, 803-896-0826</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Swabs – transport in media that will support the growth of the organism. Referred Isolate – transport on an agar slant that will support growth Food – call the food microbiology laboratory before shipping food samples (803-896-0872)</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container and Isolates must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen#. Specimen container or isolates should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC requisition 1335</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Packing and Shipping Instructions in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL. Ship swabs in transport media on frozen cold packs to be maintained in temperature range of 2-8°C until receipt at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Culture non-viable; culture mixed; improper temperature; For universal rejections, see <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>Conventional culture methods and biochemical analysis. bioMerieux VITEK MS, WGS for outbreak investigations.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>STAPHYLOCOCCUS (VISA/VRSA) ISOLATES</td>
</tr>
<tr>
<td>------</td>
<td>-------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Vancomycin Intermediate Staphylococcus aureus, Vancomycin Resistant Staphylococcus aureus <strong>Staphylococcus aureus, vancomycin-resistant or intermediate with a VA &gt; 6 MIC.</strong></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Pure, low passage isolate on a non-inhibitory, non-selective agar plate or slant that will support the growth of the isolate. Include both isolated colony and at least one original culture plate as resistance can be lost over time and subbing out organism.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Be sure to submit a pure bacterial isolate, subbed from an isolated colony.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition, Mark Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td><strong>According to CDC and CLSI, only isolates with a commercial instrument MIC or E-test &gt; 6 need sent to a reference laboratory for confirmation. CDC states results from Vitek 2, MicroScan, Phoenix, or E-test are accurate and correlate with studies performed at the CDC. MIC values of 2, 3, and 4 are not uncommon.</strong></td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <strong>Packing and Shipping Instructions in Section IV.</strong></td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Culture non-viable, culture mixed, specimens received at the improper temperature; For universal rejections, see <strong>Specimen Rejection &amp; Disclaimer Criteria in Section I.</strong></td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. Ship within 1 business day.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS, E-test</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>STREPTOCOCCUS (BETA HEMOLYTIC GROUP A)</td>
</tr>
<tr>
<td>------</td>
<td>--------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Group A Strep, <em>Streptococcus pyogenes</em></td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0803</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate on agar slant able to promote growth</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition form, mark Organism for ID, write under special instructions, “Freeze organism”.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Submit Group A Beta hemolytic <em>Streptococcus</em> (<em>S. Pyrogenes</em>) organisms that are of epidemiologic concern, to be frozen for possible surveillance studies at a later date.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Packing and Shipping Instructions in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>All Group A Strep submitted will be logged and frozen on freezer beads for possible epidemiological surveillance at a later date.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMereieux VITEK MS, freezer beads</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Test</strong></td>
<td><strong>STREPTOCOCCUS PNEUMONIAE</strong></td>
</tr>
<tr>
<td>--------------------------</td>
<td>------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Strep pneumo, invasive (pneumococcal)</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Pure isolate on a Chocolate or Blood agar slant</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Submit <em>S. pneumoniae</em> isolate from patients of any age, ALL CSF isolates, and invasive sterile body sites that are non-susceptible to any relevant antibiotics according to CLSI for further testing and serotyping.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition mark, Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Invasive disease = isolated from normally sterile site. Always specify site of isolate.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Packing and Shipping Instructions in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store in 35°C CO₂ incubator and ship in approved shippers which will maintain temperature within the range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>For universal rejections, See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the PHL is required for <em>Streptococcus pneumoniae</em>, isolate from patients of any age, ALL CSF isolates, and invasive sterile body sites that are non-susceptible to any relevant antibiotics according to CLSI for further testing and serotyping. Shipped to Wisconsin State Laboratory of Hygiene (WSLH) for serotyping by PCR.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Submission required for epidemiologic surveillance.</td>
</tr>
<tr>
<td>Method:</td>
<td>PCR</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Test</strong></td>
<td><strong>SYPHILIS SEROLOGY SCREEN</strong></td>
</tr>
<tr>
<td>----------</td>
<td>----------------------------</td>
</tr>
<tr>
<td><strong>Synonym:</strong></td>
<td>Syphilis Total Antibodies, Reverse-Algorithm, Treponemal Antibodies</td>
</tr>
<tr>
<td><strong>Lab Section/Phone:</strong></td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td><strong>Days Performed:</strong></td>
<td>Monday – Friday</td>
</tr>
<tr>
<td><strong>Turnaround Time:</strong></td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td><strong>Specimen Required:</strong></td>
<td>Serum</td>
</tr>
<tr>
<td><strong>Specimen Identification:</strong></td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td><strong>Specimen Volume (optimum):</strong></td>
<td>1.5 mL of serum</td>
</tr>
<tr>
<td><strong>Specimen Volume (minimum):</strong></td>
<td>1.0 mL of serum</td>
</tr>
<tr>
<td><strong>Collect:</strong></td>
<td>Serum-separator tube or serum. Tubes must be properly centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See <a href="#">Specimen Collection: Venipuncture Procedure in Section III</a>, if needed.</td>
</tr>
<tr>
<td><strong>Form:</strong></td>
<td>DHEC 1332 (All specimens submitted to the PHL for syphilis testing will undergo the reverse-algorithm unless otherwise indicated.)</td>
</tr>
<tr>
<td><strong>Special Instructions:</strong></td>
<td>N/A</td>
</tr>
<tr>
<td><strong>Packing and Shipping</strong>:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td><strong>Transport Conditions:</strong></td>
<td>Store and ship to be maintained at 2-8°C and received within 5 days of collection at the PHL; for storage longer than 5 days, remove the serum from the clot or gel, place in a labelled secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td><strong>Specimen Rejection Criteria:</strong></td>
<td>Plasma specimen; received after 5 days not maintained at -20°C or colder; specimens received at the improper temperature; For universal rejections, see <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td><strong>Availability:</strong></td>
<td>Monday – Friday</td>
</tr>
<tr>
<td><strong>Results and Interpretations:</strong></td>
<td>Reactive, Nonreactive, or Equivocal</td>
</tr>
<tr>
<td><strong>Additional Information:</strong></td>
<td>Reactive and Equivocal Syphilis TP specimens will automatically be reflexed for RPR testing. If the RPR is nonreactive, the specimen(s) will be automatically reflexed for manual TP-PA testing.</td>
</tr>
<tr>
<td><strong>Purpose of Test:</strong></td>
<td>The qualitative detection of antibodies (IgG and IgM) directed against Treponema pallidum (TP) in human serum.</td>
</tr>
<tr>
<td><strong>Method:</strong></td>
<td>Multiplex flow immunoassay</td>
</tr>
<tr>
<td><strong>Interfering Substances:</strong></td>
<td>Assay interference due to circulating antibodies against yaws, pinta, and bejel has not been evaluated. Cross-reactivity with these treponemal disease conditions is to be expected.</td>
</tr>
<tr>
<td><strong>Comment:</strong></td>
<td>RPR automatically performed on positives and equivocals. Specimen requirements allow for reflexed testing to be performed, if needed.</td>
</tr>
<tr>
<td>Test</td>
<td>TP-PA SEROLOGY</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>----------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>MHA-TP, Treponemal Antibody Serology</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>1 mL of serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>0.5 mL of serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum-separator tube or serum. Tubes must be properly centrifuged, and serum from red top tubes must be removed from the clot and put into a labeled secondary container/tube. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store and ship to be maintained at 2-8°C and received within 5 days of collection at the PHL; for storage longer than 5 days, remove the serum from the clot or gel, place in a secondary container and freeze at -20°C or colder, and ship on dry ice to maintain specimen at temperature of -20°C or colder until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Plasma specimen; received after 5 days not maintained at -20°C or colder; Grossly contaminated, grossly lipemic, excessively hemolyzed, or chylous; Specimens received at the improper temperature. For universal rejections, See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Reactive, Nonreactive, or Indeterminate</td>
</tr>
<tr>
<td></td>
<td>Not a screening test; Reactive test is usually reactive for life (85% of cases). Specimens are reflexed for TP-PA testing only if the initial Syphilis TP is reactive or equivocal and the RPR is non-reactive.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>N/A</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>An aid to resolve discrepant results between screening treponemal (Syphilis TP) and nontreponemal (RPR) test results.</td>
</tr>
<tr>
<td>Method:</td>
<td>Particle Agglutination</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>TRACE HEAVY METALS IN URINE</td>
</tr>
<tr>
<td>------</td>
<td>-----------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Urine Metals</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Analytical Chemistry, 803-896-0886</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As requested</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Urine</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container must be labelled with patient’s full name, and a second patient identifier such as DOB, Specimen #, etc. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 - 5 mL</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>500 µL</td>
</tr>
<tr>
<td>Collect:</td>
<td>Sterile urine cups</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332, Test #885</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Urine specimens stored at ≤ -20°C and transported frozen by packing on dry ice to maintain ≤ -20°C temperature until received at the PHL is preferred, when possible. Urine may also be stored at 2-8°C and shipped on frozen cold packs to maintain specimens at 2-8°C until receipt at the PHL. Urines stored and shipped at 2-8°C must be received at the PHL within 10 days of collection.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Insufficient quantity (QNS); improper collection container; specimens received at the improper temperature; For universal rejections, see Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday – Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Metals included: Arsenic (As), Barium (Ba), Beryllium (Be), Cadmium (Cd), Lead (Pb), Thallium (Tl), Uranium (U)</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>Identify exposure to As, Ba, Be, Cd, Pb, Tl, and U</td>
</tr>
<tr>
<td>Method:</td>
<td>Inductively Coupled Plasma Mass Spectrometry (ICP-MS)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>TRICHOMONAS VAGINALIS DETECTION BY NUCLEIC ACID AMPLIFICATION</td>
</tr>
<tr>
<td>----------------------------------------------------------------------</td>
<td>---------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Hologic Trichomonas vaginalis Amplified Nucleic Acid Test (NAAT), Trichomonas vaginalis rRNA, Aptima TV</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Diagnostic Serology, 803-896-0811</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>1 - 5 Business Days</td>
</tr>
</tbody>
</table>
| Specimen Required:                                                 | • The assay may be used to test the following specimens from symptomatic or asymptomatic female patients:  
  · Unisex swab (blue shafted swab) for endocervical specimens  
  · MTS swab (also known as the orange/coral vaginal swab) for vaginal specimens  
  · Urine transport tube for female urines  
  · Male specimens: Urine transport tube ONLY |
<p>| Specimen Identification:                                           | Specimens must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. DHEC requisition must be completed in full. |
| Specimen Volume (optimum):                                         | Urine should be collected up to the “fill area” lines. Swab collection kits should contain an adequate amount of transport media for testing. |
| Specimen Volume (minimum):                                         | Urine should be collected up to the “fill area” lines. Swab collection kits should contain an adequate amount of transport media for testing. |
| Collect:                                                           | Only use Gen-Probe Aptima Specimen Collection Kits. See Special Instructions for more information. |
| Form:                                                              | DHEC 1332                                                   |
| Special Instructions:                                              | Only use Gen-Probe Aptima Specimen Collection Kits. Female and male urine specimens: Patients should not have voided within one hour of collection. Collect first 20 - 30 mL of the first-catch urine stream into a collection cup. Transfer 2 mL of urine into Aptima Urine Specimen Transport tube so that the urine level falls within the two lines on the transport tube labeled: “fill area” (Yellow Label). See GC/Chlamydia Gen-Probe Collection Procedure, Section III. Male testing will ONLY be performed on urine specimens. |
| Packing and Shipping*:                                              | See Transporting and Shipping Infectious Substances in Section IV. |
| Transport Conditions:                                              | Store specimens at 2-30°C and ship specimens to be maintained at 2-30°C until received at the PHL. For longer storage, freeze transport tube within 7 days of collection at ≤ -20 °C and ship on dry ice to maintain at temperature of ≤ -20°C until received at the PHL; Swab specimens must be tested within 60 days of collection. Urine specimens should be tested within 30 days of collection (urine must be transferred to the Urine Collection Tubes within 24 hours). |
| Specimen Rejection Criteria:                                       | Specimens with no swab in transport media; white swab in transport media; two swabs in transport media; urine above or below designated black lines on transport tube labeled fill area; swab specimen more than 60 days old, or urine specimens more than 30 days old; specimens received at the improper temperature; For universal rejections, see Specimen Rejection &amp; Disclaimer Criteria in Section I. |
| Availability:                                                      | Monday - Friday                                              |
| Results and Interpretations:                                       | Positive or Negative                                         |
| Additional Information:                                            | N/A                                                          |
| Purpose of Test:                                                   | For the detection and aid in the diagnosis of trichomoniasis. |
| Method:                                                            | Nucleic acid amplification test (NAAT)                       |
| Interfering Substances:                                            | N/A                                                          |
| Comment:                                                           | N/A                                                          |</p>
<table>
<thead>
<tr>
<th>Test</th>
<th>VARICELLA VIRUS SEROLOGY (IgG)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Chickenpox, Varicella zoster virus</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>2 mL serum</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>1 mL serum</td>
</tr>
<tr>
<td>Collect:</td>
<td>Serum Separator vacuum tube (SST), centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged and serum is removed from the clot and put into a different container/tube). Please follow manufacturer’s guidelines. See Specimen Collection: Venipuncture Procedure in Section III, if needed.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1332</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See Transporting and Shipping Infectious Substances in Section IV.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store at 2 - 8°C and ship within 36 hours of collection to maintain specimen at 2 - 8°C until received by the PHL. If shipment is delayed longer than 36 hours, specimen should be stored at ≤ -20°C and shipped on dry ice to maintain the temperature of ≤ -20°C until received by the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>See Specimen Rejection &amp; Disclaimer Criteria in Section I.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As requested</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Immune status: Positive, Negative or Equivocal</td>
</tr>
<tr>
<td>Additional Information:</td>
<td></td>
</tr>
<tr>
<td><strong>Result</strong></td>
<td><strong>Interpretation</strong></td>
</tr>
<tr>
<td>Positive</td>
<td>Indicates IgG antibodies to Varicella virus were detected. A positive test result indicates a current or previous infection with Varicella virus, or prior vaccination against Varicella virus.</td>
</tr>
<tr>
<td>Equivocal</td>
<td>Re-evaluate by collecting and testing another specimen.</td>
</tr>
<tr>
<td>Negative</td>
<td>Indicates no detectable IgG antibodies to the Varicella virus. A non-reactive result indicates no current or previous infection with Varicella virus. Such patients are presumed to be non-immune and are therefore susceptible to a primary infection. A non-reactive result may be obtained early in seroconversion of infected individuals. If this is suspected, collect and test another specimen in 8 - 14 days.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect Varicella zoster virus IgG antibodies for determining immune status.</td>
</tr>
<tr>
<td>Method:</td>
<td>EIA (Enzyme Immunoassay)</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>VARIOLA</td>
</tr>
<tr>
<td>------------------------------------</td>
<td>----------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Small Pox</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>48 hours</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical specimens</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during Special Pathogens Laboratory notification.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 521; check “Other- (Variola)”</td>
</tr>
<tr>
<td></td>
<td>DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Pre-approval Needed - Hospitals must obtain approval from SC DHEC DADE (Division of Acute Disease Epidemiology) and the Special Pathogens Laboratory prior to submitting specimens. Contact information can be located on the back of the List of Reportable Conditions. Contact the Special Pathogens Laboratory (803-896-0777 / 803-767-8118) for test notification, specimen collection, storage conditions, and shipping conditions/methods.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during Special Pathogens Laboratory notification</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during Special Pathogens Laboratory notification</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Final results are verbally communicated to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail. The definitive identification of Variola virus requires additional testing to be performed by CDC.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Variola Virus is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of Variola detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To presumptively detect Variola DNA in clinical specimens</td>
</tr>
<tr>
<td>Method:</td>
<td>CDC/LRN Real Time PCR Assay</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
<tr>
<td>Test</td>
<td>VIBRIO, all types, including <em>Vibrio cholerae</em> O1 and O139</td>
</tr>
<tr>
<td>------</td>
<td>----------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>N/A</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0803</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Isolate or stool collected in stool transport medium.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container and isolates must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container or isolates should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>Pure isolate subbed to agar slant that supports growth.; Stool in transport medium, such as Cary Blair and Para Pak.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition; Mark “Organism for ID” for isolates and “Enteric Culture” for stool in transport medium.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <a href="#">Transporting and Shipping Infectious Substances in Section IV</a>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Ship isolates in approved shippers to maintain temperature of specimen within the range of 15-25°C until received at the PHL. Ship stool in transport medium on frozen cold packs in approved specialized insulated shippers to maintain temperature of specimen within the range of 2-8°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>For universal rejections, see <a href="#">Specimen Rejection &amp; Disclaimer Criteria in Section I</a>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Specimen submission to the Public Health Laboratory (PHL) is required. Ship +PCR specimens ASAP to promote recovery. Ship isolates within 1 business day.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux VITEK MS, conventional biochemicals, serotyping</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>WEST NILE VIRUS SEROLOGY- IgM</td>
</tr>
<tr>
<td>------</td>
<td>-------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Arbovirus Serology</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Virology &amp; Rabies, 803-896-0819</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>N/A</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>15 days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Cerebrospinal fluid (CSF) or Serum</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Patient’s full name and patient ID # (or other unique identifier) is required on the specimen and requisition.</td>
</tr>
</tbody>
</table>
| Specimen Volume (optimum): | CSF: 2 mL  
Serum: 2 mL |
| Specimen Volume (minimum): | CSF: 1 mL  
Serum: 1 mL |
| Collect: | Serum Separator vacuum tube (SST), centrifuged appropriately. (Red top vacuum tubes may be used if the specimen is centrifuged and serum is removed from the clot and put into a different container/tube). Please follow manufacturer’s guidelines. See Specimen Collection: Venipuncture Procedure in Section III, if needed.  
Sterile, leak-proof, screw-capped container for CSF. |
| Form: | DHEC 1332 |
| Special Instructions: | N/A |
| Packing and Shipping*: | See Transporting and Shipping Infectious Substances in Section IV. |
| Transport Conditions: | Store CSF at 2-8°C and ship within 24 hours to maintain the temperature at 2-8°C until received at the PHL. After 24 hours, ship CSF on dry ice to be received at the PHL at a temperature of ≤-20°C.  
Store Serum at 2 - 8°C and ship within 36 hours of collection to maintain specimen at 2 - 8°C until received by the PHL. If shipment is delayed longer than 36 hours, Serum specimen should be stored at ≤-20°C and shipped on dry ice to maintain the temperature of ≤-20°C until received by the PHL. |
<p>| Specimen Rejection Criteria: | See Specimen Rejection &amp; Disclaimer Criteria in Section I. |
| Availability: | N/A |
| Results and Interpretations: | | |
| | Result | Interpretation |
| | Positive | Indicates IgM antibodies to West Nile virus were detected. |
| | Negative | Indicates no detectable IgM antibodies to West Nile virus. |
| | Equivocal | Collect and submit another specimen for testing. |
| | Unable to Interpret | Non-specific interference. Unable to interpret. |
| Additional Information: | Positive specimens will be referred to CDC for additional testing. |
| Purpose of Test: | To detect IgM antibodies for the West Nile virus to determine a current infection. |
| Method: | EIA (Enzyme Immunoassay) |
| Interfering Substances: | N/A |
| Comment: | N/A |</p>
<table>
<thead>
<tr>
<th><strong>Test</strong></th>
<th><strong>YERSINIA ENTEROCOLITICA</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Synonym:</td>
<td>Y. enterocolitica</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Clinical Microbiology, 803-896-0805</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>10 Business Days</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Walnut sized portion of feces or 5 - 10mL of liquid stool in stool transport medium. Infant specimens may be collected in a disposable diaper with outside facing in. Submit referred isolate on agar slant in a screw capped tube.</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimen container or Isolate must be labelled with patient’s first and last name, and a second patient identifier such as DOB, MCI #, Specimen #. Specimen container and Isolate should have the date of isolate or collection, and initials of the person collecting the specimen. DHEC requisition must be completed in full.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>N/A</td>
</tr>
<tr>
<td>Collect:</td>
<td>N/A</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335 requisition, mark Enteric Culture or Organism for ID</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>N/A</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>See <em>Transporting and Shipping Infectious Substances in Section IV</em>.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Store specimen at 2-8°C. Ship stool preserved in Cary-Blair or Para-Pak transport medium on frozen cold packs in approved specialized insulated shipper to maintain specimen at a temperature range of 2-8°C until received at the PHL. Ship slants in approved shippers to maintain the temperature range of 15-25°C until received at the PHL.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Quantity insufficient; specimen too old; improper transport media or conditions; Specimens received at the improper temperature; For universal rejections, see <em>Specimen Rejection &amp; Disclaimer Criteria in Section I</em>.</td>
</tr>
<tr>
<td>Availability:</td>
<td>Monday - Friday</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>N/A</td>
</tr>
<tr>
<td>Additional Information:</td>
<td><em>Yersinia enterocolitica</em> testing is available for outbreaks as determined by the SC DHEC Division of Acute Disease Epidemiology (DADE).*</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>N/A</td>
</tr>
<tr>
<td>Method:</td>
<td>bioMerieux, Vitek MS</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>N/A</td>
</tr>
<tr>
<td>Test</td>
<td>YERSINIA PESTIS</td>
</tr>
<tr>
<td>---------------------------</td>
<td>-------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Synonym:</td>
<td>Bubonic Plague</td>
</tr>
<tr>
<td>Lab Section/Phone:</td>
<td>Special Pathogens / Daytime - (803) 896-0777 or Evenings - (803) 767-8118</td>
</tr>
<tr>
<td>Days Performed:</td>
<td>As needed</td>
</tr>
<tr>
<td>Turnaround Time:</td>
<td>7 to 10 days from the time of specimen receipt at the PHL</td>
</tr>
<tr>
<td>Specimen Required:</td>
<td>Clinical Specimens / Isolates</td>
</tr>
<tr>
<td>Specimen Identification:</td>
<td>Specimens should be labeled with patient’s first and last name, DOB, MCI # or other unique ID #, date and time of collection, initials of the person collecting the specimen, and the specimen source.</td>
</tr>
<tr>
<td>Specimen Volume (optimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Volume (minimum):</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Collect:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Form:</td>
<td>DHEC 1335; Test Number 520; “Suspect Agent” = Yersinia pestis</td>
</tr>
<tr>
<td></td>
<td>DHEC requisition must be completed in full and should include the date of birth and a second patient identifier (e.g., Local ID or Clinical ID), the date of isolate / collection, and initials of the person collecting the specimen.</td>
</tr>
<tr>
<td>Special Instructions:</td>
<td>Specimen must be pre-approved by Special Pathogens Supervisor prior to testing.</td>
</tr>
<tr>
<td>Packing and Shipping*:</td>
<td>Special handling criteria apply. Please contact the laboratory for special instructions at 803-896-0777 / 803-767-8118.</td>
</tr>
<tr>
<td>Transport Conditions:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Specimen Rejection Criteria:</td>
<td>Determined during pre-approval consultation.</td>
</tr>
<tr>
<td>Availability:</td>
<td>As needed</td>
</tr>
<tr>
<td>Results and Interpretations:</td>
<td>Preliminary (when applicable) and final results are verbally called to the sender to ensure correct interpretation. Final reports are provided via fax or e-mail.</td>
</tr>
<tr>
<td>Additional Information:</td>
<td>Yersinia pestis is designated as a Tier 1 Select Agent (Select Agent Regulation, 42 CFR, 73, Final Rule). In the event of Yersinia pestis detection, the Special Pathogens Laboratory will assist the sender in the timely submission of the required federal documentation.</td>
</tr>
<tr>
<td>Purpose of Test:</td>
<td>To detect Y. pestis in clinical specimens / To confirm referred isolates</td>
</tr>
<tr>
<td>Method:</td>
<td>A variety of sentinel and LRN methods are used to grow, isolate, confirm, and rule-out bacterial isolates.</td>
</tr>
<tr>
<td>Interfering Substances:</td>
<td>N/A</td>
</tr>
<tr>
<td>Comment:</td>
<td>Please call the Special Pathogens Laboratory with any questions or concerns.</td>
</tr>
</tbody>
</table>
SECTION III
ORDERING SUPPLIES
AND
SPECIMEN COLLECTION
ORDERING SUPPLIES/FORMS/SHIPPING CONTAINERS

The Public Health Laboratory (PHL) will provide request forms, kits, media, and shipping containers for the collection and shipping of laboratory specimens. These supplies are provided free of charge. Please use them judiciously and use them ONLY to send laboratory specimens to the Public Health Laboratory, SCDHEC, 8231 Parklane Road, Columbia, SC 29223. Supplies may be obtained by completing and submitting the DHEC 1323 form, “Request for Laboratory Supplies”. Email PHL-Supply@dhec.sc.gov to request the 1323 form. An electronic fillable form will be sent by email. Return the completed DHEC 1323 form by email to PHL-Supply@dhec.sc.gov. Be sure to provide the sender number, so the requested supplies are sent to the correct location. A confirmation email will be sent after receipt of the completed DHEC 1323 form. The Supply Section can be reached at (803) 896-0913, if needed.

COLLECTION KITS

These kits contain collection materials and a requisition form. Each kit is to be used for only one specimen.

- Enteric kit (for Bact. Culture) Pink Label
- Influenza kit Insulated Shipper
- Mycobacteriology (collection kit for TB) Yellow Label

TRANSPORT MEDIUM

(Order request forms and shipping container separately.)

- GC Culture medium
- Cary Blair Media
- Viral Transport Media

OTHER SUPPLIES

- Absorbent Packs
- Biohazard Bags
- Envelopes (for Newborn Screening and Hb electrophoresis blood spots)
- GC/Chlamydia/Trichomonas (for Antigen Detection) Unisex swab, MTS (Multitest) swab, also known as the orange/coral vaginal swab), or urine collection kit
- PPT Tubes for Viral Load
- QuantiFERON-TB Gold Plus (QFT Plus) Tubes

SHIPPING CONTAINERS

(use for shipping infectious specimens)

Commercial carriers must use special approved mailing containers. These are distributed for PHL use ONLY and will be returned to senders for re-use.

Shipping Containers
- Thermosafe and Uline
- Infecon 5000
- Infecon 5500
- Category A Cold Shipper
- Rabies Container
REQUEST FORMS

The request forms provided by the Public Health Laboratory are listed below. Forms marked with a + will be pre-addressed with your name, address and sender number. Since an over-supply cannot be returned to stock, please use discretion in the number you request. **DO NOT LOAN OR BORROW** pre-printed forms to another client. The pre-printed sender number determines where result reports are mailed or made available to electronically. Forms are periodically revised. Please discontinue use of old forms once a revision has been made.

**A separate DHEC 1323 form (Request for Laboratory Supplies) must be submitted for each location requesting supplies, using its unique sender number.**

<table>
<thead>
<tr>
<th>Form #</th>
<th>Test (revision date)</th>
<th>Form color</th>
</tr>
</thead>
<tbody>
<tr>
<td>1308</td>
<td>+Rabies</td>
<td>White</td>
</tr>
<tr>
<td>1323</td>
<td>Request for Lab Supplies</td>
<td>N/A (Electronic form)</td>
</tr>
<tr>
<td>1327</td>
<td>Newborn Screening</td>
<td>White</td>
</tr>
<tr>
<td></td>
<td>(Check expiration date on form)</td>
<td></td>
</tr>
<tr>
<td>1332</td>
<td>+GC/ Chlamydia Screening</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+Hematology</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+ HIV Hepatitis /Syphilis Serology</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+Immunology</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+Lead Analysis</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+Lymphocyte Subset Panel</td>
<td>White</td>
</tr>
<tr>
<td>1332</td>
<td>+Serum Chemistry</td>
<td>White</td>
</tr>
<tr>
<td>1335</td>
<td>+Bacteriology</td>
<td>White</td>
</tr>
<tr>
<td>1335</td>
<td>+Molecular</td>
<td>White</td>
</tr>
<tr>
<td>1335</td>
<td>+Mycobacteriology</td>
<td>White (Included in kit)</td>
</tr>
<tr>
<td>1335</td>
<td>+Parasitology</td>
<td>White</td>
</tr>
<tr>
<td>1335</td>
<td>+Virus Isolation/Herpes</td>
<td>White</td>
</tr>
<tr>
<td>1339</td>
<td>Hemoglobin Electrophoresis</td>
<td>Lt. Green</td>
</tr>
</tbody>
</table>

+Pre-addressed
### DHEC 1332 Submission Form

**DEPARTMENT OF HEALTH AND ENVIRONMENTAL CONTROL**  
Public Health Laboratory- 8231 Parklane Road Columbia, SC 29223  
(803) 896-0800

**CLIA #** 42D00658606

### Patient Information

<table>
<thead>
<tr>
<th>Patient’s Name (Last)</th>
<th>(First)</th>
<th>(MI)</th>
<th>Sex</th>
<th>Ethnicity</th>
<th>Race</th>
<th>Date of Birth</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Address</th>
<th>City</th>
<th>State</th>
<th>Zip Code</th>
<th>County of Residence</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Phone Number</th>
<th>Country of Birth</th>
<th>MCI Number</th>
<th>Local ID</th>
<th>Clinic ID</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Sender No.</th>
<th>Sender Name</th>
<th>Billing Number</th>
<th>Program Number</th>
<th>Clinic Type</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### Reason for Visit

- Contact
- Contact-Chlamydia
- Contact-Gonorrhea
- Contact-Hepatitis A
- Contact-Hepatitis B
- Contact-Hepatitis C
- Contact-HIV/HD/MD notified
- Contact-HIV Positive
- Contact-HIV/PT notified
- Contact-Jyphils
- Diagnosed
- Family Planning - Annual
- Family Planning - Initial
- Referral Agency
- Referred by outreach
- Referred by Drug Treatment Center
- Referred - Self
- Repeat Test/First Test
- Self Report (Date: ________)
- Special Project
- Survey
- Test of Cure
- Unknown
- Volunteer/Medical
- Workplace-Exposure
- Other
- Fast Track Ineligible
- Fast Track Services
- Pregnancy Test
- PEP Testing Services
- Routine Screening
- Serum
- Venipuncture
- Other

### Serology Test Symptoms

- Date of onset:
- Fever
- Duration:
- Rash (Type):
  - Conjunctivitis
  - Paralysis
  - Pericarditis
  - Cough
  - Pharyngitis
  - Diarrhea
  - Pneumonia
  - Headache
  - Rhinitis
  - Muscle Weakness
  - Vomiting
  - Nystagmus
  - Nuchal rigidity

### Specimen Information

<table>
<thead>
<tr>
<th>Collection Date</th>
<th>Collection time</th>
<th>Ordering Physician, Provider and/or Nurse:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AM</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PM</td>
<td></td>
</tr>
</tbody>
</table>

#### Blood

- Blood
  - Clotted
  - EDTA-Lavender/Purple
  - Finger, Heel
  - Plasma
  - Serum
  - Venipuncture*
  - Other
- Urine
- CSF

#### Swab

- Swab
  - Cervical
  - Rectal
  - Throat
  - Unknown
  - Urethral
  - Vaginal
  - Other

### Risk History (Past 12 months)

<table>
<thead>
<tr>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
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<th>11</th>
<th>12</th>
<th>13</th>
<th>14</th>
<th>15</th>
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<th>17</th>
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<th>19</th>
<th>20</th>
<th>21</th>
<th>22</th>
<th>23</th>
<th>24</th>
<th>25</th>
<th>26</th>
<th>27</th>
<th>28</th>
<th>29</th>
<th>30</th>
</tr>
</thead>
</table>

### Chlamydia Test

#### Pregnancy Status

<table>
<thead>
<tr>
<th>Yes</th>
<th>No</th>
<th>Unknown</th>
<th>Multiple partner</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

#### Symptoms

<table>
<thead>
<tr>
<th>Yes</th>
<th>No</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
</tbody>
</table>

#### Risk

<table>
<thead>
<tr>
<th>Yes</th>
<th>No</th>
<th>New partner</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### Special Instructions and/or Comments

**Virology**

<table>
<thead>
<tr>
<th>Mumps IgG</th>
<th>Mumps IgM</th>
<th>Rubella IgG</th>
<th>Rubella IgM</th>
<th>Rubella IgG</th>
<th>Rubella IgM</th>
<th>Chikungunya IgM</th>
<th>Dengue IgM</th>
<th>Vocrilla IgG</th>
<th>West Nile IgM</th>
<th>Zika IgM</th>
<th>Hg, Pb, Cd screen</th>
<th>Lead (Blood)</th>
<th>Trace Heavy Metals (includes As, Se, Cd, Ba, Ti, Pb, and U)</th>
<th>Individual metals upon request</th>
<th>Biomonitoring-No Demographics</th>
</tr>
</thead>
</table>

### Analytical Chemistry

#### GC/CT Detection

- GC and CT tRNA
- Trichomonas Detection
- Trichomonas tRNA

### Specimen type (Trichomonas):

- Urine
- Cervical
- Vaginal

### Diagnostic Serology

- Hepatitis A IgG
- Hepatitis A IgM
- HIV
- Hepatitis B Antibody
- Hepatitis B Surface Antibody
- Hepatitis B Diagnosis Profile
- Hepatitis B Core Antibody IgM
- Hepatitis B core antibody
- HIV Viral Load
- Hepatitis B surface antibody
- Hepatitis B anti-core
- Hepatitis C antibody
- Syphilis
- Hepatitis C virus load
- Syphilis IgM
- Syphilis IgG
- PreP Panel F/U (HIV, Syphilis, CT/GC)
- PreP Panel Initial (HIV, Syphilis, CT/GC, Hep C+B)
INSTRUCTIONS FOR COMPLETING REQUEST FORM
(May use printed patient lab label)

1. Enter patient name.
2. Write M = Male; F = Female or TX = Transgender M2F (Male to Female); or TY = Transgender F2M (Female to Male) in Sex box.
3. Enter ethnicity as follows: H = Hispanic/Latino, N = Non-Hispanic/Latino and U = Unknown
4. Enter race as follows: A = Asian, B = Black/African American, W = White, I = American Indian/Alaskan Native, P = Native Hawaiian/Other Pacific Islander, O = Other, U = Unknown/Unclassified
5. Enter date of birth (month, day and year. Example: Enter 03/06/1960 for the birthday March 6, 1960.)
6. Enter the patient address and five-digit zip code.
7. Enter county of residence and the 10-digit telephone number.
8. Enter Country of Birth.
9. Fill in patient MCI ID number (DHEC Clients only).
10. Enter local and clinic ID if applicable. (Private clients must provide a clinic ID)
11. Enter Sender number and Sender name.
12. Enter billing number if billing number is different from sender number
13. Enter Program number.
14. Enter Clinic Type.
15. In the Reason for Visit/Test box, check all that apply. Enter Date of Onset if applicable and check all symptoms that apply.
16. Enter the date and time of collection.
17. Enter Ordering Physician, Provider and/or Nurse if applicable. Note: Please print.
18. Check type/source of specimen.
19. Use the codes below to identify client and partner Risk Factors during the PAST 12 MONTHS. (Circle all that apply)

|-------------|---------------------|------------------|--------------------------|-----------------------------|---------------------------------------------------------------|-------------------------------------|-----------|-----------|-----------------------------------------------|----------|-----------|-------------------|---------|----------|

|-------------|---------------------|-----------|-------------|--------------------------|-----------------------------------------------|-------------------------------------|-----------|-----------|-----------------------------------------------|----------|-----------|-------------------|---------|----------|

20. Chlamydia test: Check pregnancy status, risk, and symptom.
21. Enter Special Instructions and/or Comments.
22. Check test(s) requested.
23. Send one copy of the form with the specimen(s) to the lab. Please Retain an Additional Copy For Your Records.

Request forms will be retained following DHEC records retention schedule 8581, "Requests for Laboratory Analysis". Records Group Number: 169.

DHEC 1332
**SC DEPARTMENT OF HEALTH AND ENVIRONMENTAL CONTROL**
**PUBLIC HEALTH LABORATORY**
8231 Parklane Road Columbia, SC 29223
(803) 796-0800

<table>
<thead>
<tr>
<th>Field</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patient's Name/Last</td>
<td></td>
</tr>
<tr>
<td>(SurName)</td>
<td></td>
</tr>
<tr>
<td>(First)</td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td></td>
</tr>
<tr>
<td>Date of Birth</td>
<td></td>
</tr>
<tr>
<td>Address</td>
<td></td>
</tr>
<tr>
<td>City</td>
<td></td>
</tr>
<tr>
<td>Zip Code</td>
<td></td>
</tr>
<tr>
<td>County of Residence</td>
<td></td>
</tr>
<tr>
<td>Phone Number</td>
<td></td>
</tr>
<tr>
<td>MSI Number</td>
<td></td>
</tr>
<tr>
<td>Local ID</td>
<td></td>
</tr>
<tr>
<td>Client ID</td>
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</tr>
<tr>
<td>Program Number</td>
<td></td>
</tr>
<tr>
<td>Country of Birth</td>
<td></td>
</tr>
<tr>
<td>Other Address</td>
<td></td>
</tr>
<tr>
<td>ORDERING PHYSICIAN, PROVIDER AND/OR NURSE:</td>
<td></td>
</tr>
<tr>
<td>Special Instructions/Notes</td>
<td></td>
</tr>
<tr>
<td>Date of Order</td>
<td></td>
</tr>
<tr>
<td>Agents/Organism(s) Specified</td>
<td></td>
</tr>
<tr>
<td>Specimen Information</td>
<td></td>
</tr>
<tr>
<td>Collection Date</td>
<td></td>
</tr>
<tr>
<td>Collection Time</td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td></td>
</tr>
<tr>
<td>Collection Site</td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td></td>
</tr>
<tr>
<td>Am/Pm</td>
<td></td>
</tr>
<tr>
<td>Specimen Type/Source</td>
<td></td>
</tr>
<tr>
<td>SYMPTOMS</td>
<td></td>
</tr>
<tr>
<td>Clinical Diagnosis</td>
<td></td>
</tr>
<tr>
<td>TEST REQUESTED</td>
<td></td>
</tr>
<tr>
<td>CLINICAL MICROBIOLOGY (BACTERIOLOGY)</td>
<td></td>
</tr>
<tr>
<td>Mycobacteriology</td>
<td></td>
</tr>
<tr>
<td>Special Pathogens</td>
<td></td>
</tr>
<tr>
<td>Molecular</td>
<td></td>
</tr>
<tr>
<td>PHIL USE ONLY</td>
<td></td>
</tr>
<tr>
<td>Date Received</td>
<td></td>
</tr>
<tr>
<td>PHI Specimen Number</td>
<td></td>
</tr>
</tbody>
</table>

**CLINICAL MICROBIOLOGY (BACTERIOLOGY)**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>521 Influenza RT-PCR</td>
<td></td>
</tr>
<tr>
<td>271 RSV PCR</td>
<td></td>
</tr>
<tr>
<td>277 Influenza RT-PCR</td>
<td></td>
</tr>
<tr>
<td>275 Respiratory Panel</td>
<td></td>
</tr>
</tbody>
</table>

**MYCOBACTERIOLOGY**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>601 Clinical Specimen for ID and serum</td>
<td></td>
</tr>
<tr>
<td>602 Drug Susceptibility</td>
<td></td>
</tr>
<tr>
<td>604 Quantiferon TB-Gold</td>
<td></td>
</tr>
</tbody>
</table>

**SPECIAL PATHOGENS**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>522 Serologic Testing</td>
<td></td>
</tr>
<tr>
<td>521 Molecular Testing for Viral Agents</td>
<td></td>
</tr>
<tr>
<td>512 Nucleic Acid</td>
<td></td>
</tr>
</tbody>
</table>

**MOLECULAR**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>520 Rule-out Testing</td>
<td></td>
</tr>
<tr>
<td>518 Clinical Specimen</td>
<td></td>
</tr>
</tbody>
</table>

**PHIL USE ONLY**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>410 Other</td>
<td></td>
</tr>
</tbody>
</table>
INSTRUCTIONS FOR COMPLETING REQUEST FORM
DHEC 1335
(May use printed patient lab label)

1. Enter patient name.
2. Write M = Male; F = Female or TX = Transgender M2F (Male to Female); or TY = F2M (Female to Male) in Sex box.
3. Enter ethnicity as follows: H = Hispanic/Latino and N = NonHispanic/Latino.
4. Enter race as follows: A = Asian
   B = Black/African American
   W = White
   I = American Indian/Alaskan Native
   P = Native Hawaiian/
   O = Other
   Other Pacific Islander
   U = Unknown/Unclassified
5. Enter date of birth (month, day and year.) Example: enter 03/06/1960 for the birthday March 6, 1960.
6. Enter the patient address and five-digit zip code.
7. Enter county of residence and the 10-digit telephone number.
8. Fill in patient MCI ID number (DHEC Clients only).
9. Enter local and clinic ID if applicable. (Private clients must provide a clinic ID)
10. Enter Program number.
11. Enter Country of Birth.
12. Enter billing number if billing number is different from sender number.
13. Enter the Outbreak number.
14. Enter the date and time of collection and initial.
15. Check type/source of specimen.
16. Enter Ordering Physician, Provider and/or Nurse if applicable. Note: Please print.
17. Enter in the Special Instructions and/or comments where you vacated (travel history).
18. Enter Date of Onset if applicable.
19. List agents, organisms, or virus suspected.
20. Enter clinical diagnosis.
21. Check symptoms that apply.
22. Mark test requested.
23. Answer the four questions in Mycobacteriology Section.
24. Send top two copies of the form with the specimen(s) to the lab. PLEASE RETAIN THIRD COPY FOR YOUR RECORDS.

DHEC 1335 (4/2016)
**DHEC 1335 Submission Form**

**DEPARTMENT OF HEALTH AND ENVIRONMENTAL CONTROL**

Public Health Laboratory
8231 Parklane Road Columbia, SC 29223
(803) 896-0800

<table>
<thead>
<tr>
<th>Patient's Name (Last)</th>
<th>(First)</th>
<th>(MI)</th>
<th>Sex</th>
<th>Ethnicity</th>
<th>Race</th>
<th>Date of Birth</th>
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<th>Country of Birth</th>
<th>MCI Number</th>
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<th>Clinic ID</th>
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<th>Sender No.</th>
<th>Sender Name</th>
<th>Billing Number</th>
<th>Program No.</th>
<th>Outbreak Number</th>
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**Ordering Physician, Provider and/or Nurse:**

**Clinical Diagnosis**

**Special Instructions and/or Comments:**

**Specimen Information**

<table>
<thead>
<tr>
<th>Collection Date</th>
<th>Collection Time</th>
<th>Date of Onset</th>
<th>Agents/Organisms/or Virus Suspected</th>
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</thead>
<tbody>
<tr>
<td></td>
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<td></td>
<td></td>
</tr>
</tbody>
</table>

**Specimen Type/Source**

- Blood/Serum
- Bronchial wash
- Nasopharyngeal Swab
- Stool specimens
- Throat swab
- Urine
- Wound pus drainage
- BAL
- Swab
- Genital
- Tissue/Biopsy
- Other

**Mycobacteriology Specimens**

- Induced sputum
- Spontaneous sputum
- Other

**Symptoms**

- Arthralgia/Myalgia
- Diarrhea
- Meningitis
- Rash Type:

- Asymptomatic
- Encephalitis
- Nausea/Vomiting
- Respiratory

- Conjunctivitis
- Fever
- Pleurodynia
- Other

**Test Requested**

**Clinical Microbiology (Bacteriology/Parasitology)**

- Was culture incubated before transport: Yes No 24 hours 48 hours

- Broth Specimen for Shiga toxin producing E. coli
- Culture/Isolate for Shiga toxin producing E. coli
- Legionella Urine Antigen
- Enteric Culture
- Non-Enteric Culture and ID
- Organism for ID-Aerobic
- Other

- CRE/CRPA/CRAHB
- Candida ID
- GC Culture and ID
- Other

**Mycobacteriology**

- Known TB case? Yes No
- 2/O new TB Case? Yes No
- Suspicious bx, sx? Yes No
- Current Rx? Yes No

- Clinical Specimen for ID and Smear
- Drug Susceptibility:
- Specimen for Genotyping

- Isolate for ID
- Blood Culture
- Clinical Specimen
- Referred Isolate

**Virology**

- BioFire Respiratory Panel (Outbreak Only)
- Herpes
- COVID RT-PCR
- First Test?
- Hospitalized?
- Employed in healthcare?
- Mumps RT-PCR
- Symptomatic (CDC defined)?
- Pregnant?
- Resident in a congregate care facility?
- QuantiFeron TB-Gold Plus
- In-patient
- Out-Patient
- Triplex RT-PCR

**Special Pathogens**

**Rule-out Testing**

- Bacterial Isolate
- Clinical Specimen
- Suspect Agent:

**Molecular Testing for Viral Pathogens**

- Avian Influenza
- Ebola
- MERS
- Other

**Serological Testing**

- BMAT
- Malaria
INSTRUCTIONS FOR COMPLETING REQUEST FORM
DHEC 1335
(May use printed patient lab label)

1. Enter patient name.
2. Enter M = Male; F = Female; TX = Transgender M2F (Male to Female); or TY = F2M (Female to Male) in Sex box.
3. Enter ethnicity as follows: H = Hispanic/Latino and N = NonHispanic/Latino.
4. Enter race as follows: A = Asian  B = Black/African American
   W = White  I = American Indian/Alaskan Native
   P = Native Hawaiian/  O = Other
   Other Pacific Islander  U = Unknown/Unclassified
5. Enter date of birth (month, day and year) Example: enter 03/06/1960 for the birthday March 6, 1960.
6. Enter the patient address and five-digit zip code.
7. Enter county of residence and the 10-digit telephone number.
8. Fill in patient MCI ID number (DHEC Clients only).
9. Enter local and clinic ID if applicable. (Private clients must provide a clinic ID)
10. Enter Program number.
11. Enter Country of Birth.
12. Enter billing number if billing number is different from sender number.
13. Enter the Outbreak number.
14. Enter the date and time of collection and initial.
15. Check type/source of specimen.
16. Enter Ordering Physician, Provider and/or Nurse if applicable. Note: Please print.
17. Enter in the Special Instructions and/or comments where you vacated (travel history).
18. Enter Date of Onset if applicable.
19. List agents, organisms, or virus suspected.
20. Enter clinical diagnosis.
21. Check symptoms that apply.
22. Mark test requested.
23. Answer the four questions in Mycobacteriology Section.
24. Send one copy of the form with the specimen(s) to the lab. PLEASE RETAIN AN ADDITIONAL COPY FOR YOUR RECORDS.

Request forms will be retained following DHEC records retention schedule 8581, “Requests for Laboratory Analysis”, Records Group Number: 169.
<table>
<thead>
<tr>
<th>Category</th>
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<tbody>
<tr>
<td>Animal Species</td>
<td>Cat, Dog, Bat, Fox, Raccoon, Skunk</td>
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<tr>
<td>Wild/Pet/Stray</td>
<td>Wild, Pet, Stray</td>
</tr>
<tr>
<td>Vaccinated against rabies</td>
<td>Yes, No, Unknown</td>
</tr>
<tr>
<td>Date of Death</td>
<td>MM DD YYYY</td>
</tr>
<tr>
<td>Date of Vaccination</td>
<td>MM DD YYYY</td>
</tr>
<tr>
<td>Address where found</td>
<td>Street: City:</td>
</tr>
<tr>
<td>County: Zip Code:</td>
<td></td>
</tr>
<tr>
<td>Animal shot in head?</td>
<td>Yes, No</td>
</tr>
<tr>
<td>Animal buried prior to shipment?</td>
<td>Yes, No</td>
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<tr>
<td>Date of Exposure</td>
<td>MM DD YYYY</td>
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<tr>
<td>Type of Exposure</td>
<td>Bite, Scratch, Contact Saliva, Unknown, Other</td>
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<tr>
<td>HUMAN EXPOSURE (Complete the following)</td>
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</tr>
<tr>
<td>Name of Owner</td>
<td>Street: City/Zip Code: Telephone</td>
</tr>
<tr>
<td>Name of Person(s) Exposed</td>
<td>Street: City/Zip Code: Telephone</td>
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<tr>
<td>DOMESTIC ANIMAL EXPOSURE (Complete the following)</td>
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<td>Name of Owner</td>
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<tr>
<td>Type of Animal Exposed</td>
<td>Dog, Cat, Livestock (Specify), Other (Specify)</td>
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<tr>
<td>CONDITION OF BRAIN</td>
<td>Acceptable, Unacceptable</td>
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<tr>
<td>LABORATORY RESULTS</td>
<td>Positive, Negative</td>
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<tr>
<td>Unsatisfactory for testing, specimen decomposed or deteriorated</td>
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<tr>
<td>Unsatisfactory for testing, brain stem unavailable for testing</td>
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<tr>
<td>EXCEPTION</td>
<td>Not tested, Brain deteriorated</td>
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<tr>
<td>Not tested, No brain present in skull</td>
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<td>Date Reported</td>
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Instruction for Completing Rabies Request Form

1. Check the appropriate box to identify the type of animal sent in for testing. If rodent or other is checked, specify the type of rodent (example: rat, mouse, etc) or type of other (example: opossum, horse, etc).
2. Check the appropriate box to identify the animal as wild, pet, or stray.
3. Enter the date of death.
4. Check box to indicate the animal’s vaccination status. If inoculated against rabies, enter the vaccination date.
5. Enter sender name if not pre-printed on form.
6. Enter sender address if not pre-printed on form.
7. Enter Abris number used by the sender to identify the animal being tested for rabies.
8. Enter a contact person who will be responsible for receiving results.
9. Enter an office and home or cell phone number for the contact person.
10. Enter the address where the animal was found.
11. Check box to indicate if the animal was shot in the head, buried, or frozen prior to shipment.
12. Check the reason for testing and the type of exposure. Enter the date of exposure.
13. Check if the exposure was provoked or unprovoked.
14. Enter the name and address of the owner of the animal being tested. If the animal is stray or wild, leave blank.
15. If there was human exposure, give the name of the person(s) exposed, address, and phone number.
16. If there was pet exposure, check the type of pet or domestic animal exposed. Fill in the name of the owner of the animal exposed, the street address, city, zip code, and phone number.
17. Do not write in the “For Laboratory Use Only” box.
18. Send the top two copies to the form with the animal head. Retain the third copy for your records.
## COUNTY CODES

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</table>

## SENDER NUMBERS

### Private Physician
Usually consists of the SC Medical License number preceded by the letter M.

### Group Practice
A number preceded by the letter G will be assigned to group practices at their request. Use of the group number will ensure that a single bill will be sent for tests submitted by all physicians in the practice. If you desire to be billed in this manner, please contact (803) 896-0800 for assignment of a group number. If each physician wishes to be billed separately, use the appropriate assigned sender number.

### Hospital
Consists of the hospital license number preceded by the letter H. If the test result is to be mailed directly to the patient’s physician, use the physician’s name, address and sender number in the appropriate spaces on the form and write the hospital sender number in the billing number space.

### Private Laboratory
A number assigned by the Public Health Laboratory. If not known, contact the lab at (803) 896-0800 for assignment.

### DHEC County Health Depts.
Consists of the assigned county code number preceded by a C.
BILLING NUMBERS
A billing number is only necessary if the test is to be billed to someone other than the sender.

PROGRAM NUMBERS
Used only when billing to a DHEC Program
0001 Immunization-VFC Operations
0002 Children with Special Health Care Needs (CSHCN)
0004 Family Planning
0005 Sickle Cell Program
0006 Maternal and Child Health (MCH)
0007 Cancer Control
0009 Tuberculosis Services – Outpatient
0011 Sexually Transmitted Diseases (STD)
0026 Adult Health
0027 Birth Defects (Metabolic Screening Program)
0031 Expanded & Integrated Human HIV Testing- Non-Clinical
0035 Expanded and Integrated HIV Testing for Populations-Clinical
0043 Environmental Health
0053 Newborn Metabolic Screening & Follow-Up
0055 Infant and Child Health Screening & Follow-Up
0059 WCS (Women & Children’s Services)
0063 Employee Health Services
0070 Epidemiology - Disease Control
0072 HIV-AIDS Alcohol & Drug Abuse Project
0095 WIC
0111 HIV/AIDS
0202 Immunization Program
0301 BT CDC Public Health Emergency Preparedness
SPECIMEN COLLECTION PROCEDURES

Specimen Collection: Venipuncture Using the Vacuum System

Precaution: Wear non-latex gloves and liquid impervious laboratory coat or apron while collecting and preparing blood for shipment.

Supplies: see Clinical Formulary on the intranet for approved supplies
1. Vacuum tube system
2. Vacuum needle, 1 inch or 1 ½ inch; 18, 20, 21, 22, or 23 gauge
3. Disposable vacuum needle holder
4. Disposable tourniquet
5. 70% isopropyl alcohol or benzylkonium chloride pads
6. Sterile gauze pads (NO COTTON BALLS!)
7. Band-aids (optional)
8. Sharps disposal container (with stand or wall mounted)
9. Biohazard waste container

Personal Protective Equipment (PPE) Requirements:
1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection; strongly recommended if you wear contacts)
3. Liquid resistant/impervious lab coat or apron (required during collection)
4. Additional protection as recommended by OSHA and/or Safety Data Sheet (SDS)

Specimen Collection: You MUST have completed a DHEC Workshop and be rated proficient and competent BEFORE you can collect a venipuncture without direct supervision.
1. While putting on the appropriate PPE, explain the procedure to the patient and ask the patient to identify themselves or check a picture identification card.
   a. BEFORE putting on ANY PPE, wash hands with soap and water or use a hand sanitizer with at least 70% alcohol content.
   b. Order for putting on PPE: put eye protection on first (optional), then put on liquid resistant/impervious lab coat/apron, and then put on non-latex gloves.
2. Position the patient for taking blood from the antecubital vein, or the median cubital vein, or the cephalic vein.
   a. DIS can ONLY use one of these sites to collect venipuncture.
   b. MDs, APRNs, RNs, MTs, MLTs can use other sites on the arm or hand, if trained using standard Training Checklist and passed Competency Evaluation annually.
3. Apply disposable tourniquet to the arm just above the elbow and instruct the patient to make a fist; it is NOT necessary for the patient to “pump” their fist.
   a. Always palpate the vein with the disposable tourniquet BEFORE making a decision to puncture the vein.
   b. DO NOT leave the tourniquet on for > 2 minutes during a venipuncture!
4. Select the best vein and clean the skin over the puncture site with 70% alcohol or benzylkonium chloride in
ONE DIRECTION!
a. **DO NOT** wipe back and forth with the 70% alcohol/benzylkonium chloride.
b. Allow to dry without blowing on the site or fanning the site.
c. Once the site is dry, **DO NOT** palpate the vein with gloved finger; these are not sterile gloves.

5. Use sterile vacuum needle and attach (screwed onto) to a disposable adaptor.
a. The vacuum collection tube may be inserted into the adaptor without danger of breaking the vacuum.
b. **DO NOT** pierce the vacuum on the tube with the adapter needle.

6. “Fix” the vein selected for the venipuncture.
a. Left thumb about an inch below where the needle is to enter.
b. Press down on the arm and pull the skin toward the hand.
c. The needle is to be in line with the vein.
d. The needle is to be **BEVEL SIDE UP**!
e. The needle is to be at approximately a 15-degree angle with the arm.
f. You can adjust the angle depending on the depth of the vein.

7. Puncture the skin with a clean, smooth motion. **BEVEL SIDE UP**!
a. **DO NOT** hesitate; this hurts.
b. As the needle enters the vein, a little “give” will be felt.
c. When inside the vein, grip the tube holder firmly and keep the holder steady.
d. Press the vacuum tube onto the needle portion inside the holder.

8. While the needle is inside the vein, collect the required tubes of blood.
a. Note: Collect blood in plain (red stopper) tubes before collecting blood in tubes with additives (e.g., EDTA)
b. Note: DIS are ONLY allowed to collect a single tube per venipuncture.
c. Mix tubes with additives by gently inverting 5-10 times to prevent clotting.
d. **DO NOT** shake the tube(s)!
e. Allow the red top tubes to stand in a test tube rack, upright, for at least 30 minutes to allow clotting before centrifugation.
f. Note: some patients may take longer to clot, so allow extra time if the patient is on maintenance doses of Coumadin and/or other platelet aggregate inhibitors.

9. Release tourniquet, withdraw needle from vein and apply pressure on venipuncture site with dry gauze.
a. **DO NOT** cover the injection site with an alcohol sponge while withdrawing the needle.
b. **DO NOT** apply pressure on the venipuncture site with gauze if the needle is still in the arm!

10. Have patient apply pressure on the venipuncture site for 2-3 minutes to prevent leakage of blood under the skin and formation of a hematoma. When site no longer bleeds, a bandage may be applied if desired.
a. Ask the patient to hold their arm straight up and lock their elbow.
b. If the patient cannot do this, hold the arm straight up for them.

11. Label specimen tube(s) with proper patient identification information; if not already done when getting supplies together.
a. Name of patient/client (first name and last name).
b. MCI number or other unique identification number.
c. Date collected, time collected and initials of person who collected specimen.
d. Note: if you use a computer label, just add time and initials of person collecting specimen.
12. Complete ALL information on the test request/lab requisition form(s): DHEC 1332/1335.
   a. Name of patient/client (first name and last name).
   b. MCI number or other unique identification number.
   c. Date collected, time collected and initials of person who collected specimen.
   d. Test required
   e. Type of specimen
   f. Ordering physician, APRN, RN, DIS
   g. Test(s) requested
   h. Sender Address or Sender code number
   i. Any specimen instructions or other important information
   j. Note: if you use a computer label, just add time and initials of person collecting specimen.
   k. Note: if you use a computer label, be sure to place the computer label with all demographic information on copy 1, copy 2 and copy 3.
   l. You will send the original top copy of the DHEC 1332/1335 with the specimen(s)
   m. Retain the third copy for your files in the County Health laboratory.

13. Properly dispose of needles (in biohazard puncture proof sharps container) and other contaminated materials used during venipuncture.
   a. Place all blood-soaked material in the contaminated waste bag (Biohazard).
   b. Place all sharps in the sharps container and DO NOT fill above 2/3!
   c. DO NOT place any non-contaminated waste in the sharps container or Biohazard waste!

14. Remove PPEs in this order:
   a. Remove contaminated gloves.
   b. Wash hands with soap and water or with hand sanitizer with >60% alcohol.
   c. Remove any eye protection.
   d. Remove liquid impervious/resistant lab coat or apron.
   e. Wash hands with soap and water or with hand sanitizer with >60% alcohol.

15. BEFORE allowing the patient/client to leave, take the gauze off of the venipuncture site to ensure it has stopped bleeding.
   a. DO NOT wipe the area with gauze since this will initiate bleeding again (subcutaneous) and may cause a hematoma (bruise).
   b. DO NOT allow patient to get up from the chair, table, etc. without being physically at the side or in front of the patient/client: THIS IS A FALL PREVENTION MEASURE!

16. NOTE: DIS staff can ONLY draw one tube; no multi-draws or multiple tubes collected from the same venipuncture collection site.

17. No DIS staff can be trained and/or use a butterfly to collect a venipuncture.

Specimen Preparation:
1. Blood collected in a plain red stopper tube or in a serum separator tube (SST): allow the tube of blood to remain undisturbed in an upright position at room temperature for 20-30 minutes.
   a. When the specimen has clotted, DO NOT allow the serum to sit on the clot, whether collected in a red tube or SST tube, without separating through centrifugation; then store according to instructions in the Reference Laboratory manual for specimen collection and transport.
   b. Note: check manufacturer’s package insert for maximum time blood can sit on clot BEFORE centrifugation, if using an SST (serum separator tube).
2. After a clot has formed, gently loosen the clot at the top; “rim” with a sterile applicator stick, if necessary.

3. Centrifuge tubes for 10-15 minutes.
   a. Since all centrifuges are calibrated by Instrumentation Department (Facilities Management Division, Public Health Laboratory), the time for most centrifugation needs will be on the instrument.
   b. CHECK OUT THE CALIBRATED TIME/SPEED ON YOUR CENTRIFUGE!

4. Remove the serum carefully with a sterile transfer pipette, and transfer to a clean sterile rubber-stoppered tube or to a screw-top, plastic vial/tube. Avoid transferring any red cells.

5. Label tube or plastic vial running up the length of the tube.
   a. **Do NOT** wrap the label around or “flag” the label by pressing ends together and extending from the tube.
   b. This includes ALL vacuum tubes for chemistry, hematology and serology; red top, SST, or purple (EDTA) top tubes would be the common ones.

6. Store tubes of labeled serum in a refrigerator until the specimens are ready to ship to the clinic laboratory or the Public Health Laboratory.

Special Procedural Notes:
1. If sending whole blood in a vacuum tube, omit steps 2 and 4 (see page III-6).

2. If using serum separator tubes (gel in the bottom of the tube, SST) omit steps 2 and 4 above. Be sure the gel forms a distinct barrier between serum and clot.

3. During the summer months, pack all SST specimens with a cold pack since the gel can possibly breakdown at temperatures experienced during the summer months. The breakdown of the gel allows the red blood cells to “leak” into the serum contaminating the specimen and possibly rendering the specimen unacceptable for testing.

4. Never use a gauge needle size smaller than a 23: this can cause hemolysis!

6. Always allow the blood to flow into a vacuum tube without adding additional pressure.

7. **DO NOT** take disposable tourniquet off until you have collected ALL of the tubes and the tubes are filled: when you take the tourniquet off once you are inside you run the risk of slow blood flow and/or short draws and/or insufficient blood volume.

8. For special considerations using a butterfly for a venipuncture, see the next procedure.
Specimen Collection: Venipuncture Using a Butterfly System

Precaution: Wear non-latex gloves and liquid impervious laboratory coat or apron while collecting and preparing blood for shipment.

Supplies: see Clinical Formulary on the intranet for approved supplies
1. Vacuum tube system
2. Butterfly needle: 21g, 22g, or 23g (NO SMALLER THAN 23G!)
3. Disposable vacuum needle holder
4. Disposable tourniquet
5. 70% isopropyl alcohol or benzylkonium chloride pads
6. Sterile gauze pads (NO COTTON BALLS!)
7. Band-aids (optional)
8. Sharps disposal container (with stand or wall mounted)
9. Biohazard waste container

Personal Protective Equipment (PPE) Requirements:
1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection; strongly recommended if you wear contacts)
3. Liquid resistant/impervious lab coat or apron (required during collection)
4. Additional protection as recommended by OSHA and/or MSDS

Specimen Collection: You MUST have completed a DHEC Workshop and be rated proficient and competent BEFORE you can collect a venipuncture without direct supervision. No DIS staff can be trained using this method.

Note: the use of a butterfly is to be used ONLY in special circumstances: elderly patients with non-patent veins; young children (less than 4 years old) or babies; patients with non-patent veins and the hand is the site of choice.

1. While putting on the appropriate PPE, explain the procedure to the patient and ask the patient to identify themselves or check a picture identification card.
   a. BEFORE putting on ANY PPE, wash hands with soap and water or use a hand sanitizer with at least 60% alcohol content.
   b. Order for putting on PPE: put eye protection on first (optional), then put on liquid resistant/impervious lab coat/apron, and then put on non-latex gloves.

2. Position the patient for taking blood from the antecubital vein, or the median cubital vein, or the cephalic vein: these are all on the arm.
   a. DIS can ONLY use one of these sites to collect venipuncture.
   b. MDs, APRNs, RNs, MTs, MLTs can use other sites on the arm or hand, if trained using standard Training Checklist and passed Competency Evaluation annually.
      1) Veins from the hand that can be used are: basilic veins (runs along the 5th digit, little finger).
      2) Veins from the hand that can be used are: metacarpal veins (runs along the 2nd or 4th digit, index/pointer finger and ring finger).
      3) Veins from the hand that can be used are: cephalic vein (runs along the side of wrist area or just above the thumb).
4) NO OTHER sites are to be used with the butterfly other than those listed in the venipuncture using the vacuum and the butterfly; no femoral, no temporal, no jugular, etc.

3. Apply disposable tourniquet to the arm just above the elbow, or on the forearm if using the hand, and instruct the patient to make a fist; it is NOT necessary for the patient to “pump” their fist.
   a. Always palpate the vein with the disposable tourniquet BEFORE making a decision to puncture the vein.
   b. **DO NOT leave the tourniquet on for >2 minutes during a venipuncture!**

4. Select the best vein and cleanse the skin over the puncture site with 70% alcohol or benzylkonium chloride in **ONE DIRECTION!**
   a. **DO NOT** wipe back and forth with the 70% alcohol/benzylkonium chloride.
   b. Allow to dry without blowing on the site or fanning the site.
   c. Once the site is dry, **DO NOT** palpate the vein with gloved finger; these are not sterile gloves.

5. Use sterile butterfly needle and attach (screwed onto) to a disposable adaptor.
   a. If a butterfly is used with a syringe (5cc, 7cc or 10cc), collect the specimen following the same steps, except you will fill the vacuum tubes with the blood from the syringe.
   b. **DO NOT** put blood into the vacuum tubes by pressing the needle through the rubber septum; take the rubber septum off and gently add blood to the tube.
   c. The vacuum collection tube may be inserted into the adaptor without danger of breaking the vacuum.
   d. **DO NOT** pierce the vacuum on the tube with the adapter needle.

6. “Fix” the vein selected for the venipuncture.
   a. Left thumb about an inch below where the needle is to enter.
   b. Press down on the arm and pull the skin toward the hand.
   c. The needle is to be in line with the vein.
   d. The needle is to be BEVEL SIDE UP!
   e. The needle is to be at approximately a 15 degree angle with the arm.
   f. You can adjust the angle depending on the depth of the vein.

7. Puncture the skin with a clean, smooth motion. BEVEL SIDE UP!
   a. **DO NOT** hesitate; this hurts.
   b. As the needle enters the vein, a little “give” will be felt.
   c. When inside the vein, grip the tube holder firmly and keep the holder steady.
   d. Press the vacuum tube onto the needle portion inside the holder.

8. While the needle is inside the vein, collect the required tubes of blood.
   a. **Note: Collect blood in plain (red stopper) tubes before collecting blood in tubes with additives (e.g. EDTA).**
   b. **Note:** DIS is ONLY allowed to collect a single tube per venipuncture.
   c. Mix tubes with additives by gently inverting 5-10 times to prevent clotting.
   d. **DO NOT** shake the tube(s)!
   e. Allow the red top tubes to stand in a test tube rack, upright, for at least 30 minutes to allow clotting before centrifugation.
   f. **Note:** some patients may take longer to clot, so allow extra time if patient is on maintenance doses of Coumadin and/or other platelet aggregate inhibitors.
9. Release tourniquet, withdraw needle from vein and apply pressure on venipuncture site with dry gauze.
   a. **DO NOT** cover the injection site with an alcohol sponge while withdrawing needle.
   b. **DO NOT** apply pressure on the venipuncture site with gauze if the needle is still in the arm!

10. Have patient apply pressure on the venipuncture site for 2-3 minutes to prevent leakage of blood under the skin and formation of a hematoma. When site no longer bleeds, a bandage may be applied if desired.
   a. Ask the patient to hold their arm straight up and lock their elbow.
   b. If the patient cannot do this, hold the arm straight up for them.

11. Label specimen tube (s) with proper patient identification information; if not already done when getting supplies together.
    a. Name of patient/client (first name and last name).
    b. MCI number or other unique identification number.
    c. Date collected, time collected and initials of person who collected specimen.
    d. Note: if you use a computer label, just add time and initials of person collecting specimen.

12. Complete ALL information on the test request/lab requisition form(s): DHEC 1332/1335.
    a. Name of patient/client (first name and last name).
    b. MCI number or other unique identification number.
    c. Date collected, time collected and initials of person who collected specimen.
    d. Test required
    e. Type of specimen
    f. Ordering physician, APRN, RN, DIS
    g. Test(s) requested
    h. Sender Address or Sender code number
    i. Any specimen instructions or other important information
    j. Note: if you use a computer label, just add time and initials of person collecting specimen.
    k. Note: if you use a computer label, be sure to place the computer label with all demographic information on copy 1, copy 2 and copy 3.
    l. You will send the original top copy of the DHEC 1332/1335 with the specimen(s)
    m. Retain the third copy for your files in the County Health laboratory.

13. Properly dispose of needles (in biohazard puncture proof sharps container) and other contaminated materials used during venipuncture.
    a. Place all blood-soaked material in the contaminated waste bag (Biohazard).
    b. Place all sharps in the sharps container and **DO NOT** fill above 2/3!
    c. **DO NOT** place any non-contaminated waste in the sharps container or Biohazard waste!

14. Remove PPEs in this order:
    a. Remove contaminated gloves.
    b. Wash hands with soap and water or with hand sanitizer with >60% alcohol.
    c. Remove any eye protection.
    d. Remove liquid impervious/resistant lab coat or apron.
    e. Wash hands with soap and water or with hand sanitizer with >60% alcohol.

15. BEFORE allowing the patient/client to leave, take the gauze off the venipuncture site to ensure it has stopped bleeding.
    a. **DO NOT** wipe the area with gauze since this will initiate bleeding again (subcutaneous) and may cause a hematoma (bruise).
    b. **DO NOT** allow patient to get up from the chair, table, etc. without being physically at the side or in
Specimen Preparation:

1. Blood collected in a plain red stopper tube or in a serum separator tube (SST): allow the tube of blood to remain undisturbed in an upright position at room temperature for 20 - 30 minutes.
   a. When the specimen has clotted, DO NOT allow the serum to sit on the clot, whether collected in a red tube or SST tube, without separating through centrifugation; then store according to instructions in the Reference Laboratory manual for specimen collection and transport.
   b. Note: check manufacturer’s package insert for maximum time blood can sit on clot BEFORE centrifugation, if using an SST (serum separator tube).

2. After a clot has formed, gently loosen the clot at the top; “rim” with a sterile applicator stick, if necessary.

3. Centrifuge tubes for 10 - 15 minutes.
   a. Since all centrifuges are calibrated by the Instrumentation Department (Facilities Management Division, Public Health Laboratory), the time for most centrifugation needs will be on the instrument.
   b. CHECK OUT THE CALIBRATED TIME/SPEED ON YOUR CENTRIFUGE!

4. Remove the serum carefully with a sterile transfer pipette, and transfer to a clean sterile rubber-stoppered tube or to a screw-top, plastic vial/tube. Avoid transferring any red cells.

6. Label tube or plastic vial running up the length of the tube.
   a. **Do NOT** wrap the label around or “flag” the label by pressing ends together and extending from the tube.
   b. This includes ALL vacuum tubes for chemistry, hematology and serology; red top, SST, or purple (EDTA) top tubes would be the common ones.

7. Store tubes of labeled serum in a refrigerator until the specimens are ready to ship to the clinic laboratory or the Public Health Laboratory.

Special Procedural Notes:

1. If sending whole blood in a vacuum tube, omit steps 2 and 4 (see page III-6).

2. If using serum separator tubes (gel in the bottom of the tube, SST) omit steps 2 and 4 above. Be sure the gel forms a distinct barrier between serum and clot.

3. During the summer months, pack all SST specimens with a cold pack since the gel can possibly breakdown at temperatures experienced during the summer months. The breakdown of the gel allows the red blood cells to “leak” into the serum contaminating the specimen and possibly rendering the specimen unacceptable for testing.

4. Always refer to the Public Health Laboratory Services Guide for complete instructions for specimen collection, specimen preparation, specimen storage and specimen transport for the specific laboratory test(s). Note: use current edition only.
5. Never use a gauge needle size smaller than a 23: this can cause hemolysis!

6. Always allow the blood to flow into a vacuum tube without adding additional pressure.

7. DO NOT take disposable tourniquet off until you have collected ALL of the tubes and the tubes are filled: when you take the tourniquet off once you are inside you run the risk of slow blood flow and/or short draws and/or insufficient blood volume.
Specimen Collection: Fingerstick Procedure for Patients Greater Than 1 Year Old

Hemoglobin or General Laboratory Procedures

Precaution: Wear gloves and liquid resistant lab coat or apron while collecting and preparing blood for shipment.

Supplies: see Clinical Formulary on the intranet for approved supplies

1. Retractable safety lancets: see Clinical Formulary on the intranet for approved lancets for adults and pediatrics
2. 70% isopropyl alcohol pads or benzylkonium chloride pads
3. Sterile gauze pads (NO COTTON BALLS!)
4. Band-aids (optional)
5. Sharps disposal container (with stand or wall mounted)
6. Biohazard waste container

Personal Protective Equipment (PPE) Requirements:

1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection; strongly recommended if wear contact lens)
3. Liquid resistant lab coat or apron (required during collection)
4. Additional protection as recommended by OSHA and/or MSDS

Specimen Collection: You MUST have completed a DHEC Workshop and be rated proficient and competent BEFORE you can collect a fingerstick without direct supervision.

1. While putting on the appropriate PPE, explain the procedure to the patient and ask the patient to identify themselves or check a picture identification card; if a child, ask parent and/or guardian to state child’s name.
   a. BEFORE putting on ANY PPE, wash hands with soap and water or use a hand sanitizer with at least 60% alcohol content.
   b. Order for putting on PPE: put eye protection on first (optional), then put on liquid resistant/impervious lab coat/apron, and then put on non-latex gloves.

2. Have all supplies within easy reach and all materials ready to use before performing the fingerstick procedure.

3. Place the sharps disposal container and waste container so you DO NOT have to cross over the patient or yourself when discarding contaminated items.

4. Wash hands with soap and water or use a hand sanitizer with at least 60% alcohol; put on disposable gloves.

5. Instruct the patient to rest his/her arm downward position for about 30 seconds to allow blood flow to the fingertips. If the patient’s hand is cold, warm the hand:
   a. Gently massage the finger a few times from the base to the tip of the finger.
   b. Stroke the arm with gentle downward motion from the forearm to the hand.
   c. Ask the patient to briskly rub both hands together.
   d. Use a warm (not more than 105 degrees F.), moist towel on the hand for a couple of minutes.
   e. Ask the patient to wash his/her hands with warm water.
6. Select the middle or ring finger for puncture on the hand used least often.

7. Do NOT choose a puncture site on a fingertip that is callused, purple, scarred, swollen, or injured.

8. Use the less painful, fleshy area of the fingertip, just off center to the finger pad, slightly to the side.

9. Clean the puncture site with an alcohol pad or benzylkonium chloride pad.
   a. Wipe in one direction ONLY!
   b. Allow the alcohol or benzylkonium chloride to evaporate.
   c. Do NOT blow on the finger or fan the area.

10. Do NOT saturate the site with alcohol.
    Note: Discard the used alcohol pad and wrapper in the regular trash can.

11. Allow the site to air dry completely.

12. Firmly hold the patient’s finger, palm side up, between your thumb and index finger.

13. Puncture the site and dispose of the used lancet in the sharps container.
    a. Note: Puncture the finger with the lancet PERPENDICULAR to the ridge swirls on the finger.
    b. Place the lancet FIRMLY on the finger pad site BEFORE triggering the lancet.

14. Wipe away the first 2-3 drops of blood with the sterile gauze.

15. Apply gentle pressure every few seconds, about ½ inches from the puncture site.

16. Collect specimen in a microtube (bullet), or for dried blood spots, or a hemoglobin cuvette.

17. Have patient apply pressure on the site for 2-3 minutes to prevent leakage of blood under the skin and formation of a hematoma. When site is no longer bleeding, a bandage may be applied if desired.
    a. Ask the patient/or parent to hold the gauze on the finger.
    b. If the patient cannot do this, hold the finger for them.

18. Label specimen tube (s) with proper patient identification information; if not already done when getting supplies together.
    a. Name of patient/client (first name and last name).
    b. MCI number or other unique identification number.
    c. Date collected, time collected and initials of person who collected specimen goes on the requisition/lab form.
    d. Note: if you use a HemoCue microcuvette, the frosted edge needs to have patient’s last name at least and the date.

19. Complete ALL information on the test request/lab requisition form(s): DHEC 1332/1335.
    a. Name of patient/client (first name and last name).
    b. MCI number or other unique identification number.
    c. Date collected, time collected and initials of person who collected specimen.
    d. Test required
    e. Type of specimen
    f. Ordering physician, APRN, RN, DIS
    g. Test(s) requested
h. Sender Address or Sender code number
i. Any specimen instructions or other important information
j. Note: if you use a computer label, just add time and initials of person collecting specimen.
k. Note: if you use a computer label, be sure to place the computer label with all demographic information on copy 1, copy 2 and copy 3.
l. You will send the original top copy of the DHEC 1332/1335 with the specimen(s)
m. Retain the third copy for your files in the County Health laboratory.

20. Properly dispose of lancets (in biohazard puncture proof sharps container) and other contaminated materials used during fingerstick.
   a. Place all blood soaked material in the contaminated waste bag (Biohazard).
   b. Place all sharps in the sharps container and DO NOT fill above 2/3!
   c. DO NOT place any non-contaminated waste in the sharps container or Biohazard waste!

21. Remove PPEs in this order:
   a. Remove contaminated gloves.
   b. Wash hands with soap and water or with hand sanitizer with >60% alcohol.
   c. Remove any eye protection.
   d. Remove liquid impervious/resistant lab coat or apron.
   e. Wash hands with soap and water or with hand sanitizer with >60% alcohol.

22. BEFORE allowing the patient/client to leave, take the gauze off of the fingerstick site to ensure it has stopped bleeding.
   a. DO NOT wipe the area with gauze since this will initiate bleeding again (subcutaneous) and may cause a hematoma (bruise).
   b. DO NOT allow patient to get up from the chair, table, etc. without being physically at the side or in front of the patient/client: THIS IS A FALL PREVENTION MEASURE!
Specimen Collection: Fingerstick for Patients Greater Than 1 Year Old

Dried Blood Spots Collection

Precaution: Wear gloves and liquid resistant lab coat or apron while collecting and preparing blood for shipment.

The filter paper to be used in the collection of dried blood spots is attached to the DHEC form 1339 for HEMOGLOBIN ELECTROPHORESIS or the DHEC form 1327 for PKU MONITORING. Envelopes for mailing specimen are also available.

Sufficient blood MUST be obtained from the fingerstick puncture to fill each preprinted, dashed circle by making a single application of blood to the filter paper. The filter paper should touch only the drop of blood and should not be pressed against the skin around the puncture. Be sure that the filter paper is saturated through with blood. DO NOT layer blood drops! This leads to inaccurate results. It is very important to fill each circle with ONE LARGE blood drop.

Supplies:
1. Retractable safety lancets for infant or pediatric: see Clinical Formulary listings on the intranet for approved lancets
2. 70% isopropyl alcohol or benzylkonium chloride pads
3. Sterile gauze pads (NO COTTON BALLS!)
4. Band-aids (optional)
5. Sharps disposal container (with stand or wall mounted)
6. Biohazard waste container

Personal Protective Equipment (PPE) Requirements:
1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection; strongly recommend if wear contact lens)
3. Liquid resistant lab coat or apron (required during collection)
4. Additional protection as recommended by OSHA and/or MSDS

Specimen Collection:
1. While putting on the appropriate PPE, explain the procedure to the patient and ask the patient to identify themselves or check a picture identification card; if a child, ask parent and/or guardian to state child’s name.
   a. BEFORE putting on ANY PPE, wash hands with soap and water or use hand sanitizer with at least 60% alcohol content.
   b. Order for putting on PPE: put eye protection on first (optional), then put on liquid resistant/impervious lab coat/apron, and then put on non-latex gloves.
2. Have all supplies within easy reach and all materials ready to use before performing the fingerstick procedure.
3. Place the sharps disposal container and waste container so you DO NOT have to cross over the patient or yourself when discarding contaminated items.
4. Wash hands with soap and water or use a hand sanitizer with at least 60% alcohol; put on disposable gloves.
5. Instruct the patient to rest his/her arm downward position for about 30 seconds to allow blood flow to the fingertips. If the patient’s hand is cold, warm the hand:
   a. Gently massage the finger a few times from the base to the tip of the finger.
   b. Stroke the arm with gentle downward motion from the forearm to the hand.
   c. Ask the patient to briskly rub both hands together.
   d. Use a warm (not more than 105 °F.), moist towel on the hand for a couple of minutes.
   e. Ask the patient to wash their hands with warm water.

6. Select the middle or ring finger for puncture on the hand used least often.

7. **Do NOT** choose a puncture site on a fingertip that is callused, purple, scarred, swollen, or injured.

8. Use the less painful, fleshy area of the fingertip, just off center to the finger pad, slightly to the side.

9. Clean the puncture site with an alcohol pad or benzylkonium chloride pad.
   a. Wipe in one direction **ONLY**!
   b. Allow the alcohol or benzylkonium chloride to evaporate.
   c. **Do NOT** blow on the finger or fan the area.

10. **Do NOT** saturate the site with alcohol.
    Note: Discard the used alcohol pad and wrapper in the regular trash can.

11. Allow the site to air dry completely.

12. Firmly hold the patient’s finger, palm side up, between your thumb and index finger.

13. Puncture the site and dispose of the used lancet in the sharps container.
    a. Note: Puncture the finger/heel with the lancet PERPENDICULAR to the ridge swirls on the finger.
    b. Place the lancet FIRMLY on the finger pad/heel site BEFORE triggering the lancet.

14. Wipe away the first drop of blood with the sterile gauze.

15. Apply gentle pressure every few seconds, about ½ inches from the puncture site.


17. Have patient apply pressure on the site for 2-3 minutes to prevent leakage of blood under the skin and formation of a hematoma. When site is no longer bleeding, a bandage may be applied if desired.
    a. Ask the patient/or parent to hold the gauze on the finger.
    b. If the patient cannot do this, hold the finger for them.

18. Complete ALL information on the 1327 or 1339 form:
    a. Name of patient/client (first name and last name).
    b. MCI number or other unique identification number.
    c. Date collected, time collected and initials of person who collected specimen.
    d. Ordering physician, APRN, RN, DIS
    e. Complete submitter and/or physician information
    f. You will send the original top copy of the DHEC 1327/1339 with the specimen(s).
    g. Retain the middle copy for your files.
19. Properly dispose of lancets (in biohazard puncture proof sharps container) and other contaminated materials used during fingerstick.
   a. Place all blood-soaked material in the contaminated waste bag (Biohazard).
   b. Place all sharps in the sharps container and DO NOT fill above 2/3!
   c. DO NOT place any non-contaminated waste in the sharps container or Biohazard waste!

20. Remove PPEs in this order:
   a. Remove contaminated gloves.
   b. Wash hands with soap and water or with hand sanitizer with >60% alcohol.
   c. Remove any eye protection.
   d. Remove liquid impervious/resistant lab coat or apron
   e. Wash hands with soap and water or with hand sanitizer with > 60% alcohol.

21. BEFORE allowing the patient/client to leave, take the gauze off of the fingerstick site to ensure it has stopped bleeding.
   a. DO NOT wipe the area with gauze since this will initiate bleeding again (subcutaneous) and may cause a hematoma (bruise).
   b. DO NOT allow patient to get up from the chair, table, etc. without being physically at the side or in front of the patient/client: THIS IS A FALL PREVENTION MEASURE!

22. Special Procedural Notes for Dried Blood Spots:
   a. When properly filled, the blood spot will be the same size on both sides of the filter paper.
   b. DO NOT send the specimen if the circles are not completely filled - collect a second specimen.
   c. All the circles are needed. If tests have to be repeated or additional tests need to be run, all 5 circles are required.

Troubleshooting:

1. Failure to wipe off alcohol residue may dilute the specimen and adversely affect test results.

2. Puncturing the heel on posterior curvature will permit blood to flow away from puncture, making proper spotting difficult.

3. DO NOT lance on previous puncture site.

4. Use of a capillary tube is not recommended since application of blood with a capillary tube results in scratching and/or abrading the surface of the filter paper which adversely affects test results.

5. Avoid touching area within filter paper circles before blood is applied.

6. DO NOT place filter paper in the envelope until thoroughly dry.

7. INSUFFICIENT DRYING ADVERSELY AFFECTS TEST RESULTS!
Specimen Collection: Heel-stick Procedure for Patients Less Than 1 Year Old

Hemoglobin or General Laboratory Testing or Newborn Screening

Precaution: Wear gloves and liquid resistant lab coat or apron while collecting and preparing blood for shipment.

Supplies: see Clinical Formulary on the intranet for approved supplies

1. Retractable safety lancets: Tenderfoot™ or lancet giving 1.0 mm – 2.0 mm depth
2. 70% isopropyl alcohol or benzylkonium chloride pads
3. Sterile gauze pads (NO COTTON BALLS!)
4. Band-aids (optional)
5. Sharps disposal container (with stand or wall mounted)
6. Biohazard waste container

Personal Protective Equipment (PPE) Requirements:

1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection; strongly recommend if wear contact lens)
3. Liquid resistant lab coat or apron (required during collection)
4. Additional protection as recommended by OSHA and/or MSDS

Protective Equipment (PPE) Requirements:

1. Disposable gloves (required during collection)
2. Safety glasses (required if there is any chance of eye/mouth contamination during collection)
3. Liquid resistant lab coat or apron (required during collection)
4. Closed-toe shoes MUST be worn when collecting ANY blood specimens
5. Additional protection as recommended by OSHA and/or safety data sheet (SDS)

Specimen Collection: You MUST have completed a DHEC Workshop and be rated proficient and competent BEFORE you can collect a fingerstick without direct supervision.

1. While putting on the appropriate PPE, explain the procedure to the patient and ask the patient to identify themselves or check a picture identification card; if a child, ask parent and/or guardian to state child’s name.
   a. BEFORE putting on ANY PPE, wash hands with soap and water or use a hand sanitizer with at least 60% alcohol content.
   b. Order for putting on PPE: put eye protection on first (optional), then put on liquid resistant/impervious lab coat/apron, and then put on non-latex gloves.

2. Have all supplies within easy reach and all materials ready to use before performing the heel stick procedure.

3. Place the sharps disposal container and waste container so you DO NOT have to cross over the patient or yourself when discarding contaminated items.

4. Wash hands with soap and water or use a hand sanitizer with at least 60% alcohol; put on disposable gloves.
5. Instruct the parent/guardian to rest the leg of the infant in a downward position for about 30 seconds to allow blood flow to the foot. If the patient’s foot is cold, warm the foot:
   1. Gently massage the foot/heel a few times from the base to the tip of the heel.
   2. Stroke the heel with gentle downward motion from the ankle to the toes.
   3. Ask the patient to briskly rub both hands together.
   4. Use a warm (not more than 105 °F.), moist towel on the heel for a couple of minutes.
   5. Ask the parent/guardian to wash child’s foot/heel with warm water.

6. Select the heel for puncture.
   Note: Use **ONLY** the lateral or medial sides of the heel.
   Note: **DO NOT** use the plantar region of the foot or great toe.

7. **Do NOT** choose a puncture site on a heel that is callused, purple, scarred, swollen, or injured.

8. Get all microcuvettes ready and LABEL NOW!!! Use a #2 pencil or black Sharpie.

9. Clean the puncture site with an alcohol pad or benzylkonium chloride pad.
   a. Wipe in one direction **ONLY**!
   b. Allow the alcohol or benzylkonium chloride to evaporate.
   c. Do **NOT** blow on the finger or fan the area.

10. Do **NOT** saturate the site with alcohol.
    a. Remove excess alcohol with a clean gauze pad.
    b. Discard the used alcohol pad and wrapper in the regular trash can.

11. Allow the site to air dry completely.

12. Firmly hold the patient’s heel between your thumb and index finger.

13. Puncture the site and dispose of the used lancet in the sharps container.
    a. Note: Puncture the heel with the lancet **PERPENDICULAR** to the ridge swirls on the heel.
    b. Place the lancet **FIRMLY** on the heel site BEFORE triggering the lancet.

14. Wipe away the first 2 - 3 drops of blood with the sterile gauze.

15. Apply gentle pressure every few seconds, about ½ inches from the puncture site.

16. Collect specimen in a microtube (bullet), or for dried blood spots, or a hemoglobin cuvette. LABEL NOW!!

17. Have patient apply pressure on the site for 2 - 3 minutes to prevent leakage of blood under the skin and formation of a hematoma. When site is no longer bleeding, a bandage may be applied if desired; elevate the leg higher than the heart.
    a. Ask the parent to hold the gauze on the puncture site.
    b. If the parent cannot do this, hold the heel elevated above the heart.

18. Label specimen tube (s) with proper patient identification information; if not already done when getting supplies together.
    a. Name of patient/client (first name and last name).
b. MCI number or other unique identification number.
c. Date collected, time collected and initials of person who collected specimen goes on the requisition/lab form.
d. Note: if you use a HemoCue microcuvette, the frosted edge needs to have patient’s last name at least and the date.

19. Complete ALL information on the test request/lab requisition form(s): DHEC 1332/1335.
   a. Name of patient/client (first name and last name).
   b. MCI number or other unique identification number.
   c. Date collected, time collected and initials of person who collected specimen.
   d. Test required
   e. Type of specimen
   f. Ordering physician, APRN, RN, DIS
   g. Test(s) requested
   h. Sender Address or Sender code number
   i. Any specimen instructions or other important information
   j. Note: if you use a computer label, just add time and initials of person collecting specimen.
   k. Note: if you use a computer label, be sure to place the computer label with all demographic information on copy 1, copy 2 and copy 3.
   l. You will send the original top copy of the DHEC 1332/1335 with the specimen(s).
   m. Retain the third copy for your files in the County Health laboratory.

20. Properly dispose of lancets (in biohazard puncture proof sharps container) and other contaminated materials used during heelstick.
   a. Place all blood-soaked material in the contaminated waste bag (Biohazard).
   b. Place all sharps in the sharps container and **DO NOT** fill above 2/3!
   c. **DO NOT** place any non-contaminated waste in the sharps container or Biohazard waste!

21. Remove PPEs in this order:
   a. Remove contaminated gloves.
   b. Wash hands with soap and water or with hand sanitizer with >60% alcohol.
   c. Remove any eye protection.
   d. Remove liquid impervious/resistant lab coat or apron.
   e. Wash hands with soap and water or with hand sanitizer with >60% alcohol.

22. **BEFORE** allowing the patient/client to leave, take the gauze off of the puncture site to ensure it has stopped bleeding.
   a. **DO NOT** wipe the area with gauze since this will initiate bleeding again (subcutaneous) and may cause a hematoma (bruise).
   b. **DO NOT** allow parent/patient to get up from the chair, table, etc. without being physically at the side or in front of the parent/patient: **THIS IS A FALL PREVENTION MEASURE!**

Special Procedural Notes for Dried Blood Spots:
1. When properly filled, the blood spot will be the same size on both sides of the filter paper.
2. **DO NOT** send the specimen if the circles are **NOT** completely filled—collect a second specimen.
3. All the circles are needed. If tests have to be repeated or additional tests need to be run, all 5 circles are required.

Troubleshooting:

1. Failure to wipe off alcohol residue may dilute the specimen and adversely affect test results.

2. Puncturing the heel on posterior curvature will permit blood to flow away from puncture, making proper spotting difficult.

3. **DO NOT** lance on previous puncture site.

4. Use of a capillary tube is not recommended since application of blood with a capillary tube results in scratching and/or abrading the surface of the filter paper which adversely affects test results.

5. Avoid touching area within filter paper circles before blood is applied.

6. **DO NOT** place filter paper in the envelope until thoroughly dry.

7. **INSUFFICIENT DRYING ADVERSELY AFFECTS TEST RESULTS!**
QuantiFERON-TB Gold Plus (QFT-Plus) Collection Procedure

Principle:
To properly collect a blood specimen for QuantiFeron-TB Gold Plus.

Supplies:
1. 4 QFT tubes
2. DHEC form 1335
3. Designated QFT shipper

Collection Procedure:

Precaution: Wear gloves when collecting blood specimens

1. For each patient, collect 1mL of blood by venipuncture directly into each of the QFT-Plus blood collection tubes (4 tubes total).
   a. As 1 mL tubes draw blood relatively slowly, keep the tube on the needle for 2-3 seconds once the tube appears to have completely filled, to ensure that the correct volume is drawn. Note: The black mark on the side of the tubes indicates 1mL fill volume. QFT-Plus blood collection tubes have been validated for volumes from 0.8 mL- 1.2 mL. If the level of blood is outside the indicator line, it is recommended to obtain another blood specimen.
   b. If a butterfly needle is being used to collect blood, a “purge” tube should be used to ensure that the tubing is filled with blood prior to the QFT-Plus tubes being used.
2. Immediately after filling tubes, shake them ten (10) times just firmly enough to ensure the entire inner surface of the tube is coated with blood to dissolve the antigens on the tube walls
   a. Tube temperature should be between 17-25°C at the time of blood tube filling.
   b. Overly vigorous shaking may cause gel disruption and could lead to aberrant results.
3. Label tubes appropriately.
4. The tubes must be transferred to a 37°C ± 1°C incubator as soon as possible, and within 16 hours of collection. Prior to incubation, maintain the tubes at room temperature (22°C ± 5°C). Do not refrigerate or freeze the blood specimens. Note: There are incubators located at specific sites in the regions, or specimens can be placed on courier for incubation, HOWEVER specimens must be received within the acceptable 16 hours post-collection if incubation is to occur at the Public Health Laboratory. If the blood is not incubated immediately after collection, re-mixing of the tubes by inverting 10 times must be performed immediately prior to incubation at 37°C.
5. Incubate the tubes UPRIGHT at 37°C ± 1°C for 16 - 24 hours.
6. After incubation at 37°C, blood collection tubes may be held between 4 - 27°C for up to 3 days before further testing. Specimens should be shipped to the Virology laboratory using the courier system in the designated boxes within the 3-day post-incubation time period.

Specimen Handling:
1. Use a patient label to properly label each QFT-Plus tube.
2. Complete a DHEC 1335. See instructions on back of form for completing. Mark QuantiFeron Gold-Plus and complete incubation start and end time.
**Specimen Preservation and Transport:**

1. Specimens should be shipped and received within 16 hours of collection if not incubated in regions, or within 3 days post-incubation.
2. Place the specimen inside designated QFT-Plus shipper (large white shipper with pink label) labeled to the attention of Virology and ship to maintain tubes in the temperature range of 17 - 27°C until receipt at the Public Health Laboratory.

**Specimen Rejection:**

1. Universal Rejections, See Section 1
2. Use of improper collection techniques and/or under- or over-filled collection tubes.
3. Specimen not incubated within the proper incubation period after collection (specimens under- or over-incubated) or specimens requiring incubation at 37°C are not received at the Public Health Laboratory within 16 hours of collection.
ENTERIC PATHOGENS

Purpose:
To properly collect a stool specimen for the isolation of the following enteric pathogens: E. coli 0157, Salmonella, Shigella, Yersinia, Campylobacter, Vibrio, Staphylococcus, Clostridium perfringens and Bacillus cereus.

Patient Preparation:
No special preparation.

Supplies:
1. Wide-mouthed container.
2. Enteric kit with Cary-Blair transport media. See Page III-1 to order.
3. DHEC form 1335

Collection Precautions:
Wear gloves when collecting stool specimens.

Collection Procedure (Stool):
1. Collect stool in a clean (not necessarily sterile) wide-mouthed container with a tight-fitting lid. These containers must be free of preservatives and detergents.
2. Do not collect specimen from toilet. Avoid contamination with urine.
3. Infant specimens may be collected in a disposable diaper with plastic side facing inside.
4. Collect a walnut sized piece if stool is formed or 5 - 10 mL if stool is liquid.

Cary-Blair Transport media
Formed feces: use tongue depressor or spoon inside the lid to transfer walnut size portion of stool. Liquid feces: use pipette to transfer 5 - 10 mL of liquid stool to the transport media. Replace cap on tube and refrigerate until transported.

Specimen Handling:
1. Place a patient identification label on the transport medium
2. Complete a DHEC form 1335 to accompany specimen. See instructions on back of form. Be sure to complete additional test specific information
   Specimen Type/Source: Mark X by Feces Date Collected
   Organism Suspected: Indicate name of suspected organism
   NOTE: Routine culture includes testing for Salmonella, Shigella, Campylobacter, and E. coli 0157. Request for other specific pathogens must be indicated on the laboratory request form.
   Test Requested: Mark 508 Enteric Culture.

Specimen Preservation and Transport:
1. Ship specimens in transport media in cooler with cold packs to be received at the temperature of 2-8°C. Specimen should be received within 48 hours of collection.
2. See Section IV for appropriate shipping container, packaging and transport instructions.

Specimen Rejection:
1. Specimen too old.
2. Use of improper transport media or transport conditions.
3. Insufficient quantity
4. Universal rejections, See Section I
**NEISSERIA GONORRHOEAE**

**Principle:**
To properly collect an eye culture, rectal culture and oropharyngeal culture for the diagnosis of *Neisseria gonorrhoeae*. To properly collect a cervical, urethral and vaginal culture in cases of assault or sexual abuse.

**Patient Preparation:**
For male urethral culture: The patient should not have voided for at least 1 hour before performing a culture, especially men without a discharge.

**Supplies:**
1. Sterile Dacron or Rayon swab
2. Sterile thin, flexible wire with Dacron or Rayon swab (males)
3. GC culture kit with Transgrow bottle for *N. gonorrhoeae* See Page III-1 to order.
4. DHEC form 1335
5. Speculum (cervical, vaginal)

**Collection Precautions: (All specimens)**
Wear disposable gloves and protective eye wear when collecting and handling specimens.
Note: Collect all specimens Monday - Wednesday. Do not ship for weekend delivery.

**Collection Procedure: (Eye)**
1. Touch a sterile swab to purulent discharge. If necessary, lower eyelid may be pulled down and the swab touched to the conjunctival mucosa.
2. Inoculate Transgrow bottles as described under Inoculation of Transgrow medium

**Collection Procedure: (Rectal)**
1. Have the patient bear down slightly for ease in insertion of swab.
2. Insert a sterile swab approximately 3 cm into the anal canal using lateral pressure to avoid entering any fecal mass. If gross fecal contamination of the swab occurs, it should be discarded into a biohazard container and a repeat specimen obtained.
3. Rotate the swab to specimen crypts just inside the anal ring and allow the swab to remain in the anal area for several seconds for better absorption onto the swab.
4. Inoculate Transgrow bottles as described under Inoculation of Transgrow medium.

**Collection Procedure: (Oropharyngeal [Throat])**
1. Using a tongue blade to hold the tongue down, take a specimen directly from the back of the throat, carefully avoiding contact with teeth, cheeks, gums or tongue when inserting or removing the swab.
2. Rub a sterile swab over the back wall of the throat and tonsillar crypts.
3. Inoculate Transgrow bottles as described under Inoculation of Transgrow Medium.

**Collection Procedure: (Cervical)**
1. Obtain the cervical specimen with the aid of a speculum that has been moistened with water. Other lubricants may contain antibacterial agents.
2. Insert the speculum and if unable to visualize the cervical os, remove excess mucus with swab.
3. Insert another sterile swab into the endocervical canal approximately 2 - 3 cm. Move the swab in a rotary motion for a few seconds to permit absorption of the exudate. If the patient is pregnant, and there has been no vaginal bleeding, insert swab into the endocervix only until the tip is no longer visible and rotate gently for a few seconds).
4. Inoculate Transgrow bottles as described under inoculation of Transgrow medium.
Collection Procedure: (Vaginal) for Children and Hysterectomy Patients Only
1. Insert the speculum.
2. With a sterile swab obtain the specimen from the posterior vaginal vault.
3. Allow a few seconds for absorption of material.
4. If the hymen is intact, a swab of the vaginal orifice will suffice.
5. Inoculate Transgrow bottles as described under Inoculation of Transgrow medium.

Collection Procedure: (Urethral Culture - Females)
1. Massage the urethra against the pubic symphysis from vagina to orifice to express discharge.
2. If no discharge is evident, insert a sterile flexible thin wire swab approximately 2 cm into the urethra and rotate for several seconds.
3. Withdraw swab and inoculate Transgrow bottle as described under Inoculation of Transgrow.

Collection Procedure: (Urethral - Males)
1. Insert a sterile flexible swab with a thin wire shaft 2 - 4 cm into the urethra.
2. Once inserted, rotate the swab gently to ensure contact with all urethral surfaces.
3. Leave inserted for 2 - 3 seconds for better absorption of material.
4. Withdraw swab and inoculate Transgrow bottle as described under Inoculation of Transgrow.

Inoculation of Transgrow Medium
1. Have Transgrow at room temperature; check the expiration date before inoculation.
2. Hold the bottle in an upright position. Remove the cap only when ready to inoculate.
3. Soak up excess moisture in the bottle with the specimen swab and roll the swab from side to side over the entire surface of the medium starting at the bottom of the bottle.
4. Remove swab from bottle and discard into a biohazard container.
5. Recap the bottle tightly.

Specimen Handling:
1. Place label with patient’s name on back of Transgrow bottle where chocolate colored medium is layered. Do not place label on clear side of bottle. This window is needed to observe growth.
2. Complete a DHEC form 1335 to accompany specimen. See instructions on back of form. Be sure to complete test specific information.
   Specimen: Mark X by the appropriate type and write in the site.
   Was Culture Incubated Before Transport?: mark X in the appropriate space(s).
   Test Requested: Mark X in the appropriate space.

Specimen Preservation and Transport:
1. Place the Transgrow bottle in an upright position in an incubator set at 35°C as soon as possible after inoculation. Never refrigerate the medium after inoculation as cold temperature will rapidly kill gonococci. Incubate until ready to ship.
2. If an incubator is not available, make sure culture is shipped on the same day as collected.
3. If the specimen is collected on Friday and cannot be shipped until Monday, incubate over the weekend, but remove first thing Monday morning to prevent contaminant overgrowth.
4. Note: Do not ship for weekend delivery.
5. Ship to be maintained at 15-25°C until received at the Public Health Laboratory.

Specimen Rejection:
1. Transgrow media not used or Transgrow media expired.
2. Specimen in transit for more than 5 days.
3. Universal rejections, See Section I.
DIPHTHERIA

Principle:
To properly collect a throat swab for the culture of *C. diphtheria*

Patient Preparation:
No special preparation

Supplies:
1. Culturette swab kit containing Stuart’s medium. Use form 1323 to order and indicate culturette in blank space on form.
2. DHEC form 1335

Collection Procedure for Throat Swab:
1. Shine a bright light, if possible, over the shoulder of the specimen collector into the oral cavity of the patient so that the swab can be guided to the posterior pharynx.
2. The patient is instructed to tilt his/her head back and breathe deeply.
3. Depress the tongue with a tongue depressor to help visualize the posterior pharynx. Use culturette kit. Do not use calcium alginate swabs.
4. Extend the swab to the back of the throat between the tonsillar pillars and behind the uvula.
5. Have the patient phonate a long aah which will lift the uvula and help to prevent gagging.
6. The tonsillar areas and posterior pharynx should be firmly rubbed with the swab.
7. Care should be taken not to touch the teeth, cheeks, gums or tongue when inserting or removing the swab to minimize contamination with normal mouth flora.
8. After collection, place the swab back into the culturette and break or squeeze the ampule. Note: Notify the DHEC PHL Clinical Microbiology Section (803-896-0803) when a diphtheria specimen is to be collected so that special isolation media can be prepared.

Specimen Handling
1. Place a patient label on a culturette swab kit.
2. Organism suspected: Indicate *Corynebacterium diphtheriae*.

Specimen Preservation and Transport
1. Store and ship culturette at room temperature. Note: Transport within 24 hours. Do not ship for weekend delivery.
2. See Section IV for appropriate shipping container, packaging and transport instructions.

Specimen Rejection
1. Ampule in culturette not crushed.
2. Universal rejections, See Section I.
MYCOBACTERIUM (TB)

**Principle:**
To properly collect a sputum or urine specimen for the diagnosing and monitoring of tuberculosis and other mycobacterial infections.

**Supplies:**
1. (a) Mycobacteriology collection kit (50 mL plastic sputum collection tube) See Page III-1 to order.
   (b) Sterile screw cap container with a round opening of at least 2 inches for urine
2. DHEC form 1335
3. Particulate respirator (PR)

**Collection Procedure: (All Specimens)**
**Wear Disposable Gloves and a Particulate Respirator When Collecting Specimens**

**Patient Preparation: (Sputum)**
1. Explain to patient the importance of how to collect and handle a sputum specimen. Give the patient the sputum collection kit and COLLECTION OF SPUTUM SPECIMENS FOR MYCOBACTERIA (TB) sheet.
2. If the nurse must remain with the patient while he/she is coughing, the nurse should wear a particulate respirator.
3. Have the patient collect an early morning sputum specimen.
4. Ask the patient to breathe deeply, exhale, and then cough deeply. Steam from a hot shower or a boiling kettle may help to stimulate the flow of secretions. Also, drinking several cups nonalcoholic liquids will assist in raising sputum.
5. Patient should brush their teeth and/or rinse with water, not an antiseptic solution before obtaining the sputum specimen to reduce the overgrowth of mouth flora,
6. The patient should submit a series of three (3) sputum specimens over a period of three days (one/day), if specimens are being collected for initial diagnosis.

**Collection Procedure (Sputum)**
1. Remove the cap from the sterile container without touching the inside of the container. This will avoid contamination of the specimen which results in having to submit another specimen.
2. Patient is instructed to take a deep breath, hold it momentarily and cough deeply from the deepest part of the chest. Saliva and nasal secretions which contain few acid-fast bacteria are not to be collected.
3. Instruct the patient to spit the sputum into the appropriate sterile container until at least 5 mL or 1 teaspoon is obtained. Replace cap on the container. A minimum of 5 mL is needed for culture.
4. Avoid soiling the outside of the container. If soiling does occur, wipe with a clean cloth wet with alcohol soap and water, or 1/10 bleach solution, and then wash hands.
5. Sputum specimens should be free of food particles and other extraneous material.
6. Place the cap on plastic tube or sterile container and screw to close tightly.

If patient is to collect sputum in the home, give patient sputum collection and mailing containers and instruction sheet on how to obtain a sputum specimen.

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Collection Procedure: (Urine)
The patient should submit a series of three (3) urine specimens over a period of three days (one/day) if specimens are being collected for initial diagnosis.

Female- midstream voided:
1. Have patient thoroughly clean the urethral area with soap and water.
2. Instruct patient to sit on toilet, and to manually separate labia minora with one hand and keep them separated while voiding the first portion of urine into the toilet.
3. After several mL have passed, have patient collect the midstream portion into the specimen container without stopping the flow of urine. Try to avoid touching the lip or inside of the container with the hand.
4. Have the patient finish voiding into the toilet.
5. Amount of urine needed is 10 mL. Screw cap on plastic container to close tightly.

Male-midstream voided:
1. Clean the glans with soap and water.
2. While holding foreskin retracted, begin voiding.
3. After several mL have passed collect the midstream portion into the appropriate container without stopping flow of urine.
4. Have the patient finish voiding into the toilet.
5. Amount of urine needed is 10 mL. Screw cap on plastic tube to close tightly.

For collection procedures on other specimens see chart on Collection and Shipment of Mycobacterial Specimens.

Specimen Handling:
1. Place a patient identification label on the 50 mL screw capped tube.
2. Complete a DHEC form 1335 to accompany specimen See instructions on back of form. Be sure to complete test specific information:
   - Agent suspected: Enter the suspected agent
   - Specimen source: Mark “X” by the appropriate source.
   - Date & Time Collected:
   NOTE: All clinical specimens should be ordered using Test Code 601. Test Code 602 is reserved exclusively for laboratories that have isolated Mycobacteria and need them identified. Do not request drug susceptibility testing (Test Code 604) when submitting specimens from suspected new cases of tuberculosis. All initial isolates of M. tuberculosis will be tested for susceptibility to INH, rifampin, ethambutol, streptomycin and pyrazinamide.

Specimen Preservation and Transport: Sputum:
1. Refrigerate specimens if shipping is delayed over 24 hours. This will decrease overgrowth of other microorganisms which delay culture results.
2. Ensure that the cap is tightly closed, secure and not cross threaded. Be sure plastic tube is not soiled with sputum or urine.
3. Place the completed DHEC 1335 laboratory form into the side pocket of the biohazard bag. Specimen goes into the large opening of the biohazard bag. If the laboratory form is soiled, the laboratory must autoclave it before it can be handled. Be sure the date the specimen was collected is on the form.
4. Ship to maintain specimens within the range of 2-30°C until received at the Public Health Laboratory.
Specimen Preservation and Transport Urine.
   1. If specimen is urine, ship cold with frozen cold packs in shipper to maintain specimen at 2-8°C until received at the Public Health Laboratory.
   2. Label outside of cooler as Urine for TB testing

Specimen Rejection:
   1. Specimen broken or leaked in transit. Sterile body fluids may be processed with the approval of the Supervisor or Division Director.
   2. Specimen > 5 days old.
   3. Universal rejections, See Section 1

### SPECIMEN COLLECTION FOR CULTURE OF MYCOBACTERIA (TB)

<table>
<thead>
<tr>
<th>SPECIMEN TYPE</th>
<th>TIME</th>
<th>AMOUNT</th>
<th>NUMBER</th>
<th>SPECIAL PROCEDURE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sputum</td>
<td>Early AM On Waking</td>
<td>5-10 mL</td>
<td>Series of 3</td>
<td>Sputum-material coughed up from deep in lungs-not saliva</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>One/Day</td>
<td></td>
</tr>
<tr>
<td>Urine</td>
<td>Early AM</td>
<td>Entire specimen,</td>
<td>Series of 3</td>
<td>Voided midstream specimen collected as aseptically as possible. Transport to lab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>centrifuge 10 mL</td>
<td>One/Day</td>
<td>immediately.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gastric Washing</td>
<td>Early AM</td>
<td>10 mL</td>
<td>1 or more as</td>
<td>No food after midnight. Pass 20-50 mL sterilised distilled water through stomach</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>needed</td>
<td>tube and draw off specimen in sterile tube.</td>
</tr>
<tr>
<td>Biopsy</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Feces</td>
<td></td>
<td>Formed-send walnut</td>
<td>1 or more as</td>
<td>No fixative or preservatives (saline only)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>sized portion</td>
<td>needed</td>
<td></td>
</tr>
<tr>
<td>Sterile body fluids</td>
<td></td>
<td>10 mL</td>
<td>1 or more as</td>
<td></td>
</tr>
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<td>other than blood</td>
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<td></td>
<td>needed</td>
<td></td>
</tr>
<tr>
<td>Swabs of drainage</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>or other material</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Use a Mycobacteriology (TB) collection kit for all specimen types

Use small amt of sterile saline to keep swab moist. Do not use transport media. Swabs are not usually productive specimens for mycobacteria.
CHLAMYDIA/GC & TRICHOMONAS VAGINALIS by GEN-PROBE APTIMA
(Endocervical, Male Urethral, Male/Female Rectal, Pharyngeal, Vaginal, Urine Specimens)

Principle:
To collect and appropriately handle specimens for nucleic acid amplification testing for Chlamydia, Gonorrhoeae, and Trichomonas vaginalis.

Patient Preparation:
See collection procedures below.

Supplies:
1. GC/ Chlamydia/Trichomonas Gen-Probe supplies See Page III-1 to order.
   For Unisex Collection Kit, the blue swab is the specimen collection swab for both male and female specimens.
2. DHEC form 1332

Collection Procedure for Endocervical Swab Specimens (FOR GC/CT/TRICH TESTING):
1. The clinician collects the specimen from the cervical and endocervical area using the APTIMA Unisex Swab (green printing) designed to collect endocervical and urethral specimens for APTIMA/TIGRIS assay. Please use the blue shaft swab for collection.
2. Remove excess mucus from cervical os and surrounding mucosa using cleaning swab (white shaft in package with red printing). Discard this swab!!!
3. Insert specimen collection swab (blue shaft) into endocervical canal.
4. Gently rotate swab clockwise for 10 to 30 seconds in endocervical canal to ensure adequate sampling.
5. Withdraw swab carefully; avoid any contact with vaginal mucosa.
6. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
7. Break off the swab at the scoreline. Use care to avoid splashing contents.
8. Re-cap swab specimen transport tube tightly.
9. Place a label with patient name, date taken, and anatomic site (cervical, Cx) on the tube.
10. Complete a laboratory test requisition for each specimen with the test(s) requested and the appropriate anatomic site (cervical, Cx) indicated on the form.
11. Specimens can be stored in the refrigerator or at room temperature, between 2-30° C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
12. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
13. The specimen is good for 60 days.
14. If longer storage is needed, freeze at -20 to -70 degrees C for up to 12 months after collection.
Collection Procedure for Vaginal Specimens (FOR GC/CT/TRICH TESTING):

1. The clinician collects the specimen from the vaginal area using the APTIMA MTS (Multitest) Swab (orange label, also known as vaginal swab) designed to collect vaginal specimens for APTIMA/TIGRIS assay. **Please use the pink shaft swab for collection.**
2. Carefully insert the swab into vagina about 2 inches inside the opening of the vagina and gently rotate swab 10-30 seconds.
3. Make sure the swab touches the walls of the vagina so that the moisture is absorbed by the swab and then withdraw the swab without touching the skin.
4. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
5. Break off the swab at the scoreline.
6. Tightly screw the cap onto the tube.
7. Place a label with patient name, date taken, and anatomic site (vaginal, vag) on the tube.
8. Complete a laboratory test requisition for each specimen with the test(s) requested and the appropriate anatomic site (vaginal, vag) indicated on the form.
9. Specimens can be stored in the refrigerator or at room temperature, between 2-30°C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
10. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
11. The specimen is good for 60 days.
12. If longer storage is needed, freeze at -20 to -70 degrees C for up to 12 months after collection.

The Aptima Vaginal Swab Specimen Collection Kit (pink shaft swab) should only be used for collection of females ≥ 14 years old and non-pregnant.

Collection Procedure for Male Urethral Specimens (FOR GC/CT TESTING ONLY):

**Patient should not have urinated for at least 1 hour prior to collection.**

1. The clinician collects the specimen from the urethral area using the APTIMA Unisex Swab (green printing) designed to collect endocervical and urethral specimens for APTIMA/TIGRIS assay. **Please use the blue shaft swab for collection.**
2. Insert the specimen collection swab (blue shaft swab in the package with the green printing) 2 to 4 cm into the urethra.
3. Gently rotate the swab clockwise for 2 to 3 seconds in the urethra to ensure adequate sampling.
4. Withdraw the swab carefully.
5. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
6. Carefully break off the swab at the scoreline. Use care to avoid splashing contents.
7. Re-cap the swab specimen transport tightly.
8. Place a label with patient name, date taken, and anatomic site (male urethral) on the tube.
9. Complete a laboratory test requisition (DHEC 1332) for each specimen with the test(s) requested and the appropriate anatomic site (male urethral) indicated on the form.
10. Specimens can be stored in the refrigerator or at room temperature, between 2-30°C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
11. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
12. The specimen is good for 60 days.
13. If longer storage is needed, freeze at -20 to -70 degrees C for up to 12 months after collection.

Collection Procedure for Pharyngeal Specimens (FOR GC/CT TESTING ONLY):

*Since this collection kit is designed to collect endocervical specimens, included is a white shaft “cleaning” swab which is NOT to be used for pharyngeal or rectal specimen collection.*

Collection Procedure for Vaginal Specimens (FOR GC/CT/TRICH TESTING):

1. The clinician collects the specimen from the vaginal area using the APTIMA MTS (Multitest) Swab (orange label, also known as vaginal swab) designed to collect vaginal specimens for APTIMA/TIGRIS assay. Please use the pink shaft swab for collection.
2. Carefully insert the swab into vagina about 2 inches inside the opening of the vagina and gently rotate swab 10-30 seconds.
3. Make sure the swab touches the walls of the vagina so that the moisture is absorbed by the swab and then withdraw the swab without touching the skin.
4. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
5. Break off the swab at the scoreline.
6. Tightly screw the cap onto the tube.
7. Place a label with patient name, date taken, and anatomic site (vaginal, vag) on the tube.
8. Complete a laboratory test requisition for each specimen with the test(s) requested and the appropriate anatomic site (vaginal, vag) indicated on the form.
9. Specimens can be stored in the refrigerator or at room temperature, between 2-30°C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
10. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
11. The specimen is good for 60 days.
12. If longer storage is needed, freeze at -20 to -70 degrees C for up to 12 months after collection.
1. The clinician collects the specimen from the pharyngeal area using the APTIMA Unisex Swab (green printing) designed to collect endocervical and urethral specimens for APTIMA/TIGRIS assay. **Please use the blue shaft swab for collection.**
2. Swab area between the tonsillar pillars and the region posterior to the pillars.
3. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
4. Break off the swab at the scoreline.
5. Place a label with patient name, date taken, and anatomic site (throat) on the tube.
6. Complete a laboratory test requisition for each specimen with the test(s) requested and the appropriate anatomic site (throat) indicated on the DHEC Form 1332.
7. Specimens can be stored in the refrigerator or at room temperature, between 4-30°C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
8. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
9. The specimen is good for 60 days.

**Collection Procedure for Male/Female Rectal Specimens (FOR GC/CT TESTING ONLY):**

Since this collection kit is designed to collect endocervical specimens, included is a white shaft “cleaning” swab which is NOT to be used for pharyngeal or rectal specimen collection.

1. The clinician collects the specimen from the rectal area using the APTIMA Unisex Swab (green printing) designed to collect endocervical and urethral specimens for APTIMA/TIGRIS assay. **Please use the blue shaft swab for collection.**
2. **Asymptomatic and/or Symptomatic Males/Females:** moisten swab with sterile saline/tap water and insert into anus and rectum approximately 2-5 cm (1 to 2 inches) and rotate 3-8 times. **NOTE: it is ok to have some fecal contamination that appears as a brown discoloration but NO frank fecal material.**
3. Remove foil cap from swab specimen transport tube and immediately place specimen collection swab into the transport tube.
4. Break off the swab at the scoreline.
5. Place a label with patient name, date taken, and anatomic site (rectal, rec) on the tube.
6. Complete a laboratory test requisition for each specimen with the test(s) requested and the appropriate anatomic site (rectal, rec) indicated on the form.
7. Specimens can be stored in the refrigerator or at room temperature, between 4-30°C, prior to transport to the Public Health Laboratory, 8231 Parklane Road, Columbia, SC 29223.
8. Specimens need to be stored in an upright position in a rack so that the buffer is in contact with the swab.
9. The specimen is good for 60 days.

**Collection Procedure for Male and Female Urine Specimens (FOR GC/CT/TRICH TESTING):**

Patient should not have urinated for at least 1 hour prior to specimen collection.

1. Direct patient to provide first-catch urine (approximately 20 to 30 mL of initial urine stream) into urine collection cup free of any preservatives. Collection of larger volumes of urine may result in specimen dilution that may reduce test sensitivity. Female patients should not cleanse labial area prior to providing specimen.
2. Remove cap from urine specimen transport tube and transfer 2 mL of urine into urine specimen transport tube using disposable pipette provided. The correct volume of urine has been added when fluid level is between black fill lines on urine specimen transport tube label.
3. Re-cap urine specimen transport tube tightly. This is now known as the “processed urine specimen.”
4. See Specimen Transport and Storage below.
Specimen Handling:
Complete DHEC form 1332 to accompany specimen. See instructions on back for completing. Be sure to complete test specific information.

Specimen Preservation and Transport
A. Swab
1. After collection, transport and store swab in swab specimen transport tube at 2°C to 30°C until tested.
2. Specimens must be assayed within 60 days of collection. If longer storage is needed, freeze at -20°C to -70°C for up to 12 months after collection.

B. Urine
1. After collection, transport the processed urine specimens in the GEN-PROBE Aptima Assay urine specimen transport tube at 2°C to 30°C and store at 2°C to 30°C until tested. Processed urine specimens should be assayed with the APTIMA Assay within 30 days of collection. If longer storage is needed, freeze at -20°C -or-70°C for up to 90 days after collection.
2. Urine specimens that are still in primary collection container must be transported to lab at 2°C to 30°C. Transfer urine specimen into APTIMA Assay urine specimen transport tube within 24 hours of collection. Store at 2°C to 30°C and test within 30 days.
3. See Section IV for appropriate shipping container, packaging and transport instructions.

Specimen Rejection:
1. No swab in tube, 2 swabs in tube, or improper (non-blue) swab used.
2. Universal rejections, See Section I.
3. Note: specimens collected with this system cannot be used for culture.

References:
1. Probtect Swab Specimen Collection and Handling by Campbell, D., SFDPH Micro Lab and Engelman, J., M.D., City Clinic, 1/2002.
2. APTIMA Swab Specimen Collection Guide; Gen-Probe Incorporated, San Diego, CA 92121.
3. City and County of San Francisco, Dept. of Public Health, City Clinic Branch Laboratory, revised 10/09.
Section IV
Transporting and Shipping Infectious Substances

A. Introduction

Patient specimens from most of the SC Health Departments and many of the SC hospitals are transported to the SC DHEC Public Health Laboratory through a DHEC contracted courier system. This courier system picks up and delivers courier mail to over 60 DHEC health departments and locations throughout the state every evening for arrival at the Public Health Laboratory the next morning.

For the protection of employees and the public, patient specimens and infectious substances must be properly packaged and labeled. As packages delivered using this courier system are transported in commerce, they must be packaged to meet all DOT requirements for shipping infectious substances. Failure to follow these regulations can result in injury, exposure, and/or fines.

B. Regulatory Requirements

There are three regulatory entities regarding the shipping of hazardous materials; the International Air Transporters Association (IATA), the United States Department of Transportation (USDOT), and the United States Postal Service (USPS). According to regulations, it is the shipper’s responsibility to properly package shipments of infectious substances and hazardous materials.

The International Air Transporters Association (IATA) is a private organization whose regulations only apply to air transport by IATA member airlines. All major airlines are members of IATA and follow the IATA Dangerous Goods Regulations taken from the International Civil Aviation Organization (ICAO) Technical Instructions for the Safe Transport of Dangerous Goods by Air.

The United States Department of Transportation (US DOT) is a government agency that regulates commercial transport. Commercial transport takes place when money is exchanged for a good or service. All modes of transportation, ground, air, and water, fall under DOT regulations. US DOT regulations are located in the Code of Federal Regulations 49 CFR 173. Updates to these regulations require congressional approval and are not frequently updated.

The United States Postal Service (USPS), has their own regulations found in the domestic mail manual. As one federal agency cannot regulate another federal agency, the USPS is not required to follow US DOT regulations. As an example, the Postal Service can transport cylindrical shippers while a private courier, like Fedex, cannot.

In addition to these regulations, private couriers can have additional regulations. As an example, Federal Express requires that a shippers declaration for Dangerous Goods be typed and not hand written.

The US Department of Transportation (DOT) and the US Postal Service (USPS) harmonized their regulations with the International Air Transporter Association (IATA) regulations in 2006. Therefore, if infectious substance is packaged and labeled to meet the IATA regulations, the package will meet or exceed the requirements for US DOT and the US Postal Service. In addition to providing uniformity, this harmonization allowed the regulations to be more adaptive. As IATA is a private organization, it has the ability to change its regulations without congressional approval.
C. **Training Requirements**

All employees who are a part of any step of classifying, packaging, labeling, marking, completing the paperwork, or transporting the specimen must be properly trained to package and ship infectious substances. Training records must be retained for a **minimum of thirty-six months**. Retraining must be completed **every two years** from the date of completion for IATA regulations and every three years for DOT regulations.

The training must include:

- An overview of the regulatory requirements
- Security awareness training
- Function specific training on the activities the employee will be responsible for, such as classification of infectious substances, packaging, labeling the outside container and completing shipping documentation.
- Safety training to include understanding the hazards of the infectious agent, safe handling and emergency response procedures.

The employer must certify the employees training as adequate and maintain a record of training which includes:

- The individual’s name
- The most recent training completion date
- A description, copy or reference to training materials used
- The name and address of the organization providing the training
- A test, which was completed satisfactorily, to verify the employee understood the training.
D. **Exemptions**

**Exempted Materials**

The following items are exempt from the shipping regulations for infectious substances, but must be packaged to avoid leaking during shipping and may require a special label.

- Specimens/samples in which all pathogens have been neutralized or inactivated
- Specimens/samples known to not contain infectious substances
- Specimens/samples which only contain micro-organisms which are non-pathogenic for humans and animals
- Dried blood spots and fecal occult blood samples
- Environmental samples (food and water) that are not considered to pose a significant health risk
- Organs for transplant and blood for transfusion

**Private Courier Exemptions**

An exemption called the “materials of trade exemption,” located at 49 CFR 173.6, is commonly used by hospital and DHEC employees. This exemption has multiple parts, but the part most useful for the transport of infectious substances is the following: “a hazardous material transported on a motor vehicle, by a private carrier in direct support of a principle means of business that is other than transportation by motor vehicle.” This exemption does not apply to all hazard classes and there are quantity limits to those materials that are allowed. For infectious substances, this exemption only applies to category B samples.

So, a hospital courier or DHEC employee that transports samples to the health department, can use this exemption, because their principle business is not the transportation of samples but the care and treatment of patients or the community. Therefore, these regulations listed above do not apply to the transport of category B infectious substances transported by a hospital courier or DHEC employee transporting samples to a health department.

However, in order to protect the safety of the employee and the public, DHEC employees and other entities shipping specimens through the DHEC contracted courier must follow all of the regulations for proper shipping described in further pages. Additionally, secure the package in the vehicle as far away as possible from the driver as possible, preferably in the trunk if available. If there is an accident, emergency responders need to know that infectious substances are in the package.
E. Definitions:

BIOLOGICAL PRODUCTS: Are those products derived from living organisms which are manufactured and distributed in accordance with the requirements of appropriate national authorities, which may have special licensing requirements, and are used either for prevention, treatment, or diagnosis of disease in humans or animals, or for development, experimental or investigational purposes related thereto. They include, but are not limited to, finished or unfinished products such as vaccines.

CARBON DIOXIDE, SOLID (DRY ICE): Carbon dioxide, solid (dry ice) is produced by expanding liquid carbon dioxide to vapor and “snow” in presses that compact the product into blocks. It is used primarily for cooling and due to its very low temperature (about -79 C) can cause sever burns to skin upon direct contact. When Carbon dioxide, solid (dry ice) converts (sublimates) directly to gaseous carbon dioxide it takes in heat from its surroundings. The resulting gas is heavier than air and can cause suffocation in confined areas as it displaces air. Packages containing Carbon dioxide, solid (dry ice) must be designed and constructed so as to prevent build-up of pressure due to the release of carbon dioxide gas.

CONSIGNEE: Any person, organization or government which is entitled to take delivery of a consignment.

CULTURES: Cultures are the result of a process by which pathogens are intentionally propagated. This definition does not include patient specimens as defined in 3.6.2.1.4.

DANGEROUS GOODS: Articles or substances which are capable of posing a risk to health, safety, property or the environment and which are shown in the list of dangerous goods in these Regulations or which are classified according to the Regulations.

EXCEPTION: A provision in these Regulations which excludes a specific item of dangerous goods from the requirements normally applicable to that item.

EXEMPTION: Authorization issued by an appropriate national authority of all States concerned providing relief from the provisions of these Regulations.

INFECTIOUS SUBSTANCES: Are substances which are known or are reasonably expected to contain pathogens. Pathogens are defined as microorganisms (including bacteria, viruses, rickettsiae, parasites, fungi) and other agents such as prions, which can cause disease in humans or animals.

INNER RECEPTACLE: Are receptacles which require an outer packaging in order to perform their containment function.
OVERPACK: An enclosure used by a single shipper to contain one or more packages and to form one handling unit for convenience of handling and stowage. Dangerous goods packages contained in the overpack must be properly packed, marked, labeled and in proper condition as required by these Regulations. For cooling purposes, an overpack may contain Carbon dioxide, solid (dry ice), provided that the overpack meets the requirements of Packing Instruction 954. (A Unit Load Device is not included in this definition.)

PACKAGE: (Non-Radioactive Material). The complete product of the packing operation consisting of the packaging and contents prepared for transport.

PACKAGING: (Non-Radioactive Material). Receptacles and any other components or materials necessary for the receptacle to perform its containment function and to ensure compliance with the minimum packing requirements of these Regulations.

PACKING: The art and operation by which articles or substances are enveloped in wrappings and/or enclosed in packaging or otherwise secured.

PATIENT SPECIMENS are those collected directly from humans or animals, including, but not limited to, excreta, secreta, blood and its components, tissue and tissue fluid swabs, and body parts being transported for purposes such as research, diagnosis, investigational activities, disease treatment and prevention.

PROPER SHIPPING NAME: The name to be used to describe a particular article or substance in all shipping documents and notifications and, where appropriate, on packaging.

RECEPACTACLE: A containment vessel, including closures, for receiving and holding substances or articles.

SELECT AGENT: microorganisms or toxins, identified by a panel of experts, which could be used for bioterrorism. A complete list of select agents and toxins may be found on the Select Agent Program’s web page http://www.cdc.gov/od/sap/docs/salist.pdf

SHIPMENT: The specific movement of a consignment from origin to destination.

UN NUMBER: The four digit number assigned by the United Nations Committee of Experts on the Transport of Dangerous Goods to identify a substance or a particular group of substances. (The prefix “UN” must always be used in conjunction with these numbers.)
F. Instructions for Packaging Infection Substances

Step 1: Classifying Infectious Substances

Infectious substances are divided into 2 categories – A and B. If you need assistance with classifying the materials you are shipping, please call the testing section which performs the test you are requesting.

Category A
An infectious substance which is transported in a form that, when exposure to it occurs, is capable of causing permanent disability, life-threatening or fatal disease in otherwise healthy humans or animals.

Indicative examples of substances that meet these criteria are given in Table 3.6.D from the IATA Dangerous Goods Regulation (see next page). This table is not exhaustive. New or emerging pathogens, which do not appear in the table but which meet the same criteria must be assigned to category A.

In this table, organisms listed with the words “cultures only” indicate that clinical specimens known to contain that organism can be shipped as category B. As an example, Ebola is not listed with “cultures only.” Therefore specimens known to contain Ebola must be shipped as Category A.

Other Examples of Category A infectious substances:
- Known culture of a Select Agent
- Known culture of \textit{Escherichia coli} (toxigenic)
- Known culture of \textit{Neisseria meningitidis}
- Known culture of \textit{Mycobacterium tuberculosis}
- Samples or cultures suspected to be Select Agents or BSL-3 organisms.
  (As an additional precaution and requested by the PHL)

Category B
An infectious substance which does not meet the criteria for inclusion in Category A.

Examples of Category B infectious substances:
- Most cultures and patient specimens shipped to the Public Health Laboratory
- A swab placed in a genprobe bottle (would not meet the IATA definition of a culture)
### Table 3.6.D from IATA Dangerous Goods Regulations

**Indicative Examples of Infectious Substances Included in Category A in Any Form Unless Otherwise Indicted** (January 2021)

[NOTE: “Select Agents or Toxins” are shown in red font]

<table>
<thead>
<tr>
<th>Vaccines</th>
<th>Vaccines</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Bacillus anthracis</em> (cultures only)</td>
<td><em>Japanese Encephalitis virus</em> (cultures only)</td>
</tr>
<tr>
<td><em>Brucella abortus</em> (cultures only)</td>
<td><em>Junin virus</em></td>
</tr>
<tr>
<td><em>Brucella melitensis</em> (cultures only)</td>
<td><em>Kyasanur Forest disease virus</em></td>
</tr>
<tr>
<td><em>Brucella suis</em> (cultures only)</td>
<td><em>Lassa virus</em></td>
</tr>
<tr>
<td><em>Burkholderia mallei - Pseudomonas mallei</em> – Glanders (cultures only)</td>
<td><em>Machupo virus</em></td>
</tr>
<tr>
<td><em>Burkholderia pseudomallei</em> – <em>Pseudomonas pseudomallei</em> (cultures only)</td>
<td><em>Marburg virus</em></td>
</tr>
<tr>
<td><em>Chlamydia psittaci</em> - avian strains (cultures only)</td>
<td><em>Monkeypox virus</em></td>
</tr>
<tr>
<td><em>Clostridium botulinum</em> (cultures only)</td>
<td><em>Mycobacterium tuberculosis</em> (cultures only)</td>
</tr>
<tr>
<td><em>Coccidioides immitis</em> (cultures only)</td>
<td><em>Nipah virus</em></td>
</tr>
<tr>
<td><em>Coxiella burnetii</em> (cultures only)</td>
<td><em>Omsk hemorrhagic fever virus</em></td>
</tr>
<tr>
<td><em>Crimean-Congo hemorrhagic fever virus</em></td>
<td>*Poliovirus (cultures only)</td>
</tr>
<tr>
<td><em>Dengue virus</em> (cultures only)</td>
<td>*Rabies virus (cultures only)</td>
</tr>
<tr>
<td><em>Eastern equine encephalitis virus</em> (cultures only)</td>
<td><em>Rickettsia prowazekii</em> (cultures only)</td>
</tr>
<tr>
<td><em>Escherichia coli</em>, verotoxigenic (cultures only)</td>
<td><em>Rickettsia rickettsii</em> (cultures only)</td>
</tr>
<tr>
<td><em>Ebola virus</em></td>
<td><em>Rift Valley fever virus</em> (cultures only)</td>
</tr>
<tr>
<td><em>Flexal virus</em></td>
<td>*Russian spring-summer encephalitis virus (cultures only)</td>
</tr>
<tr>
<td><em>Francisella tularensis</em> (cultures only)</td>
<td><em>Sabia virus</em></td>
</tr>
<tr>
<td><em>Guanarito virus</em></td>
<td><em>Shigella dysenteriae</em> type 1 (cultures only)</td>
</tr>
<tr>
<td><em>Hantaan virus</em></td>
<td><em>Tick-borne encephalitis virus</em> (cultures only)</td>
</tr>
<tr>
<td><em>Hantavirus causing hemorrhagic fever with renal syndrome</em></td>
<td><em>Variola virus</em></td>
</tr>
<tr>
<td><em>Hendra virus</em></td>
<td><em>Venezuelan equine encephalitis virus</em> (cultures only)</td>
</tr>
<tr>
<td><em>Hepatitis B virus</em> (cultures only)</td>
<td><em>West Nile virus</em></td>
</tr>
<tr>
<td><em>Herpes B virus</em> (cultures only)</td>
<td><em>Yellow fever virus</em> (cultures only)</td>
</tr>
<tr>
<td><em>Human immunodeficiency virus (HIV)</em> (cultures only)</td>
<td><em>Yersinia pestis</em></td>
</tr>
<tr>
<td><em>Highly pathogenic avian influenza virus</em> (cultures only)</td>
<td></td>
</tr>
</tbody>
</table>
Examples of Classifying Infectious Substances

<table>
<thead>
<tr>
<th>Material</th>
<th>Category A</th>
<th>Category B</th>
</tr>
</thead>
<tbody>
<tr>
<td>Culture of <em>Mycobacterium tuberculosis</em></td>
<td>X</td>
<td></td>
</tr>
<tr>
<td>Sputum from a person infected with <em>Mycobacterium tuberculosis</em></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Known culture of <em>Salmonella spp.</em></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Known culture of <em>Bacillus anthracis</em></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Suspected culture of <em>Bacillus anthracis</em></td>
<td>Safer Choice</td>
<td>Technically Correct</td>
</tr>
<tr>
<td>Tube of blood from a person known to be infected with <em>Bacillus anthracis</em></td>
<td>Safer Choice</td>
<td>Technically Correct</td>
</tr>
<tr>
<td>Tube of blood drawn from patient infected with Ebola virus</td>
<td>X</td>
<td></td>
</tr>
<tr>
<td>Animal head shipped for rabies testing</td>
<td></td>
<td>X</td>
</tr>
</tbody>
</table>

**Step 2: Proper Shipping Names and UN Numbers**

Once the proper category is determined, use the corresponding UN number and proper shipping name for your package. Both of these items are required on the outer packaging and are used in later steps. The proper shipping name must be spelled exactly as seen here.

<table>
<thead>
<tr>
<th>Classification</th>
<th>Proper shipping name</th>
<th>UN number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Infectious substance, Category A</td>
<td>“Infectious substance, affecting humans” (technical name)</td>
<td>UN 2814</td>
</tr>
<tr>
<td>Infectious substance, Category B</td>
<td>“Biological substance, Category B”</td>
<td>UN 3373</td>
</tr>
</tbody>
</table>

For category A, notice the parenthesis at the end. In these parentheses, a technical name must be entered. Abbreviations and non-standard formatting are not allowed. So, no italics for scientific names. Examples could include; “Escherichia coli” and “Neisseria meningitidis”.

If you are shipping a sample suspected, but not confirmed, to be a category A infectious substance, and have elected to ship the material as a category A as an additional safety measure, the packing name must use the text “Infectious Substance, Affecting Humans (suspected category A infectious substance).”
**Step 3: Packing Selection and Requirements**

Caution: shipping requirements for your specimen may have recently changed. Check the test section in the *Public Health Laboratory Services Guide* to ensure proper shipping conditions and see page IV-15 for temperature controlled packaging instructions.

1. **Packaging Selection**

   **Package Construction**
   Not all packages are acceptable for shipping infectious substances. Packages must follow strict DOT and IATA regulations regarding their size, shape, construction materials, and markings. Approved packaging configurations and requirements are defined by the DOT in 49 CFR 172 and 173, and by IATA in the Dangerous Goods regulations, section 5, packaging instructions 620 and 650.

   **Package Performance Testing**
   Additionally, packages must follow strict manufacturing standards and performance. Performance tests simulate the potential conditions the package may encounter during transit and test the package’s ability to contain the hazardous material while enduring stresses like drops, leaks, pressurized atmospheres, and stacking loads. Standards for specific performance tests are located in 49 CFR 178 for the DOT and in the Dangerous Goods Regulations, Section 6 for IATA. Performance tests must be documented and the records must be made available to inspectors upon request.

   **Packaging Options**
   Performance packaging accepted by the DHEC contracted courier system, also known as a shipper, falls into three general categories: UN certified shippers, PHL approved shippers, and sender verified packaging. Do not mix and match parts of packages. The package has been certified as a unit. Mixing and matching parts invalidates the UN certification.

   1. **UN certified shippers** have been tested by the manufacturer and certified to meet all performance requirements for IATA and DOT. This certification mark (seen right) indicates that the package is UN certified.

      UN certified shippers are not certified for all hazardous materials. After the UN mark will be a series of letters and numbers. As an example, 4G / CLASS 6.2 / 20 USA /. Pay special attention to the second set of information. In this example “CLASS 6.2” Class 6.2 is the class which contains infectious substances. If a box said “3” or “8” in this location, the box would not be appropriate for shipping infectious substances.

      A UN certified shipper is certified for both Category A and B infectious substances. UN certified shippers, also meet all of the requirements for air transportation, and are universally accepted by national commercial carriers like FedEx or UPS.
Berlin – HMS-69110
- UN certified for Category A and B shipping.
- Can go by ground or air transport.
- See page IV-15 for temperature controlled packaging instructions.

Infecon 5500
- UN certified for Category A and B shipping.
- Can go by ground or air transport.
- See page IV-15 for temperature controlled packaging instructions.

Infecon 5000
- UN certified for Category A and B shipping.
- Can go by ground or air transport.
- See page IV-15 for temperature controlled packaging instructions.

2. **PHL approved shippers**, indicated by the mark to the right, are shippers provided by the PHL, for which the PHL has conducted performance testing. However, the Public Health Laboratory has only conducted the testing needed for ground transportation of Category B infectious substances. Do not use them for Category A shipments and do not offer this package to a national commercial carrier like FedEx or UPS as it has not met all the requirements for air transportation.

ThermoSafe – Sonoco #311
- PHL approved for Category B shipping.
- Ground transport only.
- See page IV-15 for temperature controlled packaging instructions.
3. **Sender Verified Packaging** may be used if the shipper meets all DOT / IATA regulations and/or has been performance tested by your entity or by the manufacturer. If this option is selected, your entity will be responsible for providing USDOT inspectors with performance test results and/or a statement from the manufacturer.

**b. Triple Packaging**

The safe transport of infectious substances is based on “triple-packing.” As an example, a primary sample container, in a secondary container, in an outer shipper, offering three layers of protection.

**Primary Receptacle**

- Is the container (e.g., tube vial, bottle) that holds the specimen.
- Must be securely sealed and leak proof (screw top tubes must have a piece of waterproof tape around the top to prevent the top from coming loose in transit).
- Must be surrounded by absorbent material capable of taking up the entire liquid contents.

**NOTE:** Remember, there must always be adequate absorbent materials next to the primary receptacle to contain all liquid contents should the container leak. The PHL provided absorbent pads are rated to absorb 50ml.

- Must be packed in the secondary receptacle in such a way that it will not break.
Secondary Packaging

- Is the receptacle into which a primary receptacle and the absorbent and cushioning material are placed.
- Must be leak proof and securely sealed.
- Must be placed in the outer packaging so that it does not move.
- Must have a biohazard symbol.
- Never put dry ice inside a secondary container. The container may rupture because of trapped gasses.
- Never put paperwork inside the secondary container.

*Note: For PHIL approved containers, a ziplock biohazard bag may serve as the secondary receptacle for a patient specimen if transport is by ground with the DHEC courier system.*

Outer Packaging

- Is the receptacle into which the secondary receptacle, along with cushioning materials, is placed.
- Must be rigid.
- Bears all required markings and labels.
- At least one surface of the outer packaging must have a minimum dimension of 4 inches x 4 inches.
- Itemized list of contents, requisition forms, and other paperwork is placed here next to the secondary container.
- Dry ice and cool pack are placed here next to the secondary container.
- Seal the package with clear shipping tape. Do NOT use excessive tape to close the outside container.
- Use caution when opening outer packages. Cut the tape instead of pulling the tape to open the package. Pulling the tape can rip or tear reusable package. Also be careful not to cut the box, specifically cardboard closing tabs.

Over Packaging

- Is not required for all packages.
- Is a larger box containing one, or more, smaller completely packaged and labeled shippers.
- Must bear all the same marks and labels required by the contents of the shippers it contains and the word “Overpack.”
- Over packs may be needed if more, surface area is needed on a shipper to accommodate the required marks and labels.
**Closure Instructions**

When using a UN certified or PHL approved shipper, you must follow the manufacturer’s instructions for closing the package. If the closure instructions specify an order to close the flaps of the box, that order must be followed. Failure to follow the manufacture's closure instructions can result in a DOT fine. It is important to retain a copy of these instructions both for reference as needed and if requested by a DOT inspector.

**Quantity Limits**

For Category B infectious substances, regulations allow:
- Up to 1 liter per primary receptacle
- Up to 4 liters per outer packaging.

For Category A infectious substances, regulations allow:
- Up to 50ml or 50g per shipper on a passenger aircraft.
- Up to 4 liters per shipper on a cargo aircraft.
c. Shipping at Controlled Temperatures

**Caution:** Shipping requirements for your specimen may have recently changed. Check the test section in the Public Health Laboratory Services Guide to ensure proper shipping conditions and specimen integrity.

Generally, only three controlled shipping temperatures are used to transport specimens to the Public Health Laboratory. Samples are shipped “frozen” (≤-20°C), “refrigerated” (2-8°C), or at controlled room temperature (CRT) (15-25°C). Specimens received outside of appropriate ranges may be rejected for testing. Please carefully follow the instructions below to ensure sample integrity by following proper controlled temperature shipping.

**Caution:** It is the shipper’s responsibility to ensure proper temperature control of samples during transit where necessary. Guidance provided below is intended to assist shippers with the selection and use of materials provided through the PHL. The use of equivalent materials is acceptable. Please see, Ordering Supplies/Forms/Shipping Containers, Section III, p. 1, for information on receiving free shipping materials to submit specimens to the PHL.

**General Instructions**

**Shipping Frozen (≤ -20°C)**
- Follow all infectious substance shipping instructions.
- With UN certified shippers, do not alter the shippers packaging instructions to accommodate dry ice.
- Place dry ice between the secondary container and the outer packaging.
- **Never place dry ice inside a secondary container.**
- Generally, 6 pounds is sufficient for 24 hour shipments.

**Refrigerated Shipping (2-8°C)**
- Follow all infectious substance shipping instructions.
- With UN certified shippers, do not alter the shippers packaging instructions to accommodate temperature control materials packs.
- The use of phase changing materials (PCM) is preferred over water based gel packs. This avoids the need for pre-conditioning (aka “bench time”) to prevent freezing the sample at the beginning of transit (aka “cold shock”).
- **DO NOT USE WET ICE.**
- Gel packs should be frozen flat and allowed to equilibrate for 24 hours before use.
- Pre-condition water based gel packs by allowing them to sit at room temperature (“bench time”) until their contents begin to become fluid or they begin to sweat. This prevents “cold shocking” and freezing the sample in the beginning of transit.
- The sender will need to determine the number of packs and configurations of materials needed. See instructions below for shipper configurations for PHL provided materials.
Controlled Room Temperature (CRT) (15-25°C)

- Follow all infectious substance shipping instructions.
- With UN certified shippers, do not alter the shippers packaging instructions to accommodate temperature control materials packs.
- The use of phase changing materials (PCM) is preferred over water based gel packs. This avoids the need for pre-conditioning (aka “bench time”) to prevent freezing the sample at the beginning of transit (aka “cold shock”).
- Gel packs should be frozen flat and allowed to equilibrate for 24 hours before use.
- Pre-condition water based gel packs by allowing them to sit at room temperature (“bench time”) until their contents begin to become fluid or they begin to sweat. This prevents “cold shocking” and freezing the sample in the beginning of transit.
- Controlled Room temperature (CRT) gel packs should be allowed to equilibrate to room temperature for 24 hours before use.
- The sender will need to determine the number of packs and configurations of materials needed. See instructions below for shipper configurations for PHL provided materials.

**Instructions for Using Shippers and Temperature Control Materials Provided by the PHL**

1. Select an appropriate shipper for your situation then follow the packing instructions below.

<table>
<thead>
<tr>
<th>Shipper Name</th>
<th>Frozen &lt; -20°C</th>
<th>Refrigerated 2-8°C</th>
<th>CRT 15-25°C</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sonoco ThermoSafe #311 &quot;Head Box&quot;</strong></td>
<td>24 hours Year Round</td>
<td>24 hours Summer only</td>
<td>24 hours Summer only</td>
</tr>
<tr>
<td><strong>Uline #S-7359</strong></td>
<td>24 hours Year Round</td>
<td>24 hours Summer only</td>
<td>24 hours Summer only</td>
</tr>
<tr>
<td><strong>Berlin (UN Certified) #HMS-69110</strong></td>
<td>24 hours Year Round</td>
<td><strong>Do Not Use</strong></td>
<td><strong>Do Not Use</strong></td>
</tr>
<tr>
<td><strong>Infecon 5000 (UN Certified)</strong></td>
<td>24 hours Year Round</td>
<td>24 hours Summer only</td>
<td>24 hours Summer only</td>
</tr>
<tr>
<td><strong>Infecon 5500 (UN Certified)</strong></td>
<td>24 hours Year Round</td>
<td>24 hours Summer only</td>
<td>24 hours Summer only</td>
</tr>
</tbody>
</table>
Shipping Configurations - Sonoco ThermoSafe

Shipping Frozen (≤ -20°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place the secondary container on the dry ice.

Refrigerated Shipping (2-8°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 4 Frozen - 24oz packs (one on each side)

Controlled Room Temperature Shipping (15-25°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 2 Frozen - 24oz packs on the bottom
  - 2 CRT - 24oz packs on top of those
  - 6 CRT - 24oz packs on the sides
  - 1 CRT - 24oz pack on the top
Shipping Configurations – Uline

Shipping Frozen (≤ -20°C)
- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place the secondary container on the dry ice.

Refrigerated Shipping (2-8°C)
- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 1 frozen – 24oz pack on the bottom
  - 2 frozen – 24oz packs, one on each long side

Controlled Room Temperature Shipping (15-25°C)
- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 1 Frozen - 24oz pack on the bottom
  - 1 CRT – 24oz pack on top of that
  - 4 CRT - 24oz packs of the sizes
  - 1 CRT – 24oz pack on the top
Shipping Configurations – Infecon 5000

Shipping Frozen (≤ -20°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place dry ice around the secondary container.

Refrigerated Shipping (2-8°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - Place the cardboard insert in first
  - Insert the secondary container
  - 2 frozen – 24oz packs, one on each long side
  - 1 frozen – 24oz pack on the top

Controlled Room Temperature Shipping (15-25°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 1 Frozen - 24oz pack on the bottom
  - 1 CRT – 24oz pack on top of that.
  - Place the secondary container and cardboard insert next.
  - 4 CRT - 24oz packs on the sides
  - 1 CRT - 24oz pack on the top
Shipping Configurations – Infecon 5500

Shipping Frozen (≤ -20°C)

- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place dry ice around the secondary container.

Refrigerated Shipping (2-8°C)

- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - Place the cardboard insert in first
  - Insert the secondary container
  - 2 frozen – 24oz packs, in an L-shape around the sides
  - 1 frozen – 24oz pack on the top

Controlled Room Temperature Shipping (15-25°C)

- Follow all infectious substance shipping instructions and...
- See section for general instructions for shipping items at this temperature on the previous page.
- Place temperature controlled materials as follows:
  - 1 Frozen - 24oz pack on the bottom
  - 1 CRT – 24oz pack on top of that.
  - Place the secondary container and cardboard insert next.
  - 4 CRT - 24oz packs on the sides
  - 1 CRT - 24oz pack on the top
Shipping Configurations – Berlin

Shipping Frozen (≤ -20°C)
- Follow all infectious substance shipping instructions and…
- See section for general instructions for shipping items at this temperature on the previous page.
- Place dry ice around the secondary container.

Shipping Refrigerated (2-8°C)
- Do not use for 2-8°C Shipping.
- Cannot sustain temperature for 24 hours.

Controlled Room Temperature Shipping (15-25°C)
- Do not use for 15-25°C Shipping.
- Cannot sustain temperature for 24 hours.
**Step 4: Shipping Paperwork**

The following papers must accompany each package containing infectious substances:
- Itemized list of contents
- Paperwork related to sample testing (requisition forms, results, etc.)
- Declaration of Dangerous Goods (for shipments of Category A or dry ice)

**Itemized list of Contents**

All packages must be accompanied by an itemized list of contents. This document contains:
- To and From Address
- An Emergency Contact Name and Telephone
- The kind of specimens with a brief description
- The number and total volume of the samples
- The proper shipping classification for the hazards

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**Itemized List Used by the Public Health Laboratory**

**Shipped from:**
SC DHEC Public Health Laboratory  
8231 Parklane Road  
Columbia, SC 29223  
Emergency Contacts: Andrea Causey  
Emergency Telephone: (803) 767-8110

**Shipped to:**

---

**Shipping Temperature**

- Controlled Room Temperature (15-25°C)  
- Refrigerated (2-8°C)  
- Frozen / Dry Ice (< -20°C)

**Next day delivery required**

- No  
- Yes

<table>
<thead>
<tr>
<th>Specimen or Culture</th>
<th>Number of tubes or plates</th>
<th>Volume in each tube or plate</th>
<th>Total volume</th>
<th>Proper shipping classification (circle only one)</th>
</tr>
</thead>
</table>
| Examples – culture slant of Salmonella | | | | Infectious substance, category A or  
| | | | | Infectious substance, category B |

IV-21
Shippers Declaration for Dangerous Goods

- Required for packages containing a Category A infectious substance and/or dry ice.
- This is a legal document that declares to the courier the hazardous contents in the package.

- A pdf fillable version of this document is available at www.iata.org/whatwedo/cargo/dgr/Documents/Shippers-Declaration-Open-Format-Fillable.pdf
- Small amounts (≤ 30ml) of sample preservative which are classified as Class 3 (flammable) and/or Class 8 (corrosive) materials are not required to be listed on the declaration.
- Use the proper shipping name and UN number as determined in previous steps.
- The document must be attached to the outside (usually the top) of the package in a document pouch. The entire pouch must fit flat on one side of the package.
- The document must be completed in triplicate, each as an original, with the red stripe down each side of the paper. Two copies are given to the transporter and one copy is kept for your files.
- These documents must be kept by the sender for a minimum of two years from the date of the shipment.

**NOTE** - Federal Express does not accept hand written Shipper’s Declarations. Refer to www.fedex.com/us for details regarding acceptable electronic methods to prepare this form.
Step 5: Marks and Labels

The following marks and labels must be present, complete, and unobstructed for proper shipping. Any marks or labels which are defaced, altered, or covered up in any way are invalid.

Secondary Packaging

- Address of the sender
  (with emergency contact information)

- Biohazard Symbol
  (not required if the symbols is present on the secondary container)

Outside Packaging

- Address of the sender
  (with emergency contact information)

- Address of the intended recipient
  a. Mark the intended laboratory

- Class 6.2 Hazard Diamond

- UN number and proper shipping name(s)
  - Must be within 6 inches of the 6.2 hazard diamond and on the same side of the box

<table>
<thead>
<tr>
<th>Category</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Category A</td>
<td>“UN 2814 Infectious Substance, Affecting Humans” (technical name)</td>
</tr>
<tr>
<td>Category B</td>
<td>“UN 3373 Biological Substance, Category B”</td>
</tr>
<tr>
<td>Dry Ice</td>
<td>“UN 1845 Dry Ice”</td>
</tr>
</tbody>
</table>
**Outside Packaging** (Situational)

- If Dry Ice was used,
  - A class 9 hazard diamond
  - Must be within 6 inches of the 6.2 hazard diamond and on the same side of the box
  - Mark the weight of dry ice, in kilograms. One pound = 2.2 kg

- “Overpack” (if an overpack was used)

- Orientation Arrows (if the specimen is liquid)

**Emergency Contact Information**

- The outside packaging and the secondary container must be marked with an emergency contact name and telephone number for a point of contact of the sender.
- This person must be knowledgeable about the contents of the shipment and be able to provide guidance to first responders who call in case of a spill.
- This number must be immediately answered by the knowledgeable person. An answering service or voicemail is not acceptable.
- An outside contractor that provides this type of service may be used if you have an agreement in place.
G. Special Situations

**Newborn Screening Blood Spots**

1. Allow blood spots to **AIR DRY** thoroughly on a level non-absorbent surface such as a plastic coated test tube rack at least 4 hours at room temperature.

2. Place **dried** filter paper form(s) into the provided mailing envelope. Mail the specimen within 24 hours. No additional labeling is required on the outside of the envelope. The dried blood spots cannot leak or spill and are exempt from the dangerous goods/hazardous materials shipping regulations. **The envelopes provided to ship dried blood spots should not be used to ship any other type of patient specimen.**

3. Overnight shipping is recommended. The Public Health Lab (PHL) has a FedEx account to cover the cost of shipping newborn screening specimens to the PHL. To enroll to use this FedEx account, contact PHL at 803-896-0795.

**Suspected Bioterrorism Specimens and Cultures**

Prior notification is requested for specimens and/or cultures being sent for “rule out/rule in” testing for bioterrorism agents. Please notify: The Special Pathogens Supervisor, **Amanda Moore, 803-896-0777** before shipping these specimens or cultures. Alternate: **Megan Davis, 803-896-0870**

Use only UN certified packaging. UN certified shippers specific to the special pathogens program are available upon request. See the section on Requesting Shipping Supplies.

Classification of the infectious substance is the shipper’s responsibility and should be based on the available information. We encourage shipping suspected bioterrorism samples as Category A infectious substances as an additional precaution.

If you are shipping a sample suspected, but not confirmed, to be a category A infectious substance, and have elected to ship the material as a category A as an additional safety measure, the packing name must use the text “Infectious Substance, Affecting Humans (suspected category A infectious substance).”

To ensure that the sample is routed to the correct laboratory, **Please verify that the Special Pathogens Laboratory has been marked** on the “To” shipping label.

**[NOTE]** Special Pathogens pre-labeled shippers may be obtained by calling 803-896-0777 / 803-896-0773 (limit 2 per laboratory).
H. Contact Information and Support

Public Health Laboratory Shipping Address

<table>
<thead>
<tr>
<th>Public Health Laboratory</th>
<th>Business hours are 8:00 AM to 4:00 PM</th>
</tr>
</thead>
<tbody>
<tr>
<td>8231 Parklane Road</td>
<td>Monday through Friday, except</td>
</tr>
<tr>
<td>Columbia, SC 29223</td>
<td>for state holidays</td>
</tr>
</tbody>
</table>

Public Health Laboratory Contact Information

24/7 telephone number: 803-896-0800

Safety Office: 803-896-0956

Requesting Shipping Supplies: Email: PHL-Supply@dhec.sc.gov

Requesting Shipping and Specimen Collection Supplies

Shipping supplies are available without charge to support DHEC programs.
Supplies include:

- Shippers
- Mark and Label Stickers (hazard diamonds, UN numbers, etc.)
- Biohazard bags
- Absorbent materials
- Requisition forms

To request materials, please contact by email at PHL-Supply@dhec.sc.gov.

References for Information in This Document:

IATA Dangerous Goods Regulations, 61st edition, effective January 1, 2020 to December 31, 2020


United States Postal Service, Domestic Mail Manual

Code of Federal Regulations, 42 CFR Part 73, (Select Agent Regulations)

Centers for Disease Control and Prevention, Guidelines for the Shipment of Dried Blood Spot Specimen
SECTION V

TEST FEE POLICY
The Public Health Laboratory is only partially supported by legislative appropriations from State Funds. Therefore, we have been authorized to charge fees under certain conditions.

TEST FEES:
A fee is charged for those tests which benefit only the individual patient or which are readily available from private sources.

Exempt from charges:
A. Test (s) that is not reasonably available from qualified private laboratories.
B. Test (s) whose result is primarily of epidemiologic or public health significance.
C. Test (s) performed as a matter of lab policy which is not requested by the physician.
D. When the patient is medically indigent. In this case, the physician will be billed, but may deduct the charges before remitting. See billing procedures.
E. Repeat Tests for Newborn Screening. If the Repeat Test was requested by the Public Health Laboratory, i.e., Initial Test was invalid due to early dismissal, or improperly collected specimen or insufficient quantity or other reason, there is no charge for the Repeat Test. **All initial and second tests are subject to the full fee.**

BILLING PROCEDURE
Clients/Customers will be billed monthly by an itemized invoice that includes the patient’s name, medical record number, specimen number, date mailed, test(s) performed, and the test fees for each specimen. Billing invoices are generated by Sender and/or Billing numbers. Please note that the Public Health Laboratory does not bill Medicaid or any private insurance companies.

Payments:
1. Do not send payment with the specimen. Pay only when you receive a billing invoice. Note: Please do not send cash payments.

2. The billing invoice will consist of two copies: The remittance copy must be returned with your payment for proper crediting of your account. Please retain the provider copy for your records. On the left side of the billing invoice there is a column headed “Eligible for NON payment.” Please place an “X” in this column beside the name of any patient listed who is considered to be unable to pay for the test, i.e. indigent. Place the total charges for patients eligible for non-payment in the indicated space at the upper right-hand corner of the billing invoice and deduct this amount from the total charges. Please indicate the amount remitted on the line designated on the billing invoice. Please make check payable to South Carolina Department of Health and Environmental Control (SCDHEC) and remit to the Attention of: Bureau of Financial Management, PO Box 100103, Columbia, South Carolina 29202-3103. If you have any questions pertaining to your account, please notify the Public Health Laboratory immediately at (803) 896-0800.

3. Payment can be accessed on DHEC website at http://www.scdhec.gov. Click on “For Business” then click on “Pay Invoices”. **Note: Total payment amount online for debit/credit card payment is limited $3,000.00 with a $1.00 transaction fee. Total payment amount greater than $3,000.00 can be paid online by electronic check.**